DOI: http://dx.doi.org/10.54085/ap.2024.13.2.84

Annals of Phytomedicine: An International Journal http://www.ukaazpublications.com/publications/index.php

Print ISSN: 2278-9839

**Online ISSN : 2393-9885** 

## **Original Article : Open Access**

# Exploring the phytochemical constituents of *Plumeria obtusa* L. root and stem extracts through GC-MS analysis

### J. Jaahnavi, S.P. Thamaraiselvi<sup>4</sup>, M. Ganga, N. Sritharan<sup>\*</sup> and P. Malathi<sup>\*\*</sup>

Department of Floriculture and Landscaping, Horticultural College and Research Institute, Tamil Nadu Agricultural University, Coimbatore-641003, Tamil Nadu, India

\* Department of Rice, Tamil Nadu Agricultural University, Coimbatore-641003, Tamil Nadu, India

\*\* Department of Soil Science and Agricultural Chemistry, Tamil Nadu Agricultural University, Coimbatore-641003, Tamil Nadu, India

Article Info	Abstract
Article history	This study investigates the bioactive phytochemicals present in the root and stem extracts of Plumeria
Received 10 August 2024	obtusa L., commonly known as the 'Great white frangipani' using gas chromatography-mass spectrometry
Revised 29 September 2024	(GC-MS). Traditionally used in various medicinal applications, P. obtusa is valued for its therapeutic
Accepted 30 September 2024	properties. The GC-MS analysis identified a wide array of bioactive compounds, including ethyl alpha-d-
Published Online 30 December 2024	glucopyranoside, 2,3-dihydrobenzofuran, n-decanoic acid and methyl beta-d-glucopyranoside. These
	compounds exhibit significant pharmacological activities such as antioxidant, anticancer, antimicrobial
Keywords	and anti-inflammatory effects. Glycosides like ethyl alpha-d-glucopyranoside and methyl beta-d-
Plumeria obtusa L.	glucopyranoside showed strong antioxidant and skin-moisturizing properties, indicating their potential
GC-MS	for pharmaceutical and cosmetic applications. Fatty acids such as n-decanoic acid and methyl palmitate
Root extract	displayed anti-inflammatory and antimicrobial properties, making them suitable for treating infections
Stem extract	and inflammation related conditions. Additionally, plumericin, a terpenoid, demonstrated potent
Phytochemical compounds	antibacterial, antifungal and antiviral properties, while 9,12-octadecadienoic acid methyl ester showed
	promise as an analgesic and anticancer agent. The study highlights the potential of P. obtusa extracts for
	applications in pharmaceuticals, cosmetics and agriculture. The findings underscore the need for further
	research to explore the therapeutic potential of these bioactive compounds and to develop new applications
	in various industries.

# 1. Introduction

Plants offer everything essential for human survival, including food, fiber, shelter and especially natural remedies to treat a wide range of ailments (Sharma *et al.*, 2021). Plants play a significant role in ethnopharmacology and are widely used by numerous cultures across the globe to treat various health conditions (Sharma and Alam, 2022). Bioactive compounds from plants are present in all parts of the plant, including the bark, leaves, flowers, roots, fruits and seeds. Plant extracts and their active constituents are widely recognized for demonstrating significant biological activities, particularly antimicrobial and antioxidant properties (Musini *et al.*, 2013).

Gas chromatography-mass spectrometry (GC-MS) is a key tool for precise and detailed chemical evaluation of phytochemicals in plant extracts. It helps to identify bioactive constituents such as longchain hydrocarbons, alcohols, acids, esters, alkaloids, steroids, amino compounds and nitrocompounds (Muthulakshmi and Mohan, 2012). Volatile compounds, such as terpenoids, alkaloids and flavonoids are identified and assessed through their mass spectra (Singh *et al.*, 2024). The demand for herbal products has grown worldwide, with more

Corresponding author: Dr. S.P. Thamaraiselvi Associate Professor, Department of Floriculture and Landscaping, Horticultural College and Research Institute, Tamil Nadu Agricultural University, Coimbatore-641003, Tamil Nadu, India E-mail: thamaraiselvi.sp@tnau.ac.in Tel.: +91-9843338666

Copyright © 2024Ukaaz Publications. All rights reserved. Email: ukaaz@yahoo.com; Website: www.ukaazpublications.com people in both developed and developing countries using plantbased health solutions. Plumeria obtusa L., commonly known as Great white frangipani, is an evergreen tree, grown as an ornamental plant for its attractive and fragrant flowers. On commercial scale, essential oil extracted from its flowers is used in perfume industry (Choudhary et al., 2014; Sulaiman et al., 2008). Various parts of the plant have traditionally been used to treat a range of conditions, including diabetes mellitus, wounds, skin diseases, as well as for their diuretic, purgative and abortifacient properties. Additionally, these plant parts are utilized in cosmetics, aromatherapy, as ornaments and for ceremonial offerings (Bihani et al., 2021). The plant exhibits a broad spectrum of pharmacological activities, including antimicrobial, antiproliferative, antimutagenic, anti-inflammatory, antiulcerogenic, algicidal, antioxidant, insecticidal and wound-healing properties. Therefore, isolating phytoconstituents from plants and assessing their key pharmacological properties is crucial in the future development of new drugs (Srivani and Mohan, 2023).

