DOI: http://dx.doi.org/10.54085/ap.2023.12.1.38

Annals of Phytomedicine: An International Journal http://www.ukaazpublications.com/publications/index.php

Print ISSN: 2278-9839

Online ISSN : 2393-9885

Original Article : Open Access

Bioinformatics approach to identify molecular targets of chrysin against Alzheimer's disease

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Article history Received 2 February 2023 Breised 20 Marsh 2023	
Revised 29 March 2023 Accepted 30 March 2023 Published Online 30 June-2023 Keywords Pharmacokinetic properties Drug-likeness Chrysin Molecular pathway Protein network ACCEPTE A D by gene-set enrichment and bioinformatics approach. The chry PubChem and ChEBI database, and the targets of chrysin were estimated using DIG DisGENET, PharmGKB, and Swiss Target Prediction were used to identify poss proteins. The drug-likeness properties and toxicity characteristics of chrysin we ADME and ProToxII databases. In addition, STRING and KEGG enrichment datab the role of probable interacting proteins to construct a protein-protein intera- network of molecular targeting pathways, respectively. Based on the results of j and drug-likeness analysis, chrysin predicted to have a good drug-likeness activit is good brain barrier permeability (BBB score = 3.71) with no observable toxicit identified as the top genes that interact with chrysin against Alzheimer's disea CASP3, PSEN1, PSEN2, PTGS2, NFKB1, AKT1, GSK3B, and APP were selected as top a significant role in AD treatment. Furthermore, a total of 158 different pathways probably modulated pathways, corresponding to 38 protein targets. Besides n pathways in cancer, lipid and atherosclerosis, EGFR tyrosine kinase inhibiti signaling in diabetic complications, HIF-1 signaling, P13K-Akt signaling, MAPK neurotrophin and sphingolipid signaling were defined as the top pathways associ proteins. Overall, the results indicated that the network-based approach could to uncover the therapeutic mechanisms of chrysin against AD.	Although, the biological erties have been previously e not been fully elucidated work-based pharmacology ial interactions of chrysin ysin was entered into the GEP-Pred. Then, GeneCards, sible interacting genes and ere determined using Swiss base were used to elucidate ction (PPI) network and a pharmacokinetic properties y (score = -0.21), as well y. A total of 38 genes were use. ILB, IL6, TNF, MAPK1, to core targets that may play yays were identified as the eurodegeneration and AD, tor resistance, AGE-RAGE signaling, IL-17 signaling, ated with chrysin-regulated

1. Introduction

Chrysin (5,7-dihydroxyflavone) is a natural flavonoid found in propolis, honey, fruits, and passion flowers. It is mainly present in the species of *Pleurotus ostreatus, Oroxylum indicum, Matricaria chamomilla, Passiflora incarnata* and *P. caerulea*. This natural compound has been studied for its potential pharmacological properties and it has been reported to have various pharmacological properties, such as antioxidant, anti-inflammatory, antiviral, antitumor, anticancer, antihyperlipidemic, antidepressant, and antibacterial activities (Mani and Natesan, 2018). Chrysin has been found to increase the therapeutic efficacy of docetaxel and mitigate docetaxel-induced edema. It has also been shown to target myeloidderived suppressor cells and enhance tumor response to anti-PD-1 immunotherapy (Stompor-Goracy, 2021; Li *et al.*, 2022). Chrysin

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Copyright © 2023 Ukaaz Publications. All rights reserved. Email: ukaaz@yahoo.com; Website: www.ukaazpublications.com induces apoptosis in cancer cells by activating caspase-3 and PLC-\lambda1 degradation, downregulating XIAP and inactivating Akt (Khoo et al., 2010). Additionally, chrysin has been reported to have antiinflammatory effects by downregulating the key pro-inflammatory enzymes, inducible nitric oxide synthase (iNOS) and cyclooxy-genase-2 (COX-2) (Zeinali et al., 2017). Chrysin has also been found to possess cardioprotective activity by improving post-ischemic functional recovery and suppressing vascular endothelial growth factor (VEGF)-induced angiogenesis (Kasala et al., 2015). Chrysin also shows promise as an anxiolytic and neuroprotective agent, with some studies indicating its potential for improving cognitive function and memory. Additionally, chrysin has been shown to have antidiabetic effects, reducing blood glucose levels and improving insulin sensitivity. Its diverse range of potential therapeutic applications has made chrysin an area of interest in the field of natural medicine and drug discovery (Satyanarayana et al., 2015; Shooshtari et al., 2020).

