DOI: http://dx.doi.org/10.54085/ap.2024.13.2.54

Annals of Phytomedicine: An International Journal http://www.ukaazpublications.com/publications/index.php

Print ISSN: 2278-9839

Online ISSN : 2393-9885

Original Article : Open Access

Antidiabetic, antioxidant, and hepatoprotective activities of methanolic *Tamarindus indica* L. flower extract

Md. Nadeem Bari*•, Md. Rizwan Ansari*, Imran Mohammad* and Mohammad Anwar**

*Department of Basic Medical Sciences, College of Medicine, Prince Sattam bin Abdulaziz University, Al-Kharj-1942, Saudi Arabia **Shadan College of Allied Health Sciences, Hyderabad-500004, Telangana, India

Article Info

Abstract

Article history Received 12 October 2024 Revised 29 November 2024 Accepted 30 November 2024 Published Online 30 December 2024

Keywords Antidiabetic Antioxidant Hepatoprotective Oxidative stress Tamarindus indica L. Liver diseases are common chronic disorders that affect millions of people and often lead to serious complications, such as diabetes mellitus, which is linked to oxidative stress. Due to the side effects associated with many conventional chemical drugs, there is increasing interest in plant-based therapeutic agents. In this context, the present study investigated the antidiabetic, antioxidant, and hepatoprotective activities of the methanolic flower extract of *Tamarindus indica* L. The antidiabetic potential of the *T. indica* methanolic crude extract was demonstrated, showing 64% and 65% inhibition of α -amylase and α -glucosidase activity, respectively, at a concentration of 5 mg/ml. Additionally, antioxidant assays, including nitric oxide, DPPH free radical scavenging, ABTS scavenging, and scavenging tests, revealed significant antioxidant properties, with scavenging activities of 72%, 76%, and 82%, respectively, after treatment. The extract also exhibited hepatoprotective effects on HepG2 cells. Based on these findings, the study suggests that *T. indica* methanolic crude extract holds potential for further research as a therapeutic option for diabetes mellitus.

1. Introduction

World wide, chronic liver disease is a non-alcoholic fatty liver disease (NAFLD) and is often the cause of a chief public health concern that affects approximately 30% of the adult population and encounters two million deaths every year (Younossi et al., 2016). NAFLD comprises a huge spectrum of disease conditions characterized by simple hepatic steatosis to non-alcoholic steatohepatitis which may develop when there are no other reasons for secondary hepatic fat buildup, such as cirrhosis brought on by excessive alcohol use liver fibrosis, and elevated risk of problems from the liver, such as endstage liver disease and hepatocellular carcinoma, liver transplantation requirement leads liver associated mortality (Chalasani et al., 2018). Evidence indicates the NAFLD global burden expands apart from liver-related complications which raise the chance of getting type 2 diabetes mellitus and other metabolic diseases, metabolic syndrome, heart related conditions, and dyslipidemia (Long et al., 2020; Loomba et al., 2020; The European Association for Liver Research, 2016). Among these, the second most common metabolic liver disorder is type 2 diabetes mellitus, which is prevalent among patients with liver diseases. Liver disorders have significantly increased mortality and morbidity rates, and their progression poses a substantial challenge to humanity (Das et al., 2022; Zhou et al., 2017; Anstee et al., 2013).

Corresponding author: Dr. Md. Nadeem Bari Department of Basic Medical Sciences, College of Medicine, Prince Sattam bin Abdulaziz University, Al-Kharj, 11942, Saudi Arabia E-mail: m.bari@psau.edu.sa Tel.: +966509280389