Hence, the study aims to thoroughly investigate the root and stem extracts of *P. obtusa* to identify the biochemical compounds responsible for its extensive pharmaceutical properties, which hold significant potential for future applications.

### 2. Materials and Methods

#### 2.1 Collection of plant material

Fresh root and stem parts of *P. obtusa* cuttings were collected from the *Plumeria* germ plasm being maintained by the Department of Floriculture and Landscaping, Tamil Nadu Agricultural University,





Coimbatore district, Tamil Nadu, India for the present investigation. The plant authentication number for *Plumeria obtusa* L. was obtained from Botanical Survey of India, Southern Regional Centre, Coimbatore with Certificate No. BSI/SRC/5/23/2024/Tech./412 dt. 23.09.2024.

Figure 1: GC-MS elucidation of bioactive compounds in methanolic root extract of *P. obtusa*.

#### 2.2 Preparation of methanolic root and stem extracts of P. obtusa

The root and stem pieces of *P. obtusa* were collected and washed twice with de-ionized water and then allowed to dry naturally in a cool environment. After thorough drying, the root and stem pieces were pulverized in a grinder and then stored in an airtight container for further use (Guha *et al.*, 2010; Sultana *et al.*, 2009). The active constituents were extracted by maceration using 70% methanol and water. 50 g of the macerated plant material was weighed in to 500 ml conical flask and 300 ml of methanol was added and left to stand for three days while stirring at intervals. The extracts were then filtered and dried with a rotary evaporator (Brinkmann, R110). The concentrates were dispensed into labelled sample vials and stored in refrigerator at 4°C.

# 2.3 GC-MS analysis of root and stem extracts of P. obtusa

The volatile components of the methanolic extract of P. obtusa root and stem extracts were characterized using GC-MS, employing an Agilent Technologies model 7890A gas chromatograph equipped with a Mass Selective Detector model 5975C (MSD) operating under electron ionization (70 V) with ion source temperature set at 250°C. For analysis of this extract, a capillary column Agilent DB5MS (30 mm  $\times$  0.25 mm  $\times$  0.25 µm) was used. Helium (99.9%) was the highpurity carrier gas used, with a ml/min flow rate. The injector mode was split (1:60), with an injection volume of 1  $\mu$ l. The oven temperature program began at 100° and maintained for 0.5 min before increasing to 140° at 20°/min, keeping it there for a minute and then finally going up to 280° at 11°C/min over 20 min. Mass hunter software was used for peak area measurement and data processing. The identification of components was based on a comparison of their mass spectra with those contained in the NIST Wiley 2008 library

#### 2.4 Identification of bioactive compounds

Peaks were identified using databases namely; Wiley mass spectral library (W9N11) and the National Institute of Standards and Technology (NIST) through GC-MS for the identification of unknown volatile compounds. Information on the biological activities of these compounds was obtained from Dr. Duke's Phytochemical and Ethnobotanical Databases and the molecular weight and molecular formula were verified using PubChem.

# 3. Results

GC-MS is regarded as the "Gold standard" for substance identification because of its precise testing capabilities. This technology has been used to identify hundreds of components in natural and biological systems (Kabir and Furton, 2021). Numerous phytochemical compounds were observed in P. obtusa root extract by using this method. The present findings of GC-MS revealed the bioactive compounds in methanolic root extract of P. obtusa (Table 1, Figure 1). Analysis of peak area, retention time and molecular formulae confirmed these phytochemicals. The compounds were predicted using the NIST database. The major 20 compounds identified in root extract of P. obtusa (Table 1) are ethylalpha-d-glucopyranoside (11.39%), 2,3-dihydrobenzofuran(10.96%), n-decanoicacid (9.39%), methylbeta-d-glucopyranoside (8.78%), 2-(2-butoxyethoxy)ethyl thiocyanate(7.91%), 7-octadecenoic acid methyl ester(4.13%), methylbeta-d-galactopyranoside (3.75%),4'-methoxybiphenyl-3carboxylic acid (3.16%), linoleic acid ethyl ester (3.06%), 9,12octadecadienoic acid (Z,Z)-methyl ester (2.70%), hexadecanoic acid ethyl ester (2.68%), 2,2-diethoxytetrahydrofuran (2.65%), methylpalmitate (2.49%), ethyl oleate (2.40%), ethyl4ethoxybenzoate (1.93%), 2,6-dimethylbenzoquinone-4-oxime (1.80%), diborane(6) (1.77%), plumericin (1.48%), 7-hydroxy-6methoxy-2H-1-benzopyran-2-one (0.99%) and 6,10,14,18,22tetracosahexaene (0.85%).