Alzheimer's disease (AD) is a complex neurodegenerative disease that is attributed to a combination of multiple factors, *e.g.*, synaptic dysfunctions such as synapses loss, deficits in synaptic plasticity,

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senile dementia, and progressive disability. These dysfunctions are thought to be extremely associated with cognitive decline in Alzheimer's disease patient. The disease leads to disruption of processes vital to neurons and their networks, including communication, metabolism, and repair. It has been observed that patients with AD typically lose neurons and connections in memoryrelated parts of the brain, including the entorhinal cortex and hippocampus (Sun et al., 2017; Gezici and Sekeroglu, 2022; Ju and Tam, 2022). Alzheimer's disease is characterized by changes in the brain that result in the loss of neurons and their connections, including the development of amyloid plaques and neurofibrillary, or tau, tangles. These changes affect a person's ability to remember and think and, eventually, to live independently. The main causes of AD and other neurodegenerative diseases are aggregated protein accumulation and oxidative damage (Calderon-Garcidueñas and Duyckaerts, 2018; Espay et al., 2019). Secondary metabolites of plants, such as terpenoids and flavonoids, have been shown to possess biological activities that can be useful in the prevention and treatment of AD. Terpenoids have been found to possess antioxidant activity and the potential to increase the level of acetylcholine, which is important for cognitive function. Flavonoids have been found to inhibit acetylcholinesterase, butyrylcholinesterase, Tau protein aggregation, β -secretase, oxidative stress, inflammation, and apoptosis through modulation of signaling pathways implicated in cognitive and neuroprotective functions. Additionally, natural compounds found in various parts of the medicinal plants and/or marine sources may potentially protect against neurodegeneration alongside improve memory and cognitive function. Even though, a numerous studies have been performed toreveal biological activities and pharmacological properties of chrysin, network-based molecular and pharmacological activities of chrysin have not been performed until now (Akram and Nawaz, 2017; Gezici and Sekeroglu, 2019; Sekeroglu and Gezici, 2019; Singh et al., 2021; Choudhir et al., 2022; Wu et al., 2022). Therefore, we aimed to identify the molecular targets and potential interactions of chrysin against AD by gene-set enrichment and bioinformatics approach. This research could provide a novel approach to uncover the therapeutic mechanisms of chrysin against AD.

2. Materials and Methods

2.1 Chemical compositions and predicted targets

Chemical Entities of Biological Interest (ChEBI) database, a part of ELIXIR Core Data Resources, was used for dictionary of molecular entities and chemical properties of chrysin (Hastings *et al.*, 2016). The targets of chrysin were identified using DIGEP-Pred (Prediction of drug-induced changes of gene expression profile) based on structural formula of chrysin (Lagunin *et al.*, 2013).

2.2 Pharmacokinetic properties and drug likeness analysis

PubChem database was used to obtain chemical structure and pharmacological properties of chrysin, as well as Chemical Entities of Biological Interest (ChEBI) database. Swiss ADME and ProToxII were used to determine drug likeness possibilities and toxicity properties of chrysin, respectively (Daina et al., 2017; Banerjee et al., 2018).

2.3 Prediction of targets by gene set enrichment analysis

GeneCards, The Human Gene Database, was used to evaluate probable interacting genes of chrysin. Based on this database, top interacting genes were analyzed using unique GeneCards identifiers (GC ids), provided by the GeneLoc Algorithm (Harel *et al.*, 2009; Fishilevich *et al.*, 2016). DisGeNET (version 7.0) database and the pharmacogenomics knowledge base (Pharm GKB) were employed to reveal the data about disease associated genes and variants from multiple sources (Thorn *et al.*, 2013; Pinero *et al.*, 2020). Additionally, SMILES into Swiss Target Prediction, a network-based tool for target prediction of bioactive molecules that were used to predict all the chrysin-related targets (Daina *et al.*, 2019).

2.4 Construction protein-protein interaction (PPI) network

STRING database was used to annotate the role of probable interacting genes and proteins associated with chrysin. PPI network mapping was conducted on chrysin and protein targets using the retrieval of interacting genes database with the species limited to "homo sapiens" and a confidence score ≥ 0.4 (Wu *et al.*, 2009; Athanasios *et al.*, 2017).