Copyright © 2024Ukaaz Publications. All rights reserved. Email: ukaaz@yahoo.com; Website: www.ukaazpublications.com One of the most important chronic metabolic disordersdiabetes mellitus (DM) is characterized by elevated sugar levels in the blood sub sequently pancreatic deficiency like insulin production or a reduced cell sensitivity to insulin leads to diabetic conditions causing mild to serious complications up to death, if unnoticed or untreated (Wild et al., 2004). Worldwide, diabetes is ranked sixth leading cause of death owing to its associated complications, and diabetes patients increasing number executes an important socio-economic burden on the healthcare system and also, work loss and wages (WHO, 2016; Deshpande et al., 2008). Diabetes pathogenesis is presently attributed to risk factors including metabolic abnormalities and endogenous factors like genes and also, exogenous factors including environment and behaviour (Wu et al., 2017). Consequently, oxidative stress is an important endogenous factor and the main cause of diabetes complications (Wright et al., 2006). The relationship between diabetes and oxidative stress has been explained via molecular mechanisms wherein the elevated reactive oxygen species production is associated with hyperglycemia causingan imbalance that leads to oxidative stress (Folli et al., 2011). Therefore, a cost effective treatment option is urgently required to manage diabetes and possesses antioxidant property and also, hepatoprotective properties.

In these circumstances, the natural product is highly considered for its long lasting history of medical and health benefits on human health (Dias *et al.*, 2012). Amongst, plants are the most examined source due to their wide distribution and their variety range of biological properties like antibacterial, antiageing, anti-inflammatory, anticancer, and antiviral (Xu *et al.*, 2017). Consequently, *T. indica* generally known as tamarind is an evergreen leguminous tree from Caesalpiniaceae. The pharmaceutical values of the different plant parts are mainly associated with various phytochemicals including





saponin, tannins, flavonoids, steroids, polyphenols, and alkaloids (Komakech *et al.*, 2019). Hence, our study investigated the

2. Materials and Methods

2.1 Plant authentication

flower methanolic extract.

The plant used in the present study was identified as *T. indica* by Dr. Shamna, Department of Herbal Medicine, Deseeya Ayurvedic Pharmacy, Calicut, India with authentication number DAP/22-20/2024.

antidiabetic, antioxidant, and hepatoprotective properties of T. indica

2.2 T. indica methanolic crude extracts preparation

The gathered *T. indica* flowerwas sundried and a roughly powered 20 g. *T. indica* flower was added to a fresh cellulose thimble that was inserted into the Soxhlet device as per standard protocol (Harley *et al.*, 2022). Adding methanol to the flask initiated the reaction and continued for many hours until it wasa clear solvent and the obtained resultant was used for further studies.

2.3 Alpha-amylase inhibition activity

The ability of *T. indica* flower methanolic extract to inhibit alphaamylase activity was studied as explained earlier (Alqahtani *et al.*, 2019). The range of concentrations, in short is (1 mg/ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, and 5 mg/ml) of *T. indica* flower methanolic extract as well as the enzyme amylase in 20 mM for10 min; the buffer of sodium phosphate (pH 6.8) was kept at 25°C. A 1% starch solution was added to a 0.02 M sodium phosphate buffer (pH 6.9) to the pre-incubated above mixtures and allowed for 15 min. After adding 1.0 ml of dinitrosalicylic acid to halt the reaction, the mixtures were incubated for 5 min in a water bath. The dilution was carried out in the response mixtures and the optical density was measured at 540 nm to calculate amylase activity inhibition percentage.

2.4 Glucosidase activity inhibition assay

The ability of *T. indica* flower methanolic extract to inhibit glucosidase activity was examined as shown previously (Elya *et al.*, 2008). In short, the varying ranges of amounts of (1 mg/ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, 5 mg/ml) of *T. indica* flower methanolic extract was incubated along with α -glucosidase (0.01 mg/ml) for 10 min. The reaction was started with 5 mM of p-nitrophenyl- α -D-glucopy ranoside (pNPG) addition to the above mixture and continued for 60 min. The 0.1 M Na₂CO₃ was included to stop the response and the final product was measured at 400 nm to calculate the glucosidase action inhibition percentage.