Table 1	l: Phytochemical	compounds	identified in	ı the	methanolic	root	extract	of i	P. o	btusa
---------	------------------	-----------	---------------	-------	------------	------	---------	------	------	-------

S.No.	Chemical compound	Molecular formula	Molecular weight (g/mol)	Area %	Retention time
1.	Ethylalpha-d-glucopyranoside	$C_8 H_{16} O_6$	208.21	11.39	10.942
2.	2,3-Dihydrobenzofuran	C <sub>8</sub> H <sub>80</sub>	120.15	10.96	7.220
3.	n-Decanoic acid	$C_{10}H_{20}O_{2}$	172.64	9.39	11.041
4.	Methylbeta-d-glucopyranoside	$C_7 H_{14} O_6$	194.18	8.78	10.697
5.	2-(2-Butoxyethoxy)ethyl thiocyanate	$C_9H_{17}NO_2S$	203.30	7.91	9.431
6.	7-Octadecenoic acid, methyl ester	$C_{19}H_{36}O_{2}$	296.5	4.13	14.130
7.	Methylbeta-d-galactopyranoside	$C_{7}H_{14}O_{6}$	194.18	3.75	10.608
8.	4'-methoxybiphenyl-3-carboxylic acid	$C_{14}H_{12}O_3$	228.24	3.16	13.752
9.	Linoleic acid ethyl ester	$C_{20}H_{36}O_2$	308.5	3.06	14.508
10.	9,12-Octadecadienoic acid (Z,Z)- methyl ester	$C_{19}H_{34}O_2$	294.47	2.70	14.097
11.	Hexadecanoic acid ethyl ester	$C_{18}H_{36}O_2$	284.47	2.68	13.430
12.	2,2-Diethoxytetrahydrofuran	$C_8H_{16}O_3$	160.21	2.65	11.275
13.	Methyl palmitate	$C_{17}H_{34}O_2$	270.5	2.49	12.986
14.	Ethyl oleate	$C_{20}H_{38}O_2$	310.5	2.40	14.541
15.	Ethyl 4-ethoxybenzoate	$C_{11}H_{14}O_{3}$	194.23	1.93	10.053
16.	2,6-Dimethylbenzoquinone-4-oxime	C <sub>8</sub> H <sub>9</sub> NO <sub>2</sub>	151.16	1.80	9.642
17.	Diborane(6)	$B_2H_6$	33.71	1.77	4.876
18.	Plumericin	$C_{15}H_{14}O_{6}$	290.27	1.48	15.685
19.	7-Hydroxy-6-methoxy-2H-1-benzopyran-2-one	$C_{10}H_8O_3$	176.17	0.99	13.330
20.	2,6,10,14,18,22-Tetracosahexaene	C <sub>24</sub> H <sub>38</sub>	326.6	0.85	18.141

S.No.	Chemical compound	Nature of the compound	<b>Biological activity</b>	Reference
1.	Ethylalpha-d-glucopyranoside	Glycosides	Skin moisturizing agent, antituberculous, antioxidant, alpha amylase inhibitory, hypolipidemic, anticonvulsant	Sonia and Singh, 2019
2.	Methylbeta-d-glucopyranoside		Antimicrobial glycoside	Olawumi et al., 2020
3.	Methylbeta-d-galactopyranoside		Anticancer, CNS depressant, smart drug	Perumal et al., 2021
4.	n-Decanoic acid	Fatty acids	Anti-inflammatory agent, antimicrobial activity	Kitahara <i>et al.</i> , 2004
5.	Hexadecanoic acid ethyl ester		Antioxidant activity	Mohamad et al., 2020
6.	Methyl palmitate		Food and cosmetic additives cum anti-inflammatory, antioxidantactivity	Pinto et al., 2017
7.	7-Octadecenoic acid methyl ester		Antibacterialactivity	Shettima et al., 2013
8.	Ethyl oleate		Antibacterialactivity	Ankita et al., 2015
9.	Linoleic acid ethyl ester		Antiarthritic, antihistaminic and anti-inflammatory agents	Kolar et al., 2019
10.	9,12-Octadecadienoic acid (Z,Z)-methyl ester		Analgesic, anticancer, anti -inflammatoryactivity	Mohamad et al., 2020
11.	Plumericin	Terpenoids	Antibacterial,antifungal, anti-inflammatory, antiparasitic, antiviral, cytotoxic, algicidalactivity	Coppen and Cobb, 1983
12.	2,6,10,14,18,22 -Tetracosahexaene		Anti-inflammatory and anti- oxidant used in nutraceutical and pharmaceutical industries	Kim and Karadeniz, 2012
13.	2,3-Dihydrobenzofuran	Bicyclic derivative	Anticancer, antifungal, antibac- terial activity	Mohamad et al., 2020
14.	7-Hydroxy-6-methoxy-2H-1	Coumarin	Antioxidantactivity	De Siqueira Leite et al.,
	-benzopyran-2-one			2015
15.	2,6-Dimethylbenzoquinone-4 -oxime	Quinone	Anticancer agent	Görner, 2019
16.	4'-methoxybiphenyl-3- carboxylic acid	Biphenyl derivative	Antibacterialactivity	Kuppusamy et al., 2018
17.	2,2-Diethoxytetrahydrofuran	Cyclic ether	Antioxidantactivity	Jongcharoenkamol et al., 2023
18.	Ethyl 4-ethoxybenzoate	Esters	Antibacterialactivity	Wright et al., 2021
19.	2-(2-Butoxyethoxy) ethyl thiocyanate	Alcohol	Insecticide	Hideo and Casida, 1971
20.	Diborane(6)	Inorganic compound	Rubber vulcanizer	Borthakur et al., 2019

Table 2 : Phytochemical compounds with their biological activity in *P. obtusa* root extract