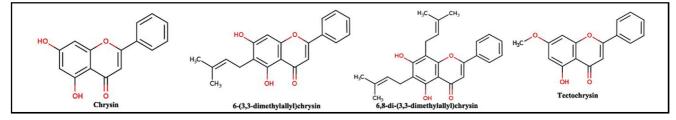
2.5 KEGG enrichment analysis

KEGG (Kyoto Encyclopedia of Genes and Genomes) is an integrated database of genes and genomes used for mapping pathways at molecular level. KEGG enrichment analysis was performed for construction the network regulated by chrysin (Aoki-Kinoshita and Kanehisa, 2007; Kanehisa *et al.*, 2017).

3. Results

3.1 Results of chemical compositions and predicted targets

Chrysin $(C_{15}H_{10}O_4)$ belonging to the class of flavone, includes a dihydroxyflavone in which the two hydroxy groups are located at positions C-5 and C-7. Similar to other flavones, chrysin is a yellow crystalline secondary compound soluble in water and ethanol. It is mainly found in the passion flowers of Passiflora incarnata and P. caerulea. The synonyms of chrysin with an average molecular mass of 254.237 g/molare chrysin, 5,7-dihydroxy-2-phenyl-4H-1benzopyran-4-one, 5,7-dihydroxy-2-phenyl-4H-benzo(b)pyran-4one, 5,7-dihydroxy-2-phenylchromen-4-one, and 5,7-dihydroxy flavone. 3-O-methyl-8 prenylgalangin (C₂₁H₂₀O₅), 6-(3,3dimethylallyl)chrysin (C₂₀H₁₈O₄), 6-geranylchrysin (C₂₅H₂₆O₄), 6,8di-(3,3-dimethylallyl) chrysin (C₂₅H₂₆O₄), 8-(3,3- dimethylallyl) chrysin ($C_{20}H_{18}O_4$), 8-geranylchrysin ($C_{25}H_{26}O_4$), chrysin 5-xyloside $(C_{20}H_{18}O_8, \text{ chrysin 5,7-dimethyl ether } (C_{17}H_{14}O_4), \text{ chrysin 7-}$ [rhamnosyl-(1->4)-glucoside $(C_{27}H_{30}O_{13})$, chrysin 7-4"acetylglucoside ($C_{23}H_{22}O_{10}$), chrysin 7-glucoronide ($C_{21}H_{18}O_{10}$), chrysin-7-O-glucoronide (C₂₁H₁₈O₁₀), chrysin-7-O-glucuronide $(C_{21}H_{18}O_{10})$, dimethylstrobochrysin $(C_{18}H_{16}O_4)$, helichrysin $(C_{22}H_{24}O_{10})$, rheochrysin ($C_{22}H_{22}O_{10}$), strobochrysin ($C_{16}H_{12}O_{4}$), and tectochrysin $(C_{16}H_{12}O_{4})$ are derivatives of chrysin. The chemical structure of chrysin and its derivatives were presented in the Figure 1.





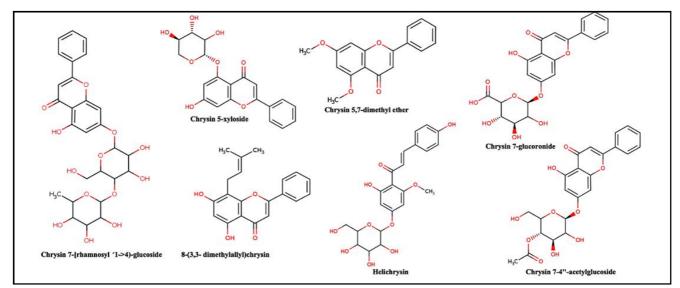


Figure 1: Chemical compositions of chrysin and some derivatives of chrysin.

The targets of chrysin were determined based on prediction of druginduced changes of gene expression profile for proteins at the pharmacological activity (Pa)>0.7. The findings were given in the Table 1 in which Pa (probability to be active) means the chance that chrysin, whereas Pi (probability to be inactive) means the chance that chrysin is belonging to the subclass of inactive compounds. According to the data presented in the table, chrysin has quite active biological activities including myosin-light-chain kinase inhibitor, anti-inflammatory, antineoplastic, antimutagenic, vasoprotector, and hepatoprotective. Actually, chlordecone reductase inhibitor, membrane integrity agonist, membrane permeability inhibitor, and kinase inhibitor were defined as the most valuable properties of chrysin (Pa>0.7).