2.5 DPPH free radical scavenging assay

T. indica flower methanolic extract the ability to scavenge radicals was investigated using the DPPH (2, 2-diphenyl-1-picryhydrazyl) assay as stated previously (Gayathri and Sathish Kumar, 2016). In short, the varying concentrations (1 mg/ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, 5 mg/ml) of *T. indica* flower methanolic extract (3 ml) were added in DPPH solution and allowed for 30 min in dark situation. The radical scavenging activity percentage was calculated after reading the final product at 517 nm:

Scavenging effect =
$$100 \times \frac{(\text{blank OD} - \text{sample OD})}{\text{blank OD}}$$

2.6 ABTS scavenging assay

The ABTS (2, 2'-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid)) scavenging activity of *T. indica* flower methanolic extract was evaluated, as previously reported by Pacifico *et al.* (2018). Briefly, ABTS solution was prepared by mixing 7 mM ABTS in water with 2.45 mM potassium persulfate (KSO) and allowing the reaction to proceed in the dark for 16 h. The resultant ABTS solution was then diluted with 0.1 M sodium phosphate buffer (pH 7.4) to achieve an absorbance of 0.750 ± 0.025 at 734 nm. Subsequently, *T. indica* flower methanolic extract at various concentrations (1 mg/ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, and 5 mg/ml) was mixed with 1 ml of ABTS solution and allowed to react for 6 min. The absorbance of the final reaction product was measured at 734 nm, and the percentage of ABTS scavenging activity was calculated:

ABTS scavenging effect (%) = $100 \times \frac{(\text{control} \text{OD} - \text{sample} \text{OD})}{\text{control} \text{OD}}$

2.7 Nitric oxide scavenging activity

The methanolic extract of *T. indica* flowers demonstrated nitric oxide scavenging activity, as previously reported by Alam *et al.* (2013). Briefly, a reaction mixture containing *T. indica* flower methanolic extract at various concentrations (1 mg/ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, and 5 mg/ml) and 10 mM sodium nitroprusside in 0.5 M phosphate buffer (pH 7.4) was incubated for 60 min. After incubation, an equal volume of Griess reagent, composed of 1% sulfanilamide and 0.1% naphthyl ethylenediamine dihydrochloride in 2.5% phosphoric acid (1:1, v/v), was added to the reaction mixture. The resulting pink-colored product was measured at 540 nm to determine the percentage of nitric oxide scavenging activity:

Scavenging activity (%) = $100 \times \frac{(\text{controlOD} - \text{sampleOD})}{\text{controlOD}}$

2.8 Hepatoprotective activity of *T. indica* methanolic crude extract

The cytotoxicity of *T. indica* methanolic crude extract on HepG2 cells was evaluated using the MTT assay, as described by Meiyazhagan *et al.* (2015). HepG2 cells cultured in DMEM medium were treated with varying concentrations of *T. indica* methanolic crude extract (1 mg ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, and 5 mg/ml). After incubation, MTT solution was added to facilitate the formation of formazan crystals, followed by the addition of DMSO to solubilize the crystals and produce a purple-colored product. The absorbance of the product was measured at 570 nm to assess cell viability.

2.9 Statistical analysis

The standard and mean deviations were used for calculating error bars for all the experiments.

3. Results

3.1 Alpha-amylase inhibition activity

The ability of *T. indica* flower methanolic extract to inhibit α -amylase activity was assessed, and the percentage inhibition of α -amylase activity is presented in Figure 1. The figure shows that α -amylase activity was inhibited by 26%, 31%, 43%, 52%, and 64% following treatment with *T. indica* flower methanolic extract at concentrations of 1 mg/ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, and 5 mg/ml, respectively.