Twenty compounds identified in *P. obtusa* stem extract (Table 3, Figure 2) are Urs-12-en-24-oic acid 3-oxo-methylester (27.43%), alpha-d-glucopyranoside (9.29%), alpha-amyrin (8.41%)9, 19-cyclolanost-24-en-3-ol (4.66%), 4'-methoxybiphenyl-3-carboxylic acid(4.14%), benzoic acid,4-ethoxy-ethylester (3.93%), 12-oleanen-3-ylacetate, (3alpha)- (3.70%), methylbeta-d-glucopyranoside (3.67%), 2-(2-Butoxyethoxy) ethyl thiocyanate (3.39%), 2,3-dihydrobenzofuran (2.72%), n-hexadecanoicacid (2.06%), alpha-

d-galactopyranoside methyl (1.91%), linoleic acid ethyl ester (1.86%), 4-O-methylmannose (1.82%), 9-octadecenoic acid (Z)ethyl ester (1.46%), 9,12-octadecadienoic acid (Z,Z)-methyl ester (1.30%), beta-amyrin (1.17%), ethyl oleate (1.12%), hexadecanoic acid methyl ester(1.12%) and hexadecanoic acid ethyl ester(1.08%). Biological activities of the phytochemical compounds of both root and stem extracts of *P.obtusa* have been mentioned in (Tables 2 and 4).





Figure 2: GC-MS elucidation of bioactive compounds in methanolic stem extract of *P. obtusa*.

S.No.	Compound	Molecular formula	Molecular weight (g/mol)	Area%	Retention time
1.	Urs-12-en-24-oic acid 3-oxo- methyl ester	$C_{31}H_{48}O_{3}$	468.7	27.43	18.063
2.	alpha-d-glucopyranoside,	$C_7 H_{14} O_6$	194.18	9.29	10.930
3.	alpha-amyrin	$C_{30}H_{50}O$	426.7	8.41	16.430
4.	9,19-Cyclolanost-24-en-3-ol	$C_{32}H_{52}O_{2}$	468.8	4.66	16.152
5.	4'-methoxybiphenyl-3-carboxylic acid	$C_{14}H_{12}O_{3}$	228.24	4.14	13.752
6.	Benzoic acid, 4-ethoxy- ethyl ester	$C_{11}H_{14}O_3$	194.23	3.93	10.053
7.	12-Oleanen-3-yl acetate (3 alpha)	$C_{32}H_{52}O_{2}$	468.8	3.70	17.163
8.	Methylbeta-d-glucopyranoside	$C_7 H_{14} O_6$	194.18	3.67	10.564
9.	2-(2-butoxyethoxy) ethyl thiocyanate	$C_9H_{17}NO_2S$	203.30	3.39	9.353
10.	2,3-Dihydrobenzofuran	C <sub>8</sub> H <sub>8</sub> O	120.15	2.72	7.220
11.	n-hexadecanoic acid	$C_{16}H_{32}O_{2}$	256.42	2.06	13.197
12.	alpha-d-galactopyranoside methyl	$C_7 H_{14} O_6$	194.18	1.91	10.597
13.	Linoleic acid ethyl ester	$C_{20}H_{36}O_{2}$	308.5	1.86	14.508
14.	4-O-methylmannose	$C_7 H_{14} O_6$	194.18	1.82	11.197
15.	9-Octadecenoic acid (Z)- methyl ester	$C_{19}H_{36}O_{2}$	296.5	1.46	14.130
16.	9,12-Octadecadienoic acid (Z,Z)- methyl ester	$C_{18}H_{32}O_{2}$	280.4	1.30	14.097
17.	Beta-amyrin	$C_{30}H_{50}O$	426.7	1.17	15.163
18.	Ethyl oleate	$C_{20}H_{38}O_2$	310.5	1.12	14.541
19.	Hexadecanoic acid methyl ester	$C_{17}H_{34}O_2$	270.45	1.12	12.986
20.	Hexadecanoic acid ethyl ester	$C_{18}H_{36}O_{2}$	284.5	1.08	13.430

Table 3: Compounds identified in the methanolic stem extract of P. obtusa

Table 4: Phytochemical compounds with their biological activity in P. obtusa stem extract

S.No.	Chemical compound	Nature of the compound	<b>Biological activity</b>	Reference
1.	Alpha-d-glucopyranoside	Glycosides	Pharmaceuticals and food industries	González et al., 2014
2.	Methylbeta-d-glucopyranoside		Antimicrobial glycoside	Olawumi et al., 2020
3.	Alpha-d-galactopyranoside methyl		Skin moisturizing agent, antitu- berculous, antioxidant, alpha amylase inhibitory, hypolipi- demic, anticonvulsant	Sonia and Singh, 2019
4.	4-O-methylmannose		Antiviral, antimicrobial activity	Ololade Zacchaeus et al., 2021
5.	n-hexadecanoic acid	Fatty acids	Antioxidant and hypocholestrol agent, in addition for use in soap and lubricant industry	Singh et al., 2024
6.	Linoleic acid ethyl ester		Antiarrithritic, antihistaminic and anti-inflammatory agents	Kolar et al., 2019
7.	9-Octadecenoic acid (Z)- methyl ester		Anti-inflammatory, anticancer activity	Mohamad et al., 2020
8.	9,12-Octadecadienoic acid (Z,Z)-methyl ester		Analgesic, anticancer, anti- inflammatory agent	Mohamad et al., 2020
9.	Ethyl oleate		Antibacterial activity	Ankita et al., 2015
10.	Hexadecanoic acid, methyl ester		Acidifier, arachidonic acid inhibitor, inhibits production of uricacid, urine acidifier	Perumal et al., 2021