Table 1: Prediction of drug-induced changes of gene expression profile for chrysin at pharmacological activity

Pa	Pi	Activity
0,967	0,002	Chlordecone reductase inhibitor
0,965	0,003	Membrane integrity agonist
0,962	0,003	HIF1A expression inhibitor
0,946	0,002	Membrane permeability inhibitor
0,944	0,002	2-Dehydropantoate 2-reductase inhibitor
0,942	0,002	Kinase inhibitor
0,942	0,005	CYP2C12 substrate
0,938	0,001	Aryl-alcohol dehydrogenase (NADP+) inhibitor
0,934	0,003	Anaphylatoxin receptor antagonist
0,933	0,003	Aldehyde oxidase inhibitor
0,928	0,001	P-benzoquinone reductase (NADPH) inhibitor
0,922	0,002	Peroxidase inhibitor
0,916	0,002	Histidine kinase inhibitor
0,915	0,002	Antimutagenic
0,913	0,001	Quercetin 2,3-dioxygenase inhibitor
0,906	0,002	CYP1A inducer
0,906	0,002	NADPH-ferrihemoprotein reductase inhibitor
0,902	0,005	TP53 expression enhancer
0,896	0,003	HMOX1 expression enhancer

0,894	0,003	Vasoprotector
0,896	0,006	Ubiquinol-cytochrome-c reductase inhibitor
0,890	0,002	Alcohol dehydrogenase (NADP+) inhibitor
0,885	0,001	Glycerol dehydrogenase (NADP+) inhibitor
0,885	0,003	27-Hydroxycholesterol 7alpha-monooxy-
0.005	0.005	genase inhibitor
0,885	0,005	CYP1A substrate
0,882	0,003	Cholesterol 26-monooxygenase inhibitor
0,877	0,002	CYP1A1 inducer
0,877	0,002	Beta-carotene 15,15'-monooxygenase inhibitor
0,879	0,006	Antiseborrheic
0,875	0,003	4-Nitrophenol 2-monooxygenase inhibitor
0,874	0,003	UGT1A9 substrate
0,883	0,012	Aspulvinonedimethylallyl transferase inhibitor
0,871	0,001	2-Dehydropantolactone reductase
		(A-specific) inhibitor
0,872	0,004	CYP1A1 substrate
0,863	0,002	2-Enoate reductase inhibitor
0,853	0,004	UGT1A6 substrate
0,846	0,002	Cystathionine beta-synthase inhibitor
0,842	0,003	MAP kinase stimulant
0,842	0,005	Apoptosis agonist
0,836	0,002	Antihemorrhagic
0,821	0,005	Alkane 1-monooxygenase inhibitor
0,815	0,003	AR expression inhibitor
0,808	0,003	UGT1A3 substrate
0,806	0,004	APOA1 expression enhancer
0,803	0,005	UGT1A substrate
0,799	0,004	CYP1A inhibitor
0,798	0,004	CYP2B5 substrate
0,793	0,003	Monophenol monooxygenase inhibitor
0,795	0,009	JAK2 expression inhibitor