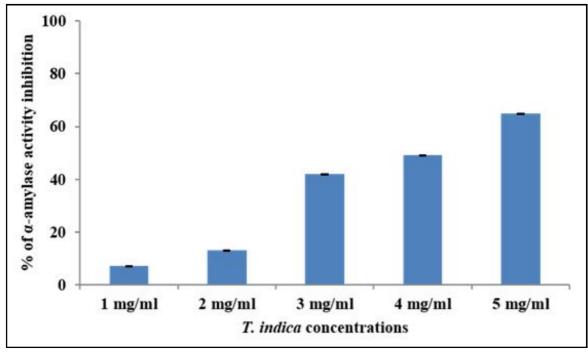


Figure 1: Effect of *T. indica* flower methanolic extract on α -amylase activity inhibition.

3.2 Glucosidase activity inhibition assay

The inhibitory effect of *T. indica* flower methanolic extract on glucosidase activity is shown in Figure 2. The graph illustrates the percentage inhibition of glucosidase activity following treatment with

varying concentrations of *T. indica* flower methanolic extract (1 mg/ ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, and 5 mg/ml). The extract effectively inhibited glucosidase activity, with inhibition rates of 7%, 13%, 42%, 49%, and 65%, respectively.

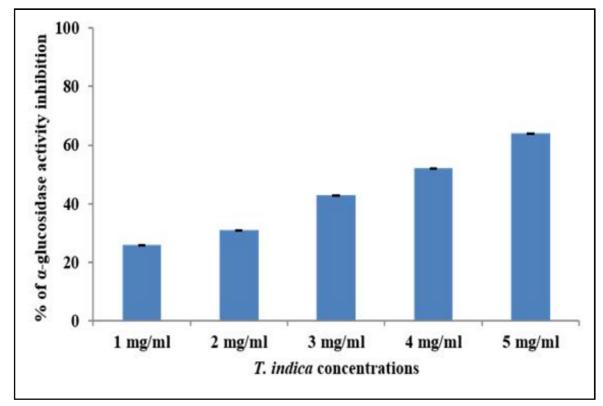


Figure 2: Effect of *T. indica* flower methanolic extract on α-glucosidase activity inhibition.

540

3.3 DPPH free radical scavenging assay

The radical scavenging capacity of *T. indica* flower methanolic extract was evaluated using the DPPH assay, and the calculated scavenging activity percentages are presented in Figure 3. The figure demonstrates that *T. indica* flower methanolic extract exhibited scavenging activities of 41%, 48%, 60%, 69%, and 72% at varying concentrations, highlighting its potent antioxidant activity.

3.4 ABTS scavenging assay

The effect of *T. indica* flower methanolic extract on ABTS scavenging activity is shown in Figure 4. The graph illustrates the calculated scavenging activity percentages after treatment with various

concentrations of *T. indica* flower methanolic extract (1 mg/ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, and 5 mg/ml). The extract exhibited ABTS scavenging activities of 47%, 49%, 58%, 70%, and 76%, respectively, demonstrating its strong antioxidant potential.

3.5 Nitric oxide scavenging activity

The effect of *T. indica* flower methanolic extract on nitric oxide scavenging activity was evaluated, and the calculated scavenging activity percentages are presented in Figure 5. The graph indicates that *T. indica* flower methanolic extract exhibited nitric oxide scavenging activities of 15%, 51%, 58%, 69%, and 82% at various concentrations, demonstrating its potent ability to neutralize nitric oxide radicals.

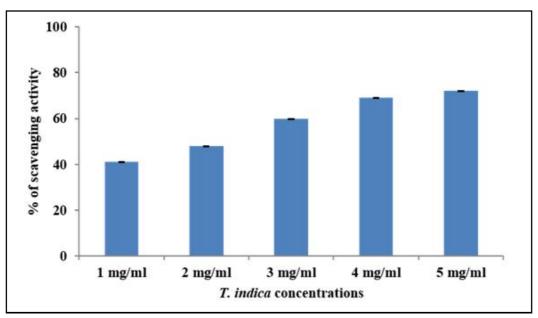


Figure 3: Effect of T. indica flower methanolic extract on free radical scavenging activity.