11.	Hexadecanoic acid, ethyl ester		Antioxidant activity	Mohamad et al., 2020
12.	Urs-12-en-24-oic acid, 3-oxo- methyl ester	Triterpenoids	Anti-inflammatory,anticancer, antioxidant activity	Rai <i>et al.</i> , 2019
13.	alpha-amyrin		Anti-inflammatory, anticancer, antioxidantactivity	Okoye et al., 2014
14.	Beta-amyrin		Anti-inflammatoryactivity	Okoye et al., 2014
15.	12-Oleanen-3-yl acetate (3 alpha)		Antioxidant, antibacterial, anti- inflammatory, antitumor activities	Fabiyi <i>et al.</i> , 2012
16.	2,3-Dihydrobenzofuran	Bicyclic derivative	Anticancer, antifungal, antibac- terial activity	Mohamad et al., 2020
17.	4'-methoxybiphenyl-3- carboxylic acid	Biphenyl derivative	Antibacterialactivity	Kuppusamy et al., 2018
18.	Benzoic acid, 4-ethoxy- ethyl ester	Ester	Antimicrobial activity	Sheela and Uthayakumari, 2013
19.	2-(2-butoxyethoxy) ethyl thiocyanate	Alcohol	Insecticide	Hideo and Casida, 1971
20.	9,19-Cyclolanost-24-en-3-ol	Steroid derivative	Antioxidant and pharmaceutical properties	Sawale et al., 2019

Differentiation and overlapping of the compounds are tabulated in (Table 5) and the interactions among them are represented by Venn diagram (Figure 3). A total of eight compounds were identified in

common to both root and stem extracts. The root and stem extracts of *P. obtusa* are rich in bioactive compounds, making them a valuable source for pharmaceuticaland industrial applications, warranting further investigation and development.

# Table 5 : Comparision of compounds in root and stem extracts of P. obtusa

Common compounds in both rootand stem extracts	8	Linoleic acid ethyl ester, ethyl oleate, 9,12-octadecadienoic acid $(Z,Z)$ - methyl ester, hexadecanoic acid ethyl ester, methylbeta-d-glucopyranoside, 2,3-dihydrobenzofuran, 4'-methoxybiphenyl-3-carboxylic acid, 2-(2-butoxyethoxy) ethyl thiocyanate
Compounds in root extract	12	Ethylalpha-d-glucopyranoside,n-decanoic acid,7-octadecenoic acid methyl ester, methylbeta-d-galactopyranoside, 2,2-diethoxytetrahydrofuran, methyl palmitate, ethyl 4-ethoxybenzoate, 2,6-dimethylbenzoquinone-4-oxime, diborane(6), plumericin, 7-hydroxy -6-methoxy-2H-1-benzopyran-2-one, 2,6,10,14,18,22-tetracosahexaene
Compounds in stem extract	12	Urs-12-en-24-oic acid3-oxo- methyl ester, alpha-d-glucopyranoside methyl ester, alpha-amyrin,9,19-cyclolanost-24-en-3-ol,benzoic acid 4-ethoxy- ethyl ester,12-oleanen- 3-yl acetate(3alpha), n-Hexadecanoic acid,alpha-d-galactopyranoside, methyl, 4-O-methylmannose,9-Octadecenoic acid (Z)- methyl ester,beta-amyrin, hexadecanoic acid methyl ester



Figure 3 : Venn diagram showing significant common and different compounds of root and stem extracts of P. obtusa.

828

# 4. Discussion

Gas chromatography, known for its speed and precision, is ideal for analyzing and identifying volatile oils and heatsensitive compounds. It plays a crucial role in accurately determining phytoconstituents and profiling complex structures (Singh et al., 2024). Phytochemical analysis was conducted in the plant extracts, revealing constituents with recognized physiological and medicinal properties in Plumeria pudica (Bari et al., 2023). It has been reported that compounds such as terpenoids, flavonoids, iridoids, cardiac glycosides, phenolic acids, fatty acid esters, coumarins, steroids and cardenolides are responsible not only for antioxidant activity but also for exhibiting antiinflammatory effects in P. obtusa (Salar et al., 2022). The GC-MS analysis of P. obtusa root extract identified a variety of bioactive compounds with significant therapeutic potential. Ethyl alpha-dglucopyranoside and methyl beta-d-glucopyranoside are the glycosides in which they exhibit strong antioxidant and antimicrobial properties, aligning with their potential in skin care and pharmaceuticals (Olawumi et al., 2020; Sonia and Singh, 2019). 2,3dihydrobenzofuran have demonstrated notable anti-allergy, antioxidant, anti-inflammatory and antimalarial properties, as well as effective in treating skin conditions (O Moustafa, 2021). Anticancer and antibacterial activities were noted in 4'-methoxybiphenyl-3carboxylic acid, indicating their relevance in cancer and infection treatments (Kuppusamy et al., 2018; Mohamad et al., 2020). Compounds like n-decanoic acid and linoleic acid ethyl ester demonstrate anti-inflammatory and antihistaminic effects, supporting their use in treating inflammatory conditions. Plumericin, an iriod with its broad spectrum of antibacterial, antifungal and antiviral activities, highlights the plant's extensive therapeutic potential (Coppen and Cobb, 1983).Plumericin has been reported to exhibit algicidal, antileishmanial, molluscicidal and antifungal activities (Singh et al., 2011). Additionally, the presence of methyl palmitate and ethyl oleate suggests applications in food, cosmetics and antibacterial formulations (Ankita et al., 2015; Pinto et al., 2017). The identification of anticancer agents like 2,6-dimethylbenzoquinone-4-oxime, along with compounds such as 7-hydroxy-6-methoxy-2H-1-benzopyran-2-one and 2,6,10,14,18,22-tetracosahexaene with antioxidant and anti-inflammatory properties, underscores the plant's value in nutraceuticals and pharmaceuticals (de Siqueira Leite et al., 2015; Görner, 2019; Kim and Karadeniz, 2012).

