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# **Processing mediated changes on protein digestibility and iron bioavailability of selected underutilized millet and legume**

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# **1. Introduction**

Roasting

Since ancient times, millets and legumes have been consumed as staple foods in many Asian diets and cultures worldwide as they are considered as rich source of nutrients, including protein, vitamins, minerals, and antioxidants. Furthermore, because they are climateresilient and use less water, these smart food crops satisfy all the criteria for being advantageous for the environment, the farmer, and human health due to their low carbon emissions. In Uttarakhand, mainly grown millets are barnyard millet, finger millet and mainly grown legumes are horse gram, soybean, *etc*. Consumption of millets or whole grains with pulses or legumes is associated with alleviating non-communicable diseases, as they consist of good quality protein, minerals (iron and calcium) in addition to dietary fiber, phenols, and antioxidants (Mounika and Hymavathi, 2021).

Millet is one of the ancient or oldest foods known to human beings and may have been the first cereal grain utilized for domestic purposes. The small-seeded grasses that are available all year long are referred to as millets. These grasses are quite significant in both Asia and Africa (Pathak and Singh, 2022). Barnyard millet (*Echinochloa frumentacea*) has been grown for centuries because of

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its great nutritional value to humans and animals, as well as its resistance to harsh environmental conditions and low input cultivation. The grains offer a variety of nutrients, including dietary fiber, minerals, other trace elements, gluten-free protein, slowly digesting starch, and phytochemicals, particularly polyphenols, and also consist of a good amount of iron (Sood *et al.,* 2015). The use of barnyard millet grains and their value-added products has consequently drawn notice all around the world. As a result, this millet has emerged as a superfood in a variety of culinary applications. In general, Indian barnyard millet is well-characterized and has 10.4% protein, bioactive components (alkaloids, tannins, and flavonoids), and mineral (iron: 15.6-18.6 mg%) content, which promote health, in comparison to other types of major food grains (Ugare *et al.,* 2014). Foods include trace amounts of bioactive molecules, and research on how these substances affect human health is ongoing (Sharma and Sarwat, 2022). Legumes are essential to human nutrition because they supply enough proteins, calories, vitamins, minerals, and other bioactive ingredients (Deshpande, 1992), and their addition to composite flour of millet or wheat could increase the protein digestibility of flours (Mounika and Hymavathi, 2021). As they are rich in amino acids like lysine and tryptophan and far less expensive than animal proteins, legumes are consumed all over the world as an alternative source of protein (Pushpangadan *et al.*, 2014). Horse gram (*Macrotyloma uniflorum*) is a leguminous crop that is grown primarily in tropical Africa and Southeast Asia and is comparatively unknown. Aside from vitamins and minerals, it is one of the underutilized legumes with high protein content. It is known as the poor man's pulse crop and is considered a great solution to combat malnutrition problems in developing nations since it provides a less



expensive source of protein for low-income populations (Kushwaha and Raghuvanshi, 2010). It is an excellent source of protein, iron, molybdenum, and calcium (Pal *et al.,* 2016).

Although, millets and legumes have a diversified high food value and high nutritional value, their consumption, especially by the Indian populace, has not reached a significant level due to the presence of antinutrients and lack of awareness. Despite their high nutritional value and major production, these crops are being underutilized mainly because of the antinutritional factors (phytates, tannins, oxalic acid) that inhibit or reduce the absorption and digestibility of certain macro and micronutrients like protein and iron in the human body and make them less bioavailable.

Conventional processing methods such as soaking, fermentation, roasting, and germination have been used to improve the nutritional value of cereals and pulses. Roasting and germination are the two most widely employed processing methods for the preparation of food. These processing techniques are also known to reduce antinutritional compounds and trypsin inhibitors (Malomo *et al.,* 2014; Wang and Malcolmson, 2003). Processing techniques responsible for decreasing the anti-nutritional components, minimizing the losses of micro and macronutrients, and thus increasing their digestibility and bioavailability in the humans are of great interest to scientists.

VL-207 was chosen and identified due to its high yield advantage and high harvest index (25%). It has shown resistance to grain smut (6.67%), which is a common disease in barnyard millet. This highyielding cultivar is probably going to contribute in increasing the yield of barnyard millet and ensuring food and nutritional security for the farmers.

VLG-19 with a maturity period of 88-94 days and an average yield of 5-6 q/ha, has low tannin content with good digestibility. In addition to showing a modest resistance to some diseases like collar rot, anthracnose, leaf spot diseases, and powdery mildew, and, it is resistant to root rot as well. Hill farmer's interest in the profitable cultivation of horse gram can be revived by the enhanced cultivars' superior performance and profitability as compared to local varieties.

Thus, an attempt was made in the present investigation to study the effects of roasting and germination on nutritional content and their availability in a specific variety of barnyard millet (VL-207) and horse gram (VLG-19).

### **2. Materials and Methods**

### **2.1 Procurement of materials**

Seeds of a selected variety of barnyard millet (VL-207) and horse gram (VLG-19) used in the current investigation were procured from the Vivekananda Parvatiya Krishi Anushandhan Sansthan (VPKAS), Almora. The seeds were properly cleaned to remove impurities and foreign material and were stored in airtight containers. Chemicals used were purchased from well-known standard companies, including Sigma, BDH Chemicals, Hi-Media, Qualigens, and Merck India.