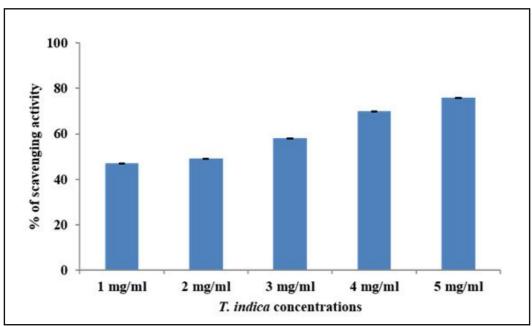


Figure 4: Effect of T. indica flower methanolic extract on ABTS scavenging activity.

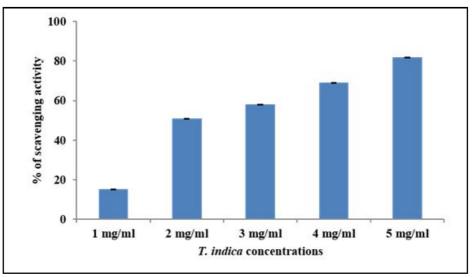


Figure 5: Effect of T. indicaflower methanolic extract on nitric oxide scavenging activity.

3.6 Hepatoprotective activity of *T. indica* methanolic crude extract

The hepatoprotective activity of *T. indica* methanolic crude extract on HepG2 cells was assessed using the MTT assay, and the calculated

cell viability percentages are presented in Figure 6. As shown in the figure, the graph illustrates the hepatoprotective activity of *T. indica* flower methanolic extract at varying concentrations (1 mg/ml, 2 mg/ml, 3 mg/ml, 4 mg/ml, and 5 mg/ml), with cell viability percentages of 93%, 87%, 41%, 40%, and 25%, respectively, after treatment.

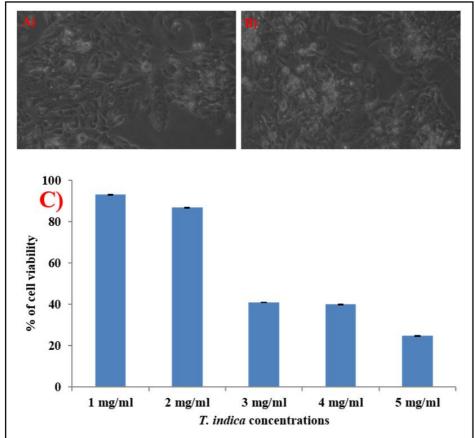


Figure 6: Effect of *T. indica* flower methanolic extract on hepatoprotective activity. (A) Untreated HepG2 cells, (B) HepG2 cells treated with *T. indica* flower methanolic extract, and (C) Graph represents the cell viability percentage after treatment.

542

4. Discussion

Liver disease is a significant chronic metabolic condition that can lead to mild to severe complications due to various factors. Among these complications, diabetes mellitus (DM) is a major metabolic disorder, often linked to risk factors such as oxidative stress. Therefore, our study investigated the antidiabetic, antioxidant, and hepatoprotective activities of the methanolic crude extract of T. indica and found it to exhibit potent antidiabetic, antioxidant, and hepatoprotective properties. α -amylase is a key enzyme responsible for breaking down dietary carbohydrates into simple sugars in the digestive tract, while α -glucosidases further degrade these simple sugars into glucose, which is then absorbed into the bloodstream. Given this, our study aimed to assess the ability of the methanolic T. indica crude extract to inhibit both α -amylase and α -glucosidase activities. Our results revealed the antidiabetic activity of T. indica crude extract. In favor of this, the study explored the methanolic raw extract of Nuxia oppositifolia antidiabetic and antioxidant of purified components. The purified compounds, katononic acid and 3oxolupenal were examined for α-glucosidase and α-amylase enzyme inhibition and revealed the potent inhibitory activity of both enzymes and also, the molecular docking highest binding affinity with active sites of both enzymes which provide primary information about N. oppositifolia use for diabetes mellitus (Algahtani et al., 2019). The in vitro analysis of various solvents (hexane, ethanol, ethyl acetate, and aqueous extracts) extracted Bridelia ferruginea investigated for antidiabetic and antioxidant analysis. The plant extract had the antioxidant property by scavenging the free radicals and exposed antidiabetic activity by inhibiting α -glucosidase, α -amylase, and lipase enzymes activities and showed good binding affinity with enzymes which exposed the antioxidant and antidiabetic potential of the plant (Oyebode et al., 2022).

