

DOI: http://dx.doi.org/10.21276/ap.2021.10.2.66

Annals of Phytomedicine: An International Journal http://www.ukaazpublications.com/publications/index.php

Print ISSN: 2278-9839

Online ISSN : 2393-9885



Effect of botanicals on collar rot of chickpea caused by *Sclerotium rolfsii* Sacc. in combination with *Trichoderma harzianum*

Suraj Kumar Patel, Siddarth N. Rahul⁺ and Sushil Kumar Singh

Department of Plant Pathology, Acharya Narendra Deva University of Agriculture and Technology, Kumarganj, Ayodhya-224229, Uttar Pradesh India

Article Info	Abstract
Article history	In this study, the effect of different botanicals in combination with Trichoderma harzianum were used to
Received 9 October 2021	observe plants mortality and yield affected by collar rot of chickpea. The effect of different treatments
Revised 29 November 2021	on germination at 20 days after sowing and vigour index 1 (germination × plant height), vigour index 2
Accepted 30 November 2021	(germination \times dry weight), 45, 60, 75 days after sowing (DAS) and also yield parameter like (yield per
Published Online 30 December 2021	plot and yield in q/h) were evaluated at after harvesting of chickpea. Significant variant in germination of
	chickpea seeds were recorded under different treatments. The lowest germination was recorded in the
Keywords	control plot (T9) (85.89%) and maximum in (T8) with (97.33%) at 20 days after sowing. Highest vigour
Botanicals	index 1 (2019.59) and vigour index 2 (147.94) were recorded in T8, maximum plant height in T8 (20.75,
Trichoderma harzianum	25.25, 33.38 cm.), highest total number of branches per plant in T8 (5.47, 6.27, 7.67), maximum fresh
Sclerotium rolfsii Sacc.	weight in T8 (6.42, 17.50, 20.72 gm.), maximum dry weight in T8 (1.52, 4.26, 5.09), minimum plant
collar rot	mortality in T8 (3.55, 7.47, 14%) and maximum plant mortality in control (9.73, 15.07, 21%) were
	recorded at 45, 60, 75 DAS, and maximum yield per plot in T8 (1.68 kg) and also the yield parameter and
	plants mortality was best observed in T& followed by in botanical T2 (Neem oil 2.5%) with combination

of Trichoderma harzianum, respectively.

1. Introduction

Chickpea (*Cicer arietinum* L.) is, a self-pollinated, diploid (2n = 2x)= 16) annual legume of family Fabaceae and known as various names, viz., Gram, Bengal gram, Garbanzo and Egyption pea. It was first cultivated in south eastern region of the world, but now it is also cultivated in semi-arid regions (Agarwal et al., 2012). Pulses contain higher proportions of protein (17%-30% by dry weight) in comparison to other plant foods. With the protein, chickpea is a good source of carbohydrates has comparison to other pulses, according to Wallace et al. (2016). There are various biotic and abiotic factors that affect the production of the chickpea in the world as well as in India too. The biotic factor includes insect pests and a number of devastating diseases caused by fungi, bacteria, viruses, and nematodes. The fungal diseases such as Fusarium wilt (Fusarium oxysporium f.sp. ciceris), Ascochyta blight (Ascochyta rabiei), collar rot (Sclerotium rolfsii), Verticillium wilt (Verticillium dahliae), black root rot (Fusarium solani), Phytophthora root rot (Phytophthora megasparma) and seed rot (Aspergillus flavus), etc. Among all of the diseases collar rot disease, caused by Sclerotium rolfsii Sacc., is a serious threat to chickpea that may cause 55-95% mortality of the crop at seedling stage under favourable environmental conditions (Gurha and Dubey, 1982).

Corresponding author: Dr. Siddarth N. Rahul

Department of Plant Pathology, Acharya Narendra Deva University of Agriculture and Technology, Kumarganj, Ayodhya-224229, Uttar Pradesh India E-mail: sagar4499@gmail.com

Tel.: +91-8795114474

Copyright © 2021 Ukaaz Publications. All rights reserved. Email: ukaaz@yahoo.com; Website: www.ukaazpublications.com

2. Materials and Methods

2.1 Preparation of botanicals

Fresh parts of the test plants (*Lantana camera*, *Eucalyptus* spp. *Ocimum sanctum* and *Azadirachta indica*) were collected and washed thoroughly in clean water. Hundred grams of each washed samples were grinded in mortar and pestle by adding equal amount (100 ml) of sterilized distilled water (1:1 W/V) and boiled at 80°C for 10 min in a hot water bath. The grinded material was filtered through muslin cloth followed by filtering through sterilized Whatman No. 1 filter paper and treated as standard 100 per cent plant extract (Nene and Thapliyal, 1982) and required concentrations of five per cent of each plant extract were prepared.