# **2.2 Processing of barnyard millet and horse gram**

Food processing is the process of converting raw ingredients into food or different forms of food through physical or chemical techniques. Processing methods such as roasting and germination

were implemented with raw samples to test the effect of processing on protein, iron content, *in vitro* protein digestibility (IVPD), and *in vitro* iron bioavailability (IVIB).

# **2.2.1 Roasting of barnyard millet**

Barnyard millet (BM) seeds were roasted by the method of Nithya *et al.* (2007) and Chabbra and Kaur (2022). BM seeds were roasted on a hot iron pan at 110°C for about 10 min. Then these roasted seeds were cooled and ground to flour using Bajaj Rex 750W Mixer Grinder (Pune, India) and passed through a 40 mesh size (BSS) sieve.

### **2.2.2 Germination of barnyard millet**

BM seeds were washed thrice and then soaked in tap water  $(1:3 \text{ w/v})$ for 12 h at  $27 \pm 3$ °C. Soaked seeds were kept for germination in muslin cloth at 33°C for about 48 h. Then the germinated seeds were dried in a tray drier (Sanco, India) at 45°C for 8 h to a final moisture content of 7%, cooled, and ground to get flour using Bajaj Rex 750W Mixer Grinder (Pune, India) and passed through a 40 mesh size (BSS) sieve (Sharma *et al.,* 2016).

### **2.2.3 Roasting of horse gram**

Horse gram (HG) seeds were roasted by the method optimized by Ojha *et al.* (2020) wherein seeds were roasted in an iron pan at 160°C for 10 min till they turned to brown colour and developed a roasted odour/smell. After letting the seeds cool, they were ground to make flour using a Bajaj Mixer Grinder, 750 W (Pune, India), and passed through a 40 mesh size (BSS) sieve.

### **2.2.4 Germination of horse gram**

HG seeds were cleaned, washed, and soaked in tap water in a glass container (1:5 w/v) at  $27 \pm 3$ °C for 18 h. Following the removal of the soaking water, the samples were left to germinate in muslin cloth at room temperature ( $27 \pm 3$ °C). Seeds were subsequently dried for 6 h at 55°C in a tray drier (Sanco, India) with up to 8% moisture content (Handa *et al.,* 2017; Banerjee *et al.,* 2022). Seeds were then cooled and ground to make flour using a Bajaj Rex 750 W Mixer Grinder (Pune, India) and passed through a 40 mesh size (BSS) sieve.

### **2.3 Nutritional analysis**

The nutritional analysis of raw, roasted, and germinated barnyard millet and horse gram involved the determination of protein content, iron content, IVPD, and IVIB. All the estimations were performed in triplicates.

### **2.3.1 Protein**

Protein content in the samples was determined by the AOAC (2010) method using the Micro Kjeldahl technique. The automatic nitrogen/ protein estimation system (Make: Borosil, Pune, India; Model: KBD061:digestor and KDI040: distillation) was used for the estimation. The protein content was obtained by multiplying per cent nitrogen by a conversion factor of 6.25 (Jones, 1941).

# **2.3.2 Iron**

For iron estimation, samples were wet-digested with 3:1 nitric acid: perchloric acid mixture, then cooled and filtered through Whatman No. 42 filter paper. The aliquot of each sample was used for the estimation of iron (Fe) using an atomic absorption spectrophotometer (Make: Electric Corporation of India Limited, New Delhi) by AOAC (1995) standard method.

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# **2.3.3** *In vitro* **protein digestibility (IVPD)**

IVPD was estimated using the procedure given by Akeson and Stahman (1964) by slightly modifying the method of Degroot and Slump (1969) in which the sample was digested with pepsin-HCl solution followed by further digestion with pancreatin and phosphate buffer mixture. After complete digestion mixture was precipitated, centrifuged, and filtered through Whatman No.54 filter paper. The nitrogen was estimated using Micro Kjeldahl assay and converted to protein using conversion factor 6.25.

# **2.3.4** *In vitro* **iron bioavailability (IVIB)**

IVIB of samples was estimated using the method described by Rao and Prabhavati (1978), which is based on the principle that food releases ionizable iron when it is exposed to the pepsin-HCl enzyme at a pH of 1.35 and is then adjusted to 7.5 to simulate the conditions found in the stomach and intestine, respectively. After the sample was extracted at pH 1.35 using pepsin-hydrochloric acid, the pH was raised to 7.5, followed by filtering the extract through Whatman No. 44 filter paper. Ionizable iron present in the filtrate reacts with alpha-alpha dipyridyl and was determined as given in the standard method (AOAC, 1965). The ionizable iron at pH 7.5 was used as an accurate measure of bioavailable iron.

### **2.4 Statistical analysis**

The study's findings were presented as the mean  $\pm$  standard deviation of three independent replications. The statistical significance of the generated results was obtained by WASP software using one-way analysis of variance. A difference was considered significant if it was *p*<0.05.