Diabetes mellitus (DM) is caused by various risk factors, including metabolic abnormalities, endogenous factors, and exogenous influences. Oxidative stress, a significant endogenous factor, plays a major role in the complications associated with hyperglycemia, leading to an imbalance that triggers oxidative stress. In this context, the antioxidant properties of the methanolic T. indica crude extract were investigated, and the results demonstrated that T. indica extract exhibits excellent antioxidant activity, as evidenced by various assays. In support of this, the antioxidant, α -glucosidase, and α -amylase inhibitory properties of Canarium tramdenum fruits were also studied. Different solvent extracts of C. tramdenum revealed the presence of various phytochemicals and exhibited the most effective antioxidant properties. Additionally, the extract showed promising inhibitory effects on α -amylase and α -glucosidase, highlighting the biological potential of C. tramdenum (Quan et al., 2019; Segwatibe et al., 2023). Similarly, the methanolic extracts of Ageratum lanatum (L.) were analyzed for their chemical composition, revealing high concentrations of phenolic acids (PAs), which contributed to their significant antioxidant properties, as demonstrated through various assays. The identified fractions showed promising antidiabetic effects by inhibiting α -amylase activity (Pieczykolan *et al.*, 2021). Additionally, the hepatoprotective ability of T. indica was studied, and no toxic effects on cells were observed. Overall, these studies highlight the promising antidiabetic, antioxidant, and hepatoprotective activities of the extracts, suggesting that they could serve as effective agents for managing diabetes mellitus (DM).

5. Conclusion

As the prolonged use of many antidiabetic drugs can lead to severe side effects, researchers are exploring plant-based alternatives. The present study evaluated *T. indica* for its antidiabetic, antioxidant, and hepatoprotective activities. The methanolic crude extract of the plant demonstrated significant antidiabetic effects by inhibiting α amylase and α -glucosidase activities. Various assays confirmed its antioxidant properties, and the extract showed promising hepatoprotective effects when tested on liver cells. Based on these findings, the study recommends further detailed investigations of *T. indica* crude methanolic flower extract as a potential alternative to chemical drugs in the treatment of diabetes mellitus.

Acknowledgments

The authors are grateful to the Deanship of Scientific Research, Prince Sattam bin Abdulaziz University, Al-Kharj, Saudi Arabia for its support and encouragement in conducting the research and publishing this report.

Conflict of interest

The authors declare no conflicts of interest relevant to this article.