2.2 Treatments details

All the treatments were used as seed treatments. (T1) Seeds treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and *Pseudomonas* spp. (10 gm/kg seed). (T2) Seeds treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and Neem oil (2.5%). (T3) Seed treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and Jeevamrit (5%). (T4) Seed treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and *Lentana camera* extract (5%). (T5) Seed treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and *Lentana camera* extract (5%). (T5) Seed treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and *Eucalyptus* spp. extract (5%). (T6) Seed treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and *Ocimum sanctum* extract (5%). (T7) Seed treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and Neem leaf extract (5%). (T8) Seed treated with combination of *Trichoderma harzianum* (10 gm/kg seed) and Carbendazim (3 gm/kg seed).

3. Results

Results presented in (Table 1), indicated that the treatment tested against *S. rolfsii* has a positive effect in germination percentage as T8 give maximum germination percentage *i.e.*, (97.33%), maximum vigour index 1 and vigour index 2 in T8, *i.e.* (2019.57) (147.94) respectively, Observations presented in (Table 2), the data was recorded at different days after sowing, *i.e.* 45, 60, 75 (DAS). Maximum plant height was

observed in T8, *i.e.* (20.75 cm) (25.25 cm) (33.80 cm), respectively, Total No. of branches per plant highest in T8, *i.e.* (5.47) (6.27) (7.67), respectively; highest total no. of pod/plant and seed index were recorded in T8, *i.e.* (53) (21.67 gm), respectively (Table 3). In Table 4, the highest fresh weight was recorded in T8 (6.42 gm) (17.50 gm) (20.72 gm) and maximum dry weight in T8 (1.52 gm)(4.26 gm) (5.09 gm).

Table 1: Effect of different treatments on plant mortality and vigour index I and II of chickpe

Treatment	Per cent germination (%)	Vigour index 1 (germination × plant length)	Vigour index 2 (germination × - dry weight)	Per cent mortality		У
				45 DAS	60 DAS	75 DAS
T1	91.11	1589.86	104.77	7.03 ± 0.15	11.67 ± 0.83	17.67 ± 0.76
T2	95.11	1882.22	133.15	4.44 ± 0.39	8.17 ± 0.76	15.33 ± 2.08
Т3	92.66	1652.12	109.33	5.90 ± 0.62	10.87 ± 0.81	17.27 ± 0.64
T4	89.77	1526.98	96.95	7.00 ± 0.50	12.50 ± 0.50	17.47 ± 1.31
Т5	88.22	1458.27	76.75	7.80 ± 0.48	13.17 ± 1.26	17.50 ± 0.70
T6	93.88	1722.69	120.16	5.77 ± 0.32	10.20 ± 0.36	16.20 ± 1.06
T7	94.33	1780.95	123.57	5.30 ± 0.36	8.97 ± 0.45	15.50 ± 1.32
T8	97.33	2019.59	147.94	3.55 ± 0.30	7.47 ± 0.55	14.00 ± 1.00
Т9	85.89	1313.25	67.85	9.73 ± 0.31	15.07 ± 1.01	21.00 ± 1.00
SE(m)	0.55	33.36	0.11	0.23	0.43	0.59
CD	1.61	96.97	0.32	0.66	1.24	1.73

(T1) Trichoderma harzianum (10 gm/kg seed) and Pseudomonas spp. (10 gm/kg seed). (T2) Trichoderma harzianum (10 gm/kg seed) and Neem oil (2.5%). (T3) Trichoderma harzianum (10 gm/kg seed) and Jeevamrit (5%). (T4) Trichoderma harzianum (10 gm/kg seed) and Lentana camera extract (5%). (T5) Trichoderma harzianum (10 gm/kg seed) and Eucalyptus spp. Extract (5%). (T6) Trichoderma harzianum (10 gm/kg seed) and Ocimum sanctum extract (5%). (T7) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Carbendazim (3 gm/kg seed), DAS-Days after showing, SE-Stander Error, CD-Critical Difference.