# **3. Results**

### **3.1 Effect of processing in BM**

The results showing the effect of roasting and germination and percent change on nutritional composition (protein, iron, IVPD, and IVIB) of BM have been presented in Table 1 and Figure 1, respectively.

# **3.1.1 Protein**

The protein content in the raw, roasted, and germinated BM flour ranged from 9.23 to 10.54% (Table 1) in which the protein content of germinated barnyard millet flour (10.54%) was significantly higher than raw BM (9.53%) followed by roasted BM (9.23%). An increase of 8.5% in protein content of germinated BM flour and a decrease of 6.82% in protein content of roasted BM flour was observed (Figure 1). The increase in protein content of germinated BM flour could

result from a drop in dry weight because germination produces some amino acids while respiration uses some lipids and carbohydrates (Jan *et al.,* 2017; Ongol *et al.,* 2013) and the decrease in protein content of roasted BM flour was due to protein degradation (Singh and Srivastava, 2006).

# **3.1.2 Iron**

Iron content in roasted (14.38 mg/100 g) and germinated barnyard millet (14.47%) flour was significantly higher than raw barnyard millet flour (14.18 mg/100 g) as shown in Table 1. The per cent increase in iron content of roasted and germinated BM is presented in Figure 1. Germination increases the mineral content in millets because, during this process, the phytase enzyme becomes more active, hydrolysing the link/bond between the phytic acid and the trapped minerals, releasing them and increasing their availability and content (Sharma *et al.,* 2015; Iswarya and Narayanan, 2016). The increase in iron content of roasted BM from raw BM could be because of leaching from the roasting iron pan into the millet (Obadina *et al.,* 2016).

#### **3.1.3** *In vitro* **protein digestibility (IVPD)**

IVPD of roasted BM flour (85.12%) and germinated BM flour (88.14%) was found to be significantly higher than raw barnyard millet flour (74.40%) as shown in Table 1 but a non-significant difference was observed between IVPD of germinated BM flour and roasted BM flour. There was a 14.41% and 18.47% increase in IVPD of roasted BM flour and germinated BM flour, respectively from raw BM flour (Figure 1). The improvement in IVPD after germination and dry heating (roasting) could be attributed to the reduction of anti-nutrients like phytic acid, tannins, and polyphenols, which interact with proteins to form complexes. It has been observed that heat processing increases the digestibility of proteins by eliminating protease inhibitors (Abbey and Berezi, 1988; Hassan *et al.*, 2006).

# **3.1.4** *In vitro* **iron bioavailability (IVIB)**

IVIB of raw, roasted, and germinated BM flour ranged from 11.02 to 24.39% in which the IVIB of germinated BM flour (24.39%) was significantly higher than roasted BM flour (21.56%), followed by raw BM flour (11.02%) as depicted in Table 1. A significant per cent increase in IVIB of roasted BM flour (95.64%) and germinated BM flour (121.32%) was observed (Figure 1). The increase in iron bioavailability after germination might result because of the leaching of the antinutritional compounds that bind the minerals (Idris *et al.,* 2007). During germination, there is a remarkable increase in phytase activity which decreases the amount of phytic acid that binds minerals and thus leads to improved mineral accessibility and availability (Luo *et al.*, 2014).

**Table 1: Effect of processing on nutritional composition of barnyard millet**

Parameter	Raw BM	<b>Roasted BM</b>	<b>Germinated BM</b>
Protein	$9.53 \pm 0.11^b$	$9.23 \pm 0.10^{\circ}$	$10.54 \pm 0.11^{\circ}$
Iron	$14.18 \pm 0.06^b$	$14.38 \pm 0.11$ <sup>a</sup>	$14.47 \pm 0.10^{\circ}$
In vitro protein digestibility	$74.40 \pm 6.45^b$	$85.12 \pm 6.41^{\circ}$	$88.14 \pm 6.11^a$
In vitro iron bioavailability	$11.02 \pm 0.87$ <sup>c</sup>	$21.56 \pm 0.70^b$	$24.39 \pm 0.73$ <sup>a</sup>

Values are the mean of replications  $(n = 3)$ 

<sup>ab</sup> values within the row with different superscript letters differed significantly ( $p$ <0.05) where, a>b>c



**Figure 1: Per cent change in nutritional composition of BM on processing.**

### **3.2 Effect of processing in HG**

Results showing the effect of roasting and germination and per cent changes on nutritional composition (protein, iron, IVPD, and IVIB) of HG have been presented in Table 2 and Figure 2, respectively.

### **3.2.1 Protein**

Protein content varied significantly with roasting and germination of HG as seen in Table 2. The germination showed a significant increase in the protein content from 24.94 to 27.06% whereas, roasting showed a significant reduction in the protein content of HG from 24.94 to 23.24% (Figure 2). The biological breakdown of several complex components into simpler molecules and the biosynthesis of proteins (amino acids) may be the cause of rise in the amount of protein during germination. The decline in protein content observed after roasting could be attributed to the denaturation of proteins and the release of nitrogenous volatile compounds at elevated temperatures (Wani *et al.,* 2017).