The GC-MS analysis of stem extract from P. obtusa identified a diverse range of bioactive compounds with significant pharmacological potential. Ursane-type pentacyclic triterpenoids, such as urs-12en-24-oic acid 3-oxo- methyl ester, have gained attention as potential therapeutic agent due to its wide range of pharmacological properties. This compound demonstrates strong anti-inflammatory, anticancer and antioxidant activities, making it promising for usage in anticancer drug development (Sultana, 2017). Alpha-amyrin is identified for anti-inflammatory, anticancer and antioxidant activities, making it promising in therapeutic applications (Okoye et al., 2014; Rai et al., 2019). The presence of 9,19-Cyclolanost-24-en-3-ol, an antioxidant with pharmaceutical relevance, highlights the plant's potential in developing new drugs (Sawale et al., 2019). Additionally, compounds like 12-oleanen-3-yl acetate and 4'-methoxybiphenyl-3-carboxylic acid exhibit strong antibacterial and anti-inflammatory effects, which could be useful in treating infections and inflammatory diseases (Fabiyi et al., 2012; Kuppusamy et al., 2018). The antimicrobial properties of benzoic acid, 4-ethoxy-ethyl ester and beta-dglucopyranoside methyl, further underscore the potential of these stem extract in developing antimicrobial agents (Olawumi et al., 2020; Sheela and Uthayakumari, 2013). The identification of 2-(2-butoxyethoxy) ethyl thiocyanatehas been reported to possess insecticidal properties (Hideo and Casida, 1971). Bioactive compounds like the methyl ester of hexadecanoic acid have demonstrated potent antifungal properties, making them useful in combating fungal infections in plants and humans. On the other hand, n-hexadecanoic acid exhibits a broad range of health benefits, including antiinflammatory effects that help reduce swelling and pain. Its antimicrobial properties make it effective in inhibiting the growth of harmful bacteria, while its antioxidant activity helps neutralize free radicals, protecting cells from oxidative stress. These compounds are valuable for their therapeutic potential in both agriculture and medicine (Sharma et al., 2021). Moreover, linoleic acid ethyl ester and 9,12-octadecadienoic acid (Z,Z)-methyl ester are known for their anti-inflammatory and analgesic properties (Zubair et al., 2024) reinforcing the therapeutic potential of P. obtusa .Other noteworthy compounds include beta-amyrin and ethyl oleate, both known for their anti-inflammatory and antibacterial properties, which could be harnessed in developing treatments for inflammation and infections (Ankita et al., 2015; Okoye et al., 2014).

## 5. Conclusion

Based on the GC-MS analysis, the root and stem extracts of *P. obtusa* have been identified to possess promising bioactive compounds with significant therapeutic potential. The methanolic root and stem extract contains a diverse array of compounds like irioids, fatty acids, triterpenoids which are rich in antioxidant, antimicrobial, anti-inflammatory and anticancer properties, making it suitable for pharmaceutical, cosmetic and agricultural applications. The presence of compounds like ethyl alpha-d-glucopyranoside, 23-dihydrobenzofuran and plumericin highlights its potential for further development in therapeutic products. The bioactive properties observe in these extracts exhibits the way for advancing research and applications in the health and wellness industries. Further investigation is warranted to explore and optimize their use across various fields.

#### Acknowledgements

The authors acknowledge M/S. Greenmakers and Landscaping Maintenance LLC, Dubai for funding the research and providing fellowship for the student. The authors also acknowledge the Department of Floriculture and Landscaping for providing the field and laboratory facilities.

#### **Conflict of interest**

The authors declare no conflicts of interest relevant to this article.

## References

- Ankita, S.; Tribhuwan, S. and Rekha, V. (2015). GC-MS Analysis of bioactive phytoconstituents from *Rumex vesicarius* L, Intl. Res. J. Pharm., 6:269-272.
- Bari, M.; Alfaki, M. and Raja, K. (2023). Antiageing properties of *Plumeria pudica* Jacq. leaf extracts and development of antiageing cream for cosmetic applications. Ann. Phytomed., 12(1):343-352.
- Bihani, T.; Tandel, P. and Wadekar, J. (2021). *Plumeria obtusa* L.: A systematic review of its traditional uses, morphology, phytochemistry and pharmacology. Phytomedicine Plus, 1(2):100052.