Treatment	Plant height (in cm.)			Total no of branches/plant		
	45 DAS	60 DAS	75 DAS	45 DAS	60 DAS	75 DAS
T1	17.45 ± 1.36	22.62 ± 1.46	28.93 ± 1.29	4.27 ± 0.42	5.20 ± 0.35	5.33 ± 0.23
T2	19.79 ± 1.39	24.87 ± 0.98	32.07 ± 1.14	$5.07~\pm~0.90$	5.93 ± 0.42	6.33 ± 0.23
Т3	17.83 ± 1.67	23.60 ± 1.34	29.47 ± 1.29	$4.33~\pm~0.50$	5.40 ± 0.00	5.47 ± 0.12
T4	17.01 ± 1.23	22.59 ± 0.95	28.47 ± 1.03	4.20 ± 0.53	5.00 ± 0.35	5.27 ± 0.12
Т5	16.53 ± 1.29	22.05 ± 1.50	27.93 ± 1.01	4.13 ± 0.46	4.60 ± 0.40	4.87 ± 0.23
T6	18.35 ± 1.68	23.79 ± 1.51	30.47 ± 0.61	$4.47~\pm~0.42$	5.60 ± 0.20	5.73 ± 0.31
T7	18.88 ± 1.52	24.17 ± 1.34	31.87 ± 1.10	$4.73~\pm~0.70$	5.67 ± 0.31	$6.00~\pm~0.00$
Т8	20.75 ± 0.66	25.25 ± 1.10	33.80 ± 0.60	$5.47~\pm~0.64$	6.27 ± 0.31	$7.67~\pm~0.58$
Т9	15.29 ± 0.71	20.94 ± 0.52	26.47 ± 1.47	$3.40~\pm~0.53$	4.47 ± 0.50	4.60 ± 0.20
SE(m)	0.29	0.33	0.30	0.15	0.13	0.16
CD	0.84	0.96	0.87	0.43	0.38	0.48

Table 2: Effect of different treatments on plant height and number of branches/plant

(T1) Trichoderma harzianum (10 gm/kg seed) and Pseudomonas spp. (10 gm/kg seed). (T2) Trichoderma harzianum (10 gm/kg seed) and Neem oil (2.5%). (T3) Trichoderma harzianum (10 gm/kg seed) and Jeevamrit (5%). (T4) Trichoderma harzianum (10 gm/kg seed) and Lentana camera extract (5%). (T5) Trichoderma harzianum (10 gm/kg seed) and Eucalyptus spp. extract (5%). (T6) Trichoderma harzianum (10 gm/kg seed) and Ocimum sanctum extract (5%). (T7) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (3 gm/kg seed), DAS-Days after showing, SE-Stander Error, CD-Critical Difference.

Treatment	Total number of	Seed index (weight	Yield		
	pods/plant	/100 Seeds) in gm.	per plot (in kg)	Q./ha	
T1	36.67 ± 0.58	17.50 ± 1.50	1.51 ± 0.08	17.26 ± 0.73	
T2	50.67 ± 5.86	20.67 ± 1.53	1.66 ± 0.15	18.70 ± 0.61	
Т3	41.33 ± 4.51	18.67 ± 1.53	1.56 ± 0.05	17.59 ± 0.73	
T4	33.33 ± 0.58	17.43 ± 1.44	1.49 ± 0.02	16.85 ± 0.85	
Т5	31.67 ± 2.31	17.33 ± 2.08	1.48 ± 0.03	16.33 ± 0.40	
T6	43.00 ± 3.00	19.17 ± 1.53	1.58 ± 0.12	17.88 ± 0.73	
T7	46.33 ± 6.11	19.67 ± 1.53	1.65 ± 0.11	18.36 ± 0.50	
Т8	53.00 ± 7.00	21.67 ± 1.53	1.68 ± 0.08	19.29 ± 1.22	
Т9	28.33 ± 1.53	15.30 ± 0.61	1.25 ± 0.04	15.29 ± 0.84	
SE(m)	1.92	0.32	0.04	0.22	
CD	5.58	0.94	0.12	0.63	

Table 3: Effect of different treatments on total number of pods/plant, seed index and yield

(T1) Trichoderma harzianum (10 gm/kg seed) and Pseudomonas spp. (10 gm/kg seed). (T2) Trichoderma harzianum (10 gm/kg seed) and Neem oil (2.5%). (T3) Trichoderma harzianum (10 gm/kg seed) and Jeevamrit (5%). (T4) Trichoderma harzianum (10 gm/kg seed) and Lentana camera extract (5%). (T5) Trichoderma harzianum (10 gm/kg seed) and Eucalyptus spp. extract (5%). (T6) Trichoderma harzianum (10 gm/kg seed) and Ocimum sanctum extract (5%). (T7) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Carbendazim (3 gm/kg seed), DAS-Days after showing, SE-Stander Error, CD-Critical Difference.