### **3.2.2 Iron**

Iron content in raw, roasted, and germinated HG is depicted in Table 2 which showed a significant increase in iron content of roasted  $(9.61 \text{ mg}/100 \text{ g})$  and germinated HG  $(10.98 \text{ mg}/100 \text{ g})$  when compared with raw HG flour (9.33 mg/100 g). A 17.68% increase in iron content was seen in germinated HG flour from raw horse gram flour (Figure 2). This increase could be due to the hydrolysis of iron compounds due to high water retention in germination (Poblete *et al.,* 2020).

### **3.2.3** *In vitro* **protein digestibility (IVPD)**

IVPD of germinated HG flour (83.75%) was significantly higher than roasted (74.56%) and raw HG flour (69.79%) as depicted in Table 2. An increase of 6.83% in roasted HG and up to 20% in germinated HG flour from raw HG flour was seen (Figure 2). Protein hydrolysis and the removal of antinutrients during germination may be the cause of the increased protein digestibility (Sharma *et al.,* 2019; Ganzle, 2020). The decrease or removal of anti-nutrients, the disintegration of phytic acid and native protein structures like lectins and enzyme inhibitors, increased activity of endogenous  $\alpha$ -galactosidase to reduce oligosaccharides might all lead to the improvement of IVPD in all processed seeds (Kalpanadevi and Mohan, 2013).

# **3.2.4** *In vitro* **iron bioavailability (IVIB)**

IVIB of raw, roasted, and germinated HG flour varied from 17.47 to 33.94% as depicted in Table 2. A significant per cent increase in IVIB of roasted (46.19%) and germinated HG flour (94.28%) from raw HG flour was observed (Figure 2). The germination process significantly increased the iron availability in horse gram from raw and roasted samples which were in agreement with the results reported by Sharma and Bhatnagar (2017). Phytic acid tends to bind cations, including calcium, iron, magnesium, and zinc. As a result of germination, the phytic acid is hydrolysed which makes these elements or minerals more bioavailable for the human body (Augustin and Klein, 1989).





Values are the mean of replications  $(n = 3)$ 

<sup>ab</sup> values within the row with different superscript letters differed significantly ( $p$ <0.05) where, a>b>c



**Figure 2: Per cent change in nutritional composition of HG on processing.**

# **4. Discussion**

Millets and legumes are abundant sources of protein, dietary fiber, polyphenols, and essential macro and micronutrients. Their nutritional content, digestibility, and nutrient availability are significantly affected by the methods of processing (Gowda *et al.,* 2022). Numerous studies demonstrate the substantial effects of different processing methods on the nutritional composition and the nutrient availability in BM and HG.

Kulla *et al.* (2021) observed a significant increase in the protein content of BM flour from 10.19 to 10.90% after germination. Sharma *et al.* (2016) also observed that the protein content of germinated BM (11.22%) increased significantly when compared to raw BM (9.14%) along with the iron content which increased from 37.68 to 39.42 mg/kg. These results were in contradiction with the study in which germination of sorghum was done at different time intervals, at different temperatures and reported that the protein content was negatively affected by the germination process (Raj and Singh, 2022). Nazni and Devi (2016) showed the high protein content in roasted BM flour  $(12.8 \text{ g})$  in comparison to raw  $(11.2 \text{ g})$  and germinated  $(8.9 \text{ g})$ g) BM flour. Contrarily, Anbalagan and Nazni (2020) observed a significant decline in the amount of protein in roasted BM (5.06%) from raw BM (5.62%). The rise in crude protein content in the sample subjected to the roasting treatment can be attributed to the use of low roasting temperatures, resulting in the generation of secondary metabolites that include protein (Mazni *et al*., 2023). Thus, low-temperature roasting increases protein content. It is recommended to roast barnyard millet at low temperatures to maximize protein availability and reduce denaturation, allowing the body to better utilize the protein.

Studies on finger millet and foxtail millet roasting showed an increase in iron content (3.45 to 3.91 mg/100 g) and (2.92 to 3.10 mg/100 g), respectively (Singh *et al.,* 2018; Khapre *et al.,* 2016) which were in contradiction with the study of Nazni and Devi (2016) who reported that the iron content of raw BM flour (9.00 mg/100 g) decreased in roasted (3.28 mg/100 g) and germinated BM flour (7.59 mg/100 g). Various researchers have reported increased protein digestibility (IVPD) of millets after a roasting or germination process (Omary *et* *al.,*2012; Pushparaj and Urooj, 2011). Experiments by Khetarpaul and Chauhan (1990) and Chaturvedi and Sarojini (1996) showed a notable increase up to 51 and 59%, respectively, in pearl millet protein digestibility after germination. Similarly, roasting significantly increased the IVPD of pearl millet (45.5 to 65.8% for the K variety and 49.3 to 75.4% for the MRB variety), indicating that dry heat treatment is superior to wet heat treatment in terms of improving protein digestibility (Pushparaj and Urooj, 2011). High protein content in germinated millet flours may be attributed to enzyme activation and synthesis of nitrogen-containing non-protein nucleic acids (Chethan *et al*., 2022). The high protein content, presence of free amino acids, and protein solubility in germinated millet flours can be linked to the activation of enzymes and the synthesis of proteins through biochemical events that occur during the germination process. Furthermore, it has been noted that the overall protein content of germinated grains is influenced by both the breakdown of proteins (proteolysis) and the creation of new proteins during the process of germination (Chetan *et al*., 2022).