References

- Alam, M.N.; Bristi, N.J. and Rafiquzzaman, M. (2013). Review on *in vivo* and *in vitro* methods evaluation of antioxidant activity. Saudi Pharm. J.,21:143-152. doi: 10.1016/j.jsps.2012.05.002.
- Alqahtani, A.S.; Hidayathulla, S.; -Naumovski, V.; Alqahtani, M.S.; El, Dib, R.A. and AlAjmi, M.F. (2019). Alpha-Amylase and Alpha-glucosidase enzyme inhibition and antioxidant potential of 3-oxolupenal and katononic acid isolated from *Nuxia oppositifolia*. Biomolecules, 10(1):61.doi: 10.3390/biom10010061.
- Anstee, Q.M.; Targher, G. and Day, C.P. (2013). Progression of NAFLD to diabetes mellitus, cardiovascular disease or cirrhosis. Nat. Rev. Gastroenterol. Hepatol., 10(6):330-44.doi: 10.1038/nrgastro. 2013. 41.
- Chalasani, N.; Younossi, Z.; Lavine, J.E.; Charlton, M.; Cusi, K. and Rinella, M. (2018). The diagnosis and management of nonalcoholic fatty liver disease: practice guidance from the American association for the study of liver diseases. Hepatol., 67(1):328-57. doi: 10.1002/hep. 29367.
- Das, R.; Mitra, S.; Tareq, A.; Emran, T.; Hossain, M. and Alqahtani, A. (2022). Medicinal plants used against hepatic disorders in Bangladesh: A comprehensive review. J. Ethnopharmacol., 282:114588. doi:10. 1016/j.jep.2021.114588.
- Deshpande, A.D.; Harris-Hayes, M. and Schootman, M (2008). Epidemiology of diabetes and diabetes-related complications. Phys. Ther., 88:1254-1264. doi: 10.2522/ptj.20080020.
- Dias, D.A.; Urban, S. and Roessner, U. A. (2012). Historical overview of natural products in drug discovery. Metabolites, 2:303-336. doi: 10.3390/ metabo2020303.
- Elya, B.; Basah, K.; Bangun, A. and Septiana, E.K. (2012). Screening of αglucosidase inhibitory activity from some plants of Apocynaceae, Clusiaceae, Euphorbiaceae, and Rubiaceae. J. Biomed. Biotechnol., 2012:1-6. doi: 10.1155/2012/281078.
- European Association for the Study of the Liver (2016). Electronic address eee, European Association for the Study of D, European Association for the Study of O. EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. J. Hepatol., 64(6):1388-402. doi: 10.1016/j.jhep.2015.11.004.

- Folli, F.; Corradi, D.; Fanti, P.; Davalli, A.; Paez, A. and Muscogiuri, G. (2011). The role of oxidative stress in the pathogenesis of type 2 diabetes mellitus micro- and macrovascular complications: Avenues for a mechanistic-based therapeutic approach. Curr. Diabetes Rev.,7:313-324. doi:10.2174/157339911797415585.
- Gayathri, P.K. and Sathish Kumar, K. (2016). Antioxidant activity of essential oil extracted from enicostemmalittorale. J. Chem. Pharm. Sci., 9(1):256-258.
- Harley, B.K.; Quagraine, A.M.; Neglo, D.; Aggrey, M.O.; Orman, E. and Mireku-Gyimah, N.A. (2022). Metabolite profiling, antifungal, biofilm formation prevention and disruption of mature biofilm activities of *Erythrina senegalens* is stem bark extract against *Candida albicans* and *Candida glabrata*. PLOS ONE., 17(11):e0278096. doi: 10.1371/journal.pone.0278096.
- Komakech, R, Kim, Y.G.; Matsabisa, G.M. and Kang, Y. (2019). Antiinflammatory and analgesic potential of *Tamarindus indica* Linn. (Fabaceae): A narrative review. Integrat. Med. Res. 8(3):181-186. doi: 10.1016/j.imr.2019.07.002.
- Long, M.T.; Zhang, X.; Xu, H.; Liu, C.T.; Corey, K.E. and Chung, R.T. (2020). Hepatic fibrosis associates with multiple cardiometabolic disease risk factors: The Framingham heart study. Hepatol., doi:10.1002/hep.31608.
- Loomba, R.; Wong, R.; Fraysse, J.; Shreay, S.; Li, S. and Harrison, S. (2020). Nonalcoholic fatty liver disease progression rates to cirrhosis and progression of cirrhosis to decompensation and mortality: A realworld analysis of Medicare data. Aliment Pharmacol.Ther., 51(11):1149-59. doi:10.1111/apt.15679.
- Meiyazhagan, G; Winfred, S.B. and Ganesh, V. (2015). Bioactivity studies of βlactam derived polycyclic fused pyrroli-dine/pyrrolizidine derivatives in dentistry: *In vitro*, *in vivo* and *in silico* studies. PLoS ONE., 10(7):e0131433. doi: 10.1371/journal.pone.0131433.
- Oyebode, O.; Erukainure, O.L.; Koorbanally, N.A. and Islam, M.S. (2022). In vitro and computational studies of the antioxidant and antidiabetic properties of Bridelia ferruginea. J. Biomol. Struct. Dyn., 40(9): 3989-4003. doi: 10.1080/07391102.2020.1852961.
- Pacifico, S.; Galasso, S.; Piccolella, S.; Kretschmer, N.; Bauer, R. and Monaco, P. (2018). Winter wild fennel leaves as a source of anti-inflammatory and antioxidant polyphenols. Arab. J. Chem., 11:513-524. doi: 10.1016/j.arabjc.2015.06.026.