Treatment	Fresh weight (in gm.)			Dry weight (in gm.)		
	45 DAS	60 DAS	75 DAS	45 DAS	60 DAS	75 DAS
T1	4.80 ± 0.56	13.92 ± 0.48	14.30 ± 1.41	1.15 ± 0.17	3.49 ± 0.13	3.76 ± 0.35
T2	6.02 ± 0.93	15.65 ± 0.08	18.13 ± 1.14	1.40 ± 0.17	3.85 ± 0.13	4.59 ± 0.22
Т3	5.12 ± 0.55	14.30 ± 0.72	15.69 ± 0.48	1.18 ± 0.19	3.51 ± 0.13	3.93 ± 0.18
T4	4.53 ± 0.55	13.44 ± 0.38	13.92 ± 1.69	1.08 ± 0.09	3.40 ± 0.12	3.61 ± 0.41
Т5	3.76 ± 0.43	12.53 ± 0.76	13.30 ± 1.97	0.87 ± 0.05	3.27 ± 0.08	3.40 ± 0.41
Т6	5.48 ± 0.56	14.83 ± 0.54	16.14 ± 0.50	1.28 ± 0.16	3.59 ± 0.07	4.17 ± 0.11
T7	5.69 ± 0.58	14.99 ± 0.52	16.78 ± 0.38	1.31 ± 0.17	3.76 ± 0.12	4.26 ± 0.03
T8	6.42 ± 0.89	17.50 ± 1.44	20.72 ± 2.61	1.52 ± 0.15	4.26 ± 0.32	5.09 ± 0.51
Т9	3.29 ± 0.26	10.59 ± 0.32	10.53 ± 2.95	0.79 ± 0.05	2.79 ± 0.05	2.61 ± 0.63
SE(m)	0.26	0.40	0.94	0.05	0.09	0.20
CD	0.77	1.16	2.73	0.14	0.26	0.58

Table 4: Effect of different treatments on Fresh and dry weight of plant

(T1) Trichoderma harzianum (10 gm/kg seed) and Pseudomonas spp. (10 gm/kg seed). (T2) Trichoderma harzianum (10 gm/kg seed) and Neem oil (2.5%). (T3) Trichoderma harzianum (10 gm/kg seed) and Jeevamrit (5%). (T4) Trichoderma harzianum (10 gm/kg seed) and Lentana camera extract (5%). (T5) Trichoderma harzianum (10 gm/kg seed) and Eucalyptus spp. extract (5%). (T6) Trichoderma harzianum (10 gm/kg seed) and Ocimum sanctum extract (5%). (T7) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T9) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/kg seed) and Neem leaf extract (5%). (T8) Trichoderma harzianum (10 gm/

As shown in Table 2, chickpea yield was recorded in per plot (in kg), significantly varied from one treatment to another. The yield of chickpea per plot ranges from 1.32 kg to 1.68 kg. Table 4 and figure showed the highest grain yield (kg/plot) (1.68 kg) in (T8) and (1.65 kg/plot) in T2 were recorded best effective on grain yield over the control, *i.e.* T9 (1.32 kg), followed by (1.65 kg), T7 (1.61 kg), T6, (1.57 kg), T3 (1.51 kg) and (1.48 kg) in T5. Likewise yield q/h was recorded significantly varied from one treatment to another. Table 4 and Figure. showed the highest grain yield (q/h) (19.29 q), (T8) and (18.70 q) in T2 were recorded best effective on grain yield over the

control, *i.e.*, T9 (15.29 q), followed by (18.36 q), T7 (17.88 q), T6 (17.59 q), T3 (17.26 q), T1 (16.85 q) and T4 (16.33 q) in T5, respectively.

The mortality presented in Table 2 showed that at 45, 60 and 75 days after sowing (DAS) (Table 6 and Figure). At 45 days after sowing, maximum mortality (9.73%) was observed in control plot (T9), which was statistically significant compared to other treatments. Minimum mortality (3.55%) was recorded in the plot where seeds were treated with T8, followed by T2 (4.44%), T7

(5.30%), T6 (7.80%). At 60 days after sowing, the Table 2 showed maximum mortality (15.07%) was recorded in control plot T9, where minimum mortality (7.47%) was found in the plot T8, followed by T2, T7, T6, T3 over the control as many worker like Pawar *et al.*, (2014) Das *et al.* (2014) and Khan and Javaid (2015) also revealed that the chemicals have the significant effect on mortality and disease inhibition of chickpea. As shown in Table 1 at 75 days after sowing maximum mortality (21%) was recorded in control plot T9. where minimum mortality (14%) was found in the plot T8, followed by T2, T7, T6, T3,T1, T4 and T5.