Additionally, germination improved the bioavailable iron content of pearl millet by up to 2.2 times when compared to unprocessed grains (from  $0.5$  to  $1.1$  mg/100 g), and in finger millet, the bioavailable iron content increased from 0.4 to 1.3 mg/100 g as reported by Anitha *et al.* (2021). Thermal processing methods like roasting can improve micronutrient density in plant-based diets. Therefore, using these measures can enhance nutrient accessibility and reduce antinutrient content in food (Hotz and Gibson, 2007). Kalse *et al*. (2022) reported that roasting millet samples leached iron from the roasting pan leading to increase in their iron content.

The research carried out by Rizvi *et al.* (2022) showed that the protein content of HG increased after 72 h of germination from 20.66 to 23.01% resulting in an increase of 11.37% along with the increase in IVPD of germinated HG (86.10%) from raw HG (77.32%) which was also in agreement with the study conducted by Kachave *et al.* (2020) and Ghavidel and Prakash (2007). The rise in protein content may be attributed to protein production occurring during germination (Kachave *et al.*, 2020). Roasting resulted in a slight decrease in the protein content of HG from 21.40 to 20.60% (Thirukkumar and Sindumathi, 2014). Similarly, Kumar *et al.* (2020) also showed a decrease in the protein content of roasted chickpeas (13.00%) from the unroasted sample (14.80%). Roasting also resulted in a slight increase (3.2-15% increase) in protein digestibility (IVPD) of roasted soybeans when compared to their raw counterpart (Baik and Han, 2012; Raigar and Mishra, 2021). Various authors reported a significant increase in the iron content of germinated HG flour than roasted and raw HG flour (Rizvi *et al.,* 2022; Atudorei *et al.,* 2021; Pal *et al.*, 2016). Germination significantly increased the iron availability in HG from raw and roasted samples which were in agreement with the results reported by Sharma and Bhatnagar (2017) in which it was seen that the germination of HG increased IVIB from 26% to 37.93% and roasting of HG increased IVIB from 26% to 28.30%. Luo *et al.* (2014) reported that germination showed an enormous improvement in the iron bioavailability (IVIB) of faba bean (6.34 to 31.47%) and soybean (6.16 to 30.67%) which was a nearly fourfold increase. Our findings align with those presented by El-Adawy *et al*. (2002), who indicated that the loss of divalent metals (Ca, Fe, and Zn) during germination is minimal. This is attributed to their binding to proteins and the creation of phytatecation-protein complexes. Similarly, Ghavidel and Prakash (2007) revealed that the percentage of bioavailable iron in chickpeas, green gram, cowpeas, and lentils increased dramatically by 64.6%, 67.8%, 75.8%, and 81.3%, respectively, over the raw counterparts after germination. However, the iron content of chickpeas and cowpeas decreased during the same period. The increase in iron bioavailability in germinated horse gram flour is attributed to the soaking and germination treatment, which leads to a reduction in phytate concentration (Sharma and Bhatnagar, 2017). Phytate is a major inhibitor of iron and zinc absorption (Sandberg, 2002).

# **5. Conclusion**

The study demonstrates that germination and roasting significantly enhance the protein and iron content, as well as the digestibility and availability, of BM and HG flours. Empirical data show that germination notably increases nutrient content and availability compared to raw and roasted counterparts, primarily by reducing anti-nutritional factors through enzyme activation. However, future research is yet required to determine how these processing techniques affect other nutrients as well. These traditional processing methods improve nutrient bioavailability, making roasted and germinated millet or legume flours ideal for developing nutrient-rich products and therapeutic diets.

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# **Conflict of interest**

The authors declare no conflict of interest relevant to this article.