- Pieczykolan, A.; Pietrzak, W.; Gawlik-Dziki, U. and Nowak, R. (2021). Antioxidant, anti-inflammatory, and antidiabetic activity of phenolic acids fractions obtained from *Aerva lanata* (L.) Juss. Molecules, 26(12):3486. doi:10.3390/molecules26123486.
- Quan, N.V.; Andriana, Y. and Tuyen, P.T. (2019). Antioxidant, α-amylase and α-glucosidase inhibitory activities and potential constituents of *Canarium tramdenum* Bark. Molecules, 24(3):605. doi: 10.3390/ molecules24030605.
- Segwatibe, M.K.; Cosa, S. and Bassey, K. (2023). Antioxidant and antimicrobial evaluations of *Moringa oleifera* Lam. leaves extract and isolated compounds. Molecules, 28(2):899. doi: 10.3390/molecules 28020 899.
- Wild, S.; Roglic, G.; Green A.; Sicree, R. and King, H. (2004). Global prevalence of diabetes: Estimates for the year 2000 and projections for 2030. Diabetes Care. 27:1047-1053. doi: 10.2337/diacare.27.5.1047.
- World Health Organization (2016). Global report on diabetes. World Health Organization; Geneva, Switzerland: 2016. pp:1-88. Accessed on: 31.11.2024, https://www.who.int/publications/i/item/97892415 65257.
- Wright, E.; Scism-Bacon, J.L. and Glass L.C. (2006). Oxidative stress in type 2 diabetes: The role of fasting and postprandial glycaemia. Int. J. Clin. Pract. 60:308-314. doi: 10.1111/j.1368-5031.2006.00825.x.
- Wu, J.; Fang, X.; Yuan, Y.; Dong, Y.; Liang Y. and Xie, Q. (2017). UPLC/Q-TOF-MS profiling of phenolics from *Canarium pimela* leaves and its vasorelaxant and antioxidant activities. Braz. J. Pharmacogy., 27:716-723. doi: 10.1016/j.bjp.2017.10.005.
- Xu, D.P.; Li, Y.; Meng, X.; J.J. and Li, H.B. (2017). Natural antioxidants in foods and medicinal plants: Extraction, assessment and resources. Int. J. Mol. Sci., 18:96. doi: 10.3390/ijms18010096.
- Younossi, Z.M.; Koenig, A.B.; Abdelatif, D.; Fazel, Y.; Henry, L. and Wymer, M. (2016). Global epidemiology of nonalcoholic fatty liver disease-metaanalytic assessment of prevalence, incidence, and outcomes. Hepatol., 64(1):73-84. doi:10.1002/hep.28431.
- Zhou, M.; Wang, H.; Zeng, X.; Yin, P. and Zhu, J. (1990). Mortality, morbidity, and risk factors in China and its provinces, 1990-2017: A systematic analysis for the global burden of disease study 2017. Lancet. 394:1145-58. 10.1016/S0140-6736(19)30427-1.

Md. Nadeem Bari, Md. Rizwan Ansari, Imran Mohammad and Mohammad Anwar (2024). Antidiabetic, antioxidant, Citation and hepatoprotective activities of methanolic *Tamarindus indica* L. flower extract. Ann. Phytomed., 13(2):537-543. http://dx.doi.org/10.54085/ap.2024.13.2.54.