4. Discussion

Many researchers like Khan *et al.* (2020) and Tewari and Mukhopadhyay (2000) used botanicals and bioagents as seed treatments increased seedling emergence. More *et al.*, (2016) revealed that maximum seedling vigour index was recorded in Carbendazim followed by *Trichoderma viride* and *Azadirachta indica* in the requirement which support and shows the similarity observations are recorded in present investigation. Singh *et al.* (2018) revealed that the number of healthy pod per plant and seed index is similar as the present investigation. Tewari and Mukhopadhyay (2000) used botanicals as seed treatments and found affective in increases seedling emergence and grain yield. Many worker like Dutta *et al.* (1991) and the findings of Asgharian and Mayee (1991), Jhonson *et al.* (2008) was also similar as grain yield was increased while using of botanicals and bioagents. Kumar *et al.* (2008) also used bioagent and revealed that higher yield of chickpea.

Nagamma and Nagaraja (2015) revealed that the maximum inhibition of mycelial growth (71.67%) was noticed in *Trichoderma harzianum* which was followed by *Trichoderma viride* (63.33%) under *in vitro* condition. Bhuiyan *et al.* (2012) also revealed that the bioagents have the significant effect on *Sclerotium rolfsii* and maximum inhibition was recorded and many worker like More *et al.* (2016), Khan *et al.* (2020) also revealed that the botanicals have the significant effect on mortality of chickpea plant. Prabhu (2003) revealed that the different systemic and non-systemic fungicides against *Sclerotium rolfsii* and reported 100% inhibition by carboxin, which was followed by carbendazim (63%) + mancozeb (12%) and propiconazole.

5. Conclusion

In the current investigation, it is concluded that application of T8 [*Trichoderma herzianum* (10 g/kg seed) + Carbendazim (3 g/kg seed)] showed better performance in enhancing germination and reducing mortality percentage and appearance of the disease. This is also true that use of *Trichoderma herzianum* (10 g/kg seed) + Carbendazim (3 g/kg seed) promoted plant height, total number of branches/plant, total number of pod/plant reduced disease incidence and showed maximum grain yield of chickpea. In the botanicals, T2 [*Trichoderma harzianum* (10 g/kg seed) + Neem oil (2.5%)] was found to be the best in all prospects. In the remaining treatments, all were effectively controlling the disease and enhance the plant growth and production. For batter environment and to avoid hazardous effect of chemicals, the botanicals can be the alternative option to manage such kind of plant diseases.

Conflict of interest

The authors declare no conflicts of interest relevant to this article.