#### **References**

- **Abbey, B. W. and Berezi, P. E. (1988).** Influence of processing on the digestibility of African yam bean flour. Nutr. Rep. Int., **32**:819-827.
- **Akeson, W. R. and Stahmann, M. A. (1964).** A pepsin pancreatin digest index of protein quality evaluation. J. Nutr., **83**(3):257-261.
- **Anbalagan, S. and Nazni, P. (2020).** Effect of processing techniques on nutritional, viscosity and osmolarity of barnyard millet based diarrheal replacement fluids. Curr. Res. Nutr. Food Sci., **8**(1):164
- **Anitha, S.; Kane-Potaka, J.; Botha, R.; Givens, D. I.; Sulaiman, N. L. B.; Upadhyay, S. and Bhandari, R. K. (2021).** Millets can have a major impact on improving iron status, hemoglobin level, and in reducing iron deficiency anemia: A systematic review and meta-analysis. Front. Nutr., **8**:725529.
- **AOAC (1965).** Official Methods of Analysis. Association of Official Agricultural Chemists. 10th edn, AOAC International, Washington, **IISA**
- **AOAC (1995).** Official Methods of Analysis. Association of Official Agricultural Chemists. Arlington, Virginia. USA
- **AOAC (2010).** Official Methods of Analysis for protein in flour. Association of Official Analytical Chemistry. 19th edn, AOAC International, Washington, USA.
- **Atudorei, D.; Stroe, S. G. and Codinã, G. G. (2021).** Impact of germination on the microstructural and physicochemical properties of different legume types. Plants, **10**(3):592.
- **Augustin, J. and Klein, B. P. (1989).** Nutrient composition of raw, cooked, canned, and sprouted legumes. In: Mathews R.H., editor. Legumes: Chemistry. Technology and Human Nutrition, Mercel Dekker Inc.; New York, pp: 187-217.
- **Baik, B. K. and Han, I. H. (2012).** Cooking, roasting, and fermentation of chickpeas, lentils, peas, and soybeans for fortification of leavened bread*.* Cereal Chem., **89**(6):269-275.
- **Banerjee, S.; Haldar, S.; Reddy, N.; Reddy, R.; Nagananda, G. S. and Mitra, J. (2022).** Under-utilized germinated horse gram (*Macrotyloma uniflorum*) protein-Extraction, process optimization, characterization and its use in cookies fortification. LWT - Food Sci. Technol., **160**:113- 276.
- **Chaturvedi, A. and Sarojini, G. (1996).** Malting of pearl millet (*Pennisetum typhoideum*): its effect on starch and protein digestibilities. J. Food Sci. Technol*.*, **33**(4):342-344.
- **Chethan, K.P, Amutha, S.; Oberoi, H.S.; Kanchana, S. Azeez, S. and Rupa, T.R. (2022).** Germination induced changes in bioactive compounds and nutritional components of millets. J. Food Sci. Technol., **59**(11):4244-4252.
- **Chhabra, I. and Kaur, A. (2022).** Development of a convenient, nutritious ready to cook packaged product using millets with a batch scale process development for a small-scale enterprise. J. Food Sci. Technol*.*, **59**(2):488-497.
- **De Groot, A. P. and Slump, P. (1969).** Effects of severe alkali treatment of proteins on amino acid composition and nutritive value. J. Nutr., **98**(1):45-56.
- **Deshpande, S. S. (1992).** Food legumes in human nutrition: a personal perspective. Crit. Rev. Food Sci. Nutr., **32**(4):333-363.
- **Deshpande, S. S.; Mohapatra, D.; Tripathi, M. K. and Sadvatha, R. H. (2015).** Kodo millet-nutritional value and utilization in Indian foods. J. Grain Process. Storage, **2**(2):16-23.
- **El-Adawy, T.A. (2002).** Nutritional composition and antinutritional factors of chickpeas (*Cicer arietinum* L.) undergoing different cooking methods and germination. Plant Foods Hum. Nutr., **57**:83-97.
- **Gänzle, M. G. (2020).** Food fermentations for improved digestibility of plant foods: An essential ex situ digestion step in agricultural societies? Curr. Opin. Food Sci., **32**:124-132.
- **Ghavidel, R. A. and Prakash, J. (2007).** The impact of germination and dehulling on nutrients, antinutrients, *in vitro* iron and calcium bioavailability and *in vitro* starch and protein digestibility of some legume seeds. LWT-Food Sci. Technol., **40**(7):1292-1299.
- **Gowda, N. N.; Siliveru, K.; Prasad, P. V.; Bhatt, Y.; Netravati, B. P. and Gurikar, C. (2022).** Modern processing of Indian millets: A perspective on changes in nutritional properties. Foods, **11**(4):499.
- **Handa, V.; Kumar, V.; Panghal, A.; Suri, S. and Kaur, J. (2017).** Effect of soaking and germination on physicochemical and functional attributes of horsegram flour. J. Food Sci. Technol*.*, **54**:4229-4239.
- **Hassan, A. B.; Mohamed Ahmed, I. A.; Osman, N. M.; Eltayeb, M. M.; Osman, G. A. and Babiker, E. E. (2006).** Effect of processing treatments followed by fermentation on protein content and digestibility of pearl millet (*Pennisetum typhoideum*) cultivars. Pak. J. Nutr., **5**:86-89.
- **Hotz, C. and Gibson, R.S. (2007).** Traditional food-processing and preparation practices to enhance the bioavailability of micronutrients in plantbased diets. J. Nutr., 137(4):1097-1100.