- Borthakur, R.; Saha, K.; Kar, S. and Ghosh, S. (2019). Recent advances in transition metal diborane (6), diborane (4) and diborene (2) chemistry. Coordination Chemistry Reviews, 399:213021.
- Choudhary, M.; Kumar, V. and Singh, S. (2014). Phytochemical and pharmacological activity of Genus *Plumeria*: An updated review. Interna-tional Journal of Biomedical and Advance Research, 5(6):266-271.
- Coppen, J. J. and Cobb, A. L. (1983). The occurrence of iridoids in Plumeria and Allamanda. Phytochemistry, 22(1):125-128.
- de Siqueira Leite, K. C.; Torres, L. M.; Garcia, L. F.; Rezende, S. G; de Oliveira-Neto, J. R.; Lopes, F. M. and de Souza Gil, E. (2015). Eletrochemical characterization of scopoletin, a 7-hydroxy-6-methoxy-coumarin. International Journal of Electrochemical Science, 10(7):5714-5725.
- Fabiyi, O. A.; Atolani, O.; Adeyemi, O. S. and Olatunji, G. A. (2012). Antioxidant and cytotoxicity of β-amyrin acetate fraction from *Bridelia ferruginea* leaves. Asian Pacific Journal of Tropical Biomedicine, 2(2):S981-S984.
- González, C.; Tapia, M.; Pérez, E.; Pallet, D. and Dornier, M. (2014). Main properties of steviol glycosides and their potential in the food industry: A review. Fruits, 69(2):127-141.
- Gorner, H. (2019). Handbook of Organic Photochemistry and Photobiology. CRC press, pp:683-714.
- Guha, G; Rajkumar, V.; Kumar, R. A. and Mathew, L. (2010). Aqueous extract of *Phyllanthus amarus* inhibits chromium (VI)-induced toxicity in MDA-MB-435S cells. Food and Chemical Toxicology, 48(1):396-401.
- Hideo, O. and Casida, J. E. (1971). Glutathione S-transferases liberate hydrogen cyanide from organic thiocyanates. Biochemical Pharmacology, 20(7):1708-1711.
- Jongcharoenkamol, J.; Jancharoen, K.; Batsomboon, P.; Ruchirawat, S. and Ploypradith, P. (2023). Divergent synthesis of isochroman-4-ols, 1, 3dihydroisobenzofurans, and tetrahydro-2H-indeno [2, 1-b] furan-2-ones via epoxidation/cyclization strategy of (E)-(2-stilbenyl/ styrenyl) methanols. Synthesis, 55(17):2757-2772.
- Kabir, A. and Furton, K. G (2021).Gas Chromatography. Elsevier, pp:745-791.
- Kim, S.-K. and Karadeniz, F. (2012). Biological importance and applications of squalene and squalane. Advances in Food and Nutrition Research, 65:223-233.
- Kitahara, T.; Koyama, N.; Matsuda, J.; Aoyama, Y.; Hirakata, Y.; Kamihira, S. and Sasaki, H. (2004). Antimicrobial activity of saturated fatty acids and fatty amines against methicillinresistant *Staphylococcus aureus*. Biological and Pharmaceutical Bulletin, 27(9):1321-1326.
- Kolar, M. J.; Konduri, S.; Chang, T.; Wang, H.; McNerlin, C.; Ohlsson, L. and Saghatelian, A. (2019). Linoleic acid esters of hydroxy linoleic acids are anti-inflammatory lipids found in plants and mammals. Journal of Biological Chemistry, 294(27):10698-10707.
- Kuppusamy, R.; Yasir, M.; Berry, T.; Cranfield, C. G; Nizalapur, S.; Yee, E. and Cornell, B. (2018). Design and synthesis of short amphiphilic cationic peptidomimetics based on biphenyl backbone as antibacterial agents. European Journal of Medicinal Chemistry, 143:1702-1722.
- Mohamad, S. A.; Adzahar, N. S.; Akhtar, M. N.; Zareen, S. and Lee, T. C. (2020). Phytochemical analysis and GC-MS profiling in the flower of *Plumeria alba*. Materials Science Forum, 981:282-284
- Musini, A.; Rao, M. J. P. and Giri, A. (2013). Phytochemical investigations and antibacterial activity of *Salacia oblonga* Wall ethanolic extract. Ann. Phytomed., 2(1):102-107.