References

- Agarwal, G.; Jhanwar, S.; Priya, P.; Singh, V.K. and Jain, M. (2012). Comparative analysis of kabuli chickpea transcriptome with desi and wild chickpea provides a rich resource for development of functional markers. Jour. of Plt. Path., 7:441-443.
- Asghari, M.A and Mayee, C.D. (1991). Comparative efficacy of management practices on stem and pod rots of groundnut. In. Phytopath., 44: 328-332.
- Ayock, R. (1966). Stem rot and other disease caused by *Sclerotium rolfsii*. North Carolina Agricultural Experiment Station. Tech. Bullt., 174-202.
- Bhuiyan, M.A.H.B.; Rahman, M.T. and Bhuiyan, K.A. (2012). In vitro screening of fungicides and antagonists against *Sclerotium rolfsii*, Afr. Jour. of Biotech.11(82):14822-14827
- Das, N.C.; Dutta, B.K. and Ray, D.C. (2014). Potential of some fungicides on the growth and dovelopement of *Sclerotium rolfsii* Sacc. *In vitro*. Inter. Jour. of Scie. and Rese. Publ., 4(12):ISSN 2250-3153.
- Gupta, S.K. and Sharma, A. (2004). Symptomology and management of crown rot of (*Sclerotium rolfsii*) of French bean. Jour. of Myco. and Pl. Path., 34:820-823.
- Gurha, S.N. and R.S. Dubey (1982). Occurrence of possible sources of resistance in chickpea (*Cicer arietinum L.*) against Sclerotium rolfsii Sacc. Madras Agri. Jour., 70:63-64
- Harinath, N. (2000). Crossandra-A new host record for *Sclerotium rolfsii*. Ind. Phyto., 53:496-497.
- Hind, T.S. (2005). Disease of field crop and their management, Daya Publishing House, New Delhi, 171 pp:171.
- Jhonson, M.; Reddy, P.N and Reddy, D.R. (2008). Comparative efficacy of rhizosphere mycoflora, fungicides, insecticides and herbicides against groundnut stem rot caused by *Sclerotium rolfsii*. Ann. of Pl.Prote. Sci., 16(2):414-418.
- Khan, I.H.and Javaid, A. (2015). Chemical control of collar rot disease of chickpea. Pak., Jour. of Phytopath. 27:(01)61-68.
- More, P.S.; Parate, R.L. and Mairan, N.R. (2016). Evaluation of botanicals and bioagents to record the root, shoot length and vigour index of chickpea. Inter. Jour. of Pl. Prot., 9(2):483-488.
- Nagamma, G. and Nagaraja, A. (2015). Efficacy of biocontrol agents against Sclerotium rolfsii causing collar rot disease of chickpea, under in vitro conditions. Inter. Jour. of Pl. Prot., 8(2):222-227.
- Narasimha, R.S.; Anahosur, K.H. and Srikant, K. (2004) Eco-friendly approaches for management of wilt of potato (*Sclerotium rolfsii*). Jour. of Myco. and Pl. Path., 34:327-329.
- Nene, Y.L., Reddy; M.V., Haware; M.P., Ghanekar; A.M., Amin; K.S. Pande and Sharma, M. (2012) Field diagnosis of chickpea disease and their control. Information Bulletin No. 28 (Revised). Inter. Crop Rese. Inst. for the Semi-Arid Trop. Patancheru, A.P. 502 324, India, 2627 pp:2627.
- Nyvall, R.F.(1989) Field Crop Disease Handbook, Second edition, Published by Van Nostrand Reinhold, New York, 817 pp:817.
- Pawar, A.K.; Surywanshi, A.P.; Gawade, D.B.; Zagade, S.N and Wadje, A.G (2014). Effects of organic amendments and fungicides on the survival of collar rot fungus of soybean incited by *Sclerotium rolfsi*. Afr. Jour. of Agri. Res., 9(27):2124-2131.

506

- Prabhu, H.V. (2003). Studies on collar rot of soybean caused Sclerotium rolfsii Sacc. M.Sc. (Agri.) Thesis, University of Agricultural Sciences. Dharwad.
- Rajalakshmi (2002) Studies on variability among the isolates of *Sclerotium rolfsii*. M. Sc.(Ag) Thesis, Acharya N. G. Ranga Agricultural University, Hyderabad, Andhra Pradesh.
- Rakholiya, K.B. and Jadeja, K.B. (2011). Morphological diversity of *Sclerotium rolfsii* caused and pod rot of groundnut. Jour. of Myco. and Pl. Path., 41(4):500-504.
- Ramesh, A.; Gupta, O. and Mishra M. (2014) Technique for screening of chickpea genotype against collar rot, its management through host plant rasistance and fungicides. Leg. Res., 37(1):110-114.
- Reddi, M.K.; Santhoshi, M.V.M.; Krishna, T.G. and Reddy, K.R. (2014) Cultural and morphological variability *Sclerotium rolfsii* isolates infecting ground nut and its reaction to some fungicides. Inter. Jou. of Currt. Microb. and App. Sci., 3(10):553-561.
- Shivani, B.R.; Dubey, R.C. and Maheshwari, D.K. (2005) Enhancement of plant growth and suppression of collar rot of sunflower caused by *Sclerotium rolfsii* through *Pseudomonas fluorescens*. In: Phytopath., 58(1):17-42.
- Tewari, A.K and Mukhopadhyay, A.N. (2000). Management of chickpea root rot and collar rot by integration of biological and chemical seed treatment. In: Phytopath., 56(1):39-42.
- Wallace, T.C.; Murray, R. and Zelman, K.M. (2016). The nutritional value and health benefits of chickpeas and hummus. Nutrients, 8(12):E766.

Suraj Kumar Patel, Siddarth N. Rahul and Sushil Kumar Singh (2021). Effect of botanicals on collar rot of chickpea caused by *Sclerotium rolfsii* Sacc in combination with *Trichoderma harzianum*. Ann. Phytomed., 10(2):502-506. http://dx.doi.org/10.21276/ap.2021.10.2.66