- **Idris, W. H.; AbdelRahaman, S. M.; Elmaki, H. B.; Babikar, E. E. and Eltinay, A. H. (2007).** Effect of malt pre- treatment on HCL extractability of calcium, phosphorus and iron of sorghum (*Sorghum bicolor*) cultivars*.* Int. J. Food Sci., **42**(2):194-199.
- **Iswarya and Narayanan, A. (2016).** Effect of germination on biofortified pearl millet cultivars' nutrient content. Int. J. Innov*.* Res. Edu. Sci., **3**(6):2349-5219.
- **Jan, R.; Saxena, D. C. and Singh, S. (2017)**. Physico- chemical, textural, sensory and antioxidant characteristics of gluten free cookies made from raw and germinated Chenopodium (*Chenopodium album*) flour. LWT - Food Sci. Technol., **71**:281-287.
- **Jones, D. B. (1941).** Factors for converting percentages of nitrogen in foods and feeds into percentages of proteins. Washington, DC: US Department of Agriculture, **183**:22
- **Kachave, K. D.; Pawar, V. S. and Shinde, E. M. (2020).** Effect of germination treatment on chemical and nutritional properties of horse gram. The Pharma Innov. J., **9**(12):213-216
- **Kalpanadevi, V. and Mohan, V. R. (2013).** Effect of processing on antinutrients and *in vitro* protein digestibility of the underutilized legume, *Vigna unguiculata* (L.) Walp subsp. unguiculata. LWT - Food Sci. Technol., **51**(2):455-461.
- **Kalse, S.B.; Swami, S.B. and Jain, S.K. (2022).** Millet: A review of its nutritional content, processing and machineries. Int. J. Food Ferment. Technol. **12**(01):47-70.
- **Khapre, A.; Shere, D. and Deshpande, H. (2016).** Studies on effect of roasting on nutritional and anti-nutritional components of foxtail millet (*Setaria italica*). The Bioscan., **11**(1):177-179.
- **Khetarpaul, N. and Chauhan, B. M. (1990).** Effect of germination and fermentation on *in vitro* starch and protein digestibility of pearl millet. J. Food. Sci., **55**(3):883-884.
- **Kulla, S.; Hymavathi, T. V.; Kumari, B. A.; Reddy, R. G. and Rani, C. V. D. (2021).** Impact of germination on the nutritional, antioxidant and antinutrient characteristics of selected minor millet flours. Ann. Phytomed., **10**(1):178-184.
- **Kumar, Y.; Sharanagat, V. S.; Singh, L. and Mani, S. (2020).** Effect of germination and roasting on the proximate composition, total phenolics, and functional properties of black chickpea (*Cicer arietinum*). Legume Sci., **2**(1):e20.
- **Kushwaha, A. and Raghuvanshi, R.S. (2010).** Physicochemical and functional characteristics of different varieties of horse gram grown in Uttarakhand. Int. J. Food Sci. Technol. Nutr., **4**(1):81-92.
- **Luo, Y. W.; Xie, W. H.; Jin, X. X.; Wang, Q. and He, Y. J. (2014).** Effects of germination on iron, zinc, calcium, manganese and copper availability from cereals and legumes. CyTA- J. Food, **12**(1):22-26
- **Malomo, O.; Alamu, E. A. and Oluwajoba, S. O. (2014).** Effect of germination on the anti-nutritional and toxic factors of cowpea. Int. J. Fd. Sci. Nut. Eng., **4**(2):49–53.
- **Mazni, S.M.; Nurul, H.S.; Fairus, A. and Siti, R.S. (2023)**. Effect of roasting on whole grain barnyard millet to the proximate composition, amino acid profile, total phenolic content and antioxidant activity. Malays. J. Med. Health Sci., **19**(6):69-76.
- **Mounika, M. and Hymavathi, T. V. (2021).** Nutrient and phytonutrient quality of nutricereals incorporated flour mix suitable for diabetics. Ann. Phytomed., **10**(1):132-140.
- **Nazni, P. and Devi, R. S. (2016).** Effect of processing on the characteristics changes in barnyard and foxtail millet. J. Food Process. Technol.**, 7**:3
- **Nithya, K. S.; Ramachandramurty, B. and Krishnamoorthy, V. V. (2007).** Effect of processing methods on nutritional and anti-nutritional qualities of hybrid (COHCU-8) and traditional (CO7) pearl millet varieties of India. J. Biol. Sci., **7**(4):643-647.
- **Obadina, A.; Ishola, I. O.; Adekoya, I. O.; Soares, A. G.; de Carvalho, C. W. P. and Barboza, H. T. (2016).** Nutritional and physicochemical properties of flour from native and roasted whole grain pearl millet (*Pennisetum glaucum* (L.) R. Br.). J. Cereal Sci., **70**:247-252.
- **Ojha, P.; Bhurtel, Y.; Karki, R. and Subedi, U. (2020).** Processing effects on anti-nutritional factors, phytochemicals, and functional properties of horse gram (*Macrotyloma uniflorum*) flour. J. Microbiol. Biotechnol. Food Sci., **9**(6):1080-1086.
- **Omary, M. B.; Fong, C.; Rothschild, J. and Finney, P. (2012).** Effects of germination on the nutritional profile of gluten free cereals and pseudocereals: A review. Cereal Chem., **89**(1):1-14.
- **Ongol, M. P.; Nyozima, E.; Gisanura, I. and Vasanthakaalam, H. (2013).** Effect of germination and fermentation on nutrients in maize flour. Pak. J. Food Sci., **23**:183-188.