- Muthulakshmi, A. and Mohan, V. (2012). GC-MS analysis of bioactive components of *Feronia elephantum Correa* (Rutaceae). Journal of Applied Pharmaceutical Science, 2(2):69-74.
- Moustafa, G. (2021). Synthesis of dibenzofurans possessing antiallergy, antioxidant, anti-inflammatory, antimalarial and treatment of skin conditions. Egyptian Journal of Chemistry, 64(5):2539-2556.
- Okoye, N. N.; Ajaghaku, D. L.; Okeke, H. N.; Ilodigwe, E. E.; Nworu, C. S. and Okoye, F. B. C. (2014).Beta-amyrin and alpha-amyrin acetate isolated from the stem bark of *Alstonia boonei* display profound anti-inflammatory activity. Pharmaceutical Biology, 52(11):1478-1486.
- Olawumi, O. O.; Olakunle, F. A. and Koma, O. S. (2020). Stigma-5, 22,-diene-3-O-beta-D-glucopyranoside: A new antimicrobial glycoside from *Tetrapleura tetraptera*. World Journal of Biology Pharmacy and Health Sciences, 4(3):021-042.
- Ololade Zacchaeus, S.; Anuoluwa Iyadunni, A.; Adejuyitan Johnson, A. and Uyaboerigha Daubotei, I. (2021). Black velvet tamarind: phytochemical analysis, antiradical and antimicrobial properties of the seed extract for human therapeutic and health benefits. J. Phytopharm., 10:249-255.
- Perumal, G; Prabhu, K.; Rao, M.; Janaki, C.; Kalaiyannan, J. and Kavimani, M. (2021). The GC-MS analysis of ethyl acetate extract of one herbal plant, *Canthium parviflorum*. NVEO-Natural Volatiles and Essential Oils Journal, 20:4041-4047.
- Pinto, M. E.; Araújo, S. G.; Morais, M. I.; Sa, N. P.; Lima, C. M.; Rosa, C.A. and Lima, L. A. (2017). Antifungal and antioxidant activity of fatty acid methyl esters from vegetable oils. Anais da Academia Brasileira de Ciências, 89(03):1671-1681.
- Rai, S. N.; Zahra, W.; Singh, S. S.; Birla, H.; Keswani, C.; Dilnashin, H. and Singh, S. P. (2019). Anti-inflammatory activity of ursolic acid in MPTPinduced Parkinsonian mouse model. Neurotoxicity Research, 36:452-462.
- Salar, S.; Sharma, P.; Lamba, H. S.; Sharma, J. and Kaur, A. (2022). Exploration of antioxidant activity of *Plumeria obtusa* L. Ann. Phytomed., 11(2):532-539.
- Sawale, J.; Patel, J. and Kori, M. (2019). Antioxidant properties of cycloartenol isolated from *Euphorbia neriifolia* leaves. Indian Journal of Natural Products, 33(1):60-64.
- Sharma, B. and Alam, A. (2022). Phytochemical profiling, antioxidant potential and antimicrobial activities of *Dalbergia sissoo* Roxb. Ann. Phytomed., 11(1):383-388.
- Sharma, B.; Sharma, S. C. and Alam, A. (2021). Phytochemical screening and GC-MS analysis of *Tamarindus indica* L.(Angiosperms: Fabaceae). Ann. Phytomed., 10(1):215-221.
- Sheela, D. and Uthayakumari, F. (2013). GC-MS analysis of bioactive constituents from coastal sand dune taxon-Sesuvium portulacastrum (L.). Bioscience Discovery, 4(1):47-53.
- Shettima, A.; Karumi, Y.; Sodipo, O.; Usman, H. and Tijjani, M. (2013). Gas Chromatography-Mass Spectrometry (GC-MS) analysis of bioactive components of ethyl acetate root extract of *Guiera senegalensis* JF Gmel. Journal of Applied Pharmaceutical Science, 3(3):146-150.
- Singh, D.; Sharma, U.; Kumar, P.; Gupta, Y. K.; Dobhal, M. and Singh, S. (2011). Antifungal activity of plumericin and isoplumericin. Natural Product Communications, 6(11):1934578X1100601101.
- Singh, M.; Khan, M. I.; Badruddeen, J.A.; Ahmad, M.; Fatima, G.; Siddiqui, Z. and Islam, A. (2024). GC-MS analysis of bioactive compounds,

standardization, and assessment of antimicrobial efficacy of Himalayan *Juniperus communis* L. stems. Ann. Phytomed., **13**(1):815-824.

- Sonia, S. and Singh, S. (2019). Phytoconstituents of Ziziphus nummularia (Burm. f.) Wight and Arn. leaves extracts using GC-MS spectroscopy. Res. Rev. J. Life Sci., 9:109-118.
- Srivani, A. and Mohan, G (2023). GC-MS analysis and isolation of few bioactive phytoconstituents from *Ixora parviflora* Lam. Ann. Phytomed., 12(1):783-794.
- Sulaiman, S. F.; Yaacob, S. S.; Lan, T. M. and Muhammad, T. S. T. (2008). Chemical components of the essential oils from three species of *Malaysian Plumeria* L. and their effects on the growth of selected microorganisms. Tropical Life Sciences Research, 19(2):1-7.
- Sultana, B.; Anwar, F. and Ashraf, M. (2009). Effect of extraction solvent/ technique on the antioxidant activity of selected medicinal plant extracts. Molecules, 14(6):2167-2180.
- Sultana, N. (2017). Triterpenes and triterpenoids clinically useful with multiple targets in Cancer, Malaria and more treatment: Focus on potential therapeutic value. International Journal of Biochemistry Research and Review, 16(2):1-35.
- Wright, J.; Masown, K. and Ruffell, S. E. (2021). Comparison of antibacterial *Apis mellifera* honey varietals from North Eastern United States. Bios., 91(3):180-187.
- Zubair, A.; Akhtar, J.; Badruddeen, B.; Ahmad, M. and Khan, M. (2024). GC-MS profiling and phytopharmacological analysis of aqueous distillate of *Cyperus scariosus* R.Br. Ann. Phytomed., 13(1):1132-1140.

J. Jaahnavi, S.P. Thamaraiselvi, M. Ganga, N. Sritharan and P. Malathi (2024). Exploring the phytochemical constituents of *Plumeria obtusa* L. root and stem extracts through GC-MS analysis. Ann. Phytomed., 13(2):822-831. http://dx.doi.org/10.54085/ap.2024.13.2.84.