- **Pal, R. S.; Bhartiya, A.; ArunKumar, R.; Kant, L.; Aditya, J. P. and Bisht, J. K. (2016).** Impact of dehulling and germination on nutrients, antinutrients, and antioxidant properties in horsegram. J. Food Sci. Technol*.*, **53**(1):337-347.
- **Pathak, A. and Singh, S. P. (2022).** Study on the health benefit and utilization of sprouted grains for development of value-added food products: A review. Ann. Phytomed., **11**(2):155-165.
- **Poblete, T.; Rebolledo, K.; Barrera, C.; Ulloa, D.; Valenzuela, M.; Valenzuela, C. and González, C. (2020).** Effect of germination and cooking on iron content, phytic acid and lectins of four varieties of Chilean beans (*Phaseolus vulgaris*). J. Chil. Chem. Soc*.*, **65**(4):4937-4942.
- **Pushpangadan, P.; George, V.; Sreedevi, P.; Bincy, A. J.; Anzar, S.; Aswany, T.; Ninawe, A. S. and Ijinu, T. P. (2014).** Functional foods and nutraceuticals with special focus on mother and child care. Ann. Phytomed., **3**(1):4-  $24.$
- **Pushparaj, F. S. and Urooj, A. (2011).** Influence of processing on dietary fiber, tannin and *in vitro* protein digestibility of pearl millet. Food Nutr. Sci., **2**(8):895-900.
- **Raigar, R. K. and Mishra, H. N. (2021).** Impact of pilot scale roasting treatment on physical and functional properties of soybean (*Glycine max* L.). J. Inst. Eng. (India): Series A, **102**:489-498.
- **Raj, R. and Singh, S. P. (2022).** Study of functional properties of sorghum and their utilization in development of value-added functional bakery products. Ann. Phytomed., **11**(2):137-146.
- **Rao, B. N. and Prabhavathi, T. (1978).** An *in vitro* method for predicting the bioavailability of iron from foods. Am. J. Clin. Nutr*.*, **31**(1):169- 175.
- **Rizvi, Q. U. E. H.; Kumar, K.; Ahmed, N.; Chauhan, D.; Thakur, P.; Jan, S. and Sheikh, I. (2022).** Effect of processing treatments on nutritional, antinutritional, and bioactive characteristics of horse gram (*Macrotyloma uniflorum* L.). J. Postharvest Technol., **10**(2):48-59.
- **Sandberg, A.S. (2002).** Bioavailability of minerals in legumes. British J. Nutr., **88**:281-285.
- **Sharma, N. and Sarwat, M. (2022).** Functional foods for better health and weight loss. Ann. Phytomed., **11**(2):114-121.
- **Sharma, S.; Saxena, D. C. and Riar, C. S. (2015).** Antioxidant activity, total phenolics, flavonoids and anti-nutritional characteristics of germinated foxtail millet (*Setaria italica*). Cogent Food Agri., **1**(1):1081728.
- **Sharma, S.; Saxena, D. C. and Riar, C. S. (2016).** Analysing the effect of germination on phenolics, dietary fibres, minerals and  $\gamma$ -amino butyric acid contents of barnyard millet (*Echinochloa frumen taceae*). Food Biosci., **13**:60-68.
- **Sharma, S.; Singh, A. and Singh, B. (2019).** Characterization of *in vitro* antioxidant activity, bioactive components, and nutrient digestibility in pigeon pea (*Cajanus cajan*) as influenced by germination time and temperature. J. Food Biochem., **43**(2):e12706.
- **Sharma, V. and Bhatnagar, V. (2017).** Effects of processing on phytic acid, iron and its bioavailability of *Macrotyloma uniflorum*. Food. Sci. Res. J., **8**(1):128-131.
- **Singh, N.; David, J.; Thompkinson, D. K.; Seelam, B. S.; Rajput, H. and Morya, S. (2018).** Effect of roasting on functional and phytochemical constituents of finger millet (*Eleusine coracana* L.). J. Pharm. Innov., **7**(4):414-418.
- **Singh, P. and Srivastava, S. (2006).** Nutrient composition of some new varieties of finger millet (*Eleusine coracana*). J. Community Mobilization Sustainable Dev., **1**(1and2):115-120.
- **Sood, S.; Khulbe, R. K.; Kumar, A.; Agrawal, P. K. and Upadhyaya, H. D. (2015).** Barnyard millet global core collection evaluation in the submontane Himalayan region of India using multivariate analysis. The Crop J., **3**(6): 517-525.
- **Thirukkumar, S. and Sindumathi, G. (2014).** Studies on preparation of processed horse gram (*Macrotyloma uniflorum*) flour incorporated chappathi. Int. J. Sci. Res., **3**(3):110-111.
- **Ugare, R.; Chimmad, B.; Naik, R.; Bharati, P. and Itagi, S. (2014).** Glycemic index and significance of barnyard millet (*Echinochloa frumentacae*) in type II diabetics. J. Food Sci. Technol*.*, **51**:392-395.
- **Wang, N.; Daun, J. K. and Malcolmson, L. J. (2003).** Relationship between physicochemical and cooking properties, and effects of cooking on antinutrients of yellow field peas (*Pisum sativum*). J. Sci. Food Agric., **83**(12):1228-1237.
- **Wani, I. A.; Hamid, H.; Hamdani, A. M.; Gani, A. and Ashwar, B. A. (2017).** Physicochemical, rheological and antioxidant properties of sweet chestnut (*Castanea sativa* Mill.) as affected by pan and microwave roasting. J. of Adv. Res.**, 8**(4):399405.

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