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0,793	0,007	UDP-glucuronosyltransferase substrate
0,787	0,002	NADPH oxidase inhibitor
0,788	0,004	Histamine release inhibitor
0,786	0,003	Leukotriene-B4 20-monooxygenase inhibitor
0,799	0,019	Mucomembranous protector
0,774	0,002	1-Alkylglycerophosphocholine
		O-acetyltransferase inhibitor
0,779	0,008	CYP3A4 inducer
0,774	0,004	CYP1A2 inhibitor
0,773	0,004	UGT1A1 substrate
0,778	0,012	Dehydro-L-gulonate decarboxylase inhibitor
0,767	0,003	UGT1A7 substrate
0,766	0,004	MMP9 expression inhibitor
0,760	0,004	Tetrahydroxynaphthalene reductase inhibitor
0,761	0,006	CYP1A2 substrate
0,758	0,003	Beta glucuronidase inhibitor
0,759	0,004	CYP2A4 substrate
0,756	0,002	CYP1A1 inhibitor
0,769	0,016	Antineoplastic
0,757	0,004	Pectate lyase inhibitor
0,776	0,027	CYP2J substrate
0,778	0,031	Testosterone 17beta-dehydrogenase (NADP+) inhibitor
0,746	0,002	CF transmembrane conductance regulator agonist
0,748	0,005	Pin1 inhibitor
0,746	0,004	Xenobiotic-transporting ATPase inhibitor
0,741	0,002	CYP19A1 expression inhibitor
0,747	0,009	CYP3A inducer
0,748	0,012	Glutathione thiolesterase inhibitor
0,755	0,021	CYP2J2 substrate
0,738	0,004	Nitrite reductase [NAD(P)H] inhibitor
0,735	0,002	Alcohol dehydrogenase [NAD(P)+] inhibitor
0,732	0,005	Insulysin inhibitor
0,727	0,003	UGT1A10 substrate
0,729	0,006	Ecdysone 20-monooxygenase inhibitor
0,734	0,015	Glucan endo-1,6-beta-glucosidase inhibitor
0,721	0,005	5 Hydroxytryptamine release inhibitor
0,719	0,009	CYP2A6 substrate
0,713	0,003	CYP1B substrate
0,711	0,002	GABA C receptor antagonist
0,707	0,002	Iodide peroxidase inhibitor
0,708	0,004	Antioxidant
0,706	0,003	UGT2B15 substrate
0,705	0,003	NOS2 expression inhibitor
0,710	0,011	NAD(P)+-arginine ADP-ribosyltrans
		ferase inhibitor
0,701	0,005	UGT2B12 substrate
0,704	0,010	Thioredoxin inhibitor
0,706	0,013	Nitrate reductase (cytochrome) inhibitor

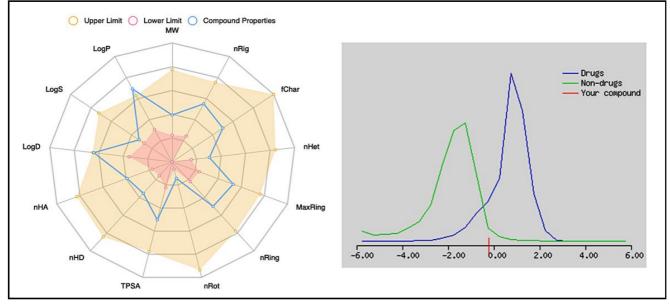
3.2 Results of pharmacokinetic properties and drug-likeness analysis

The chemical and molecular information of chrysin obtained from ChEBI and PubChem, and the relevant drug-likeness properties obtained from SwissADME were summarized in the Table 2. Swiss ADME predicted pharmacokinetic of the chrysin, including topological polar surface area and Lipinski's Rule of 5 recommendations were shown in the Table 2.

of chrysin			
ID	ChEBI: 75095		
Name	Chrysin		
Synonyms	Chrysine, 5,7-Dihydroxy-2-phenyl-4H-		
	1-benzopyran-4-one, 5,7-Dihydroxy-2-		
	phenyl-4H-benzo(b)pyran-4-one, 5,7-		
	dihydroxy-2-phenylchromen-4-one, and 5,7-Dihydroxyflavone		
Formula	$C_{15}H_{10}O_4$		
Net Charge	0 = 0		
MW	254.23750 g/mol		
Monoisotopic Mass	254.05791 g/mol		
Hdon	2		
Hacc	4		
Rbon	1		
TPSA (+! ²)	70.67		
Melting/boiling point	285.5 °C / 491.91 °C at 760.00 mm Hg		
	(est)		
W S	0.105 mg/ml		
InChI	InChI = 1S/C15H10O4/c16-10-6-		
	11(17)15-12(18)8-13(19-14(15)7-10)0421250(118)8-13(19-14(15)7-10)0421250(118)16(17)100000000000000000000000000000000000		
SMILES	10)9-4-2-1-3-5-9/h1-8,16-17H		
Canonical SMILES	Ocl cc(O)c2c(cl)oc(cc2=O)-cl cccccl C l = C C = C (C = C l) C 2 = C C (
Canonical SWILLS	=0)C3=C(C=C(C=C3O2)O)O		
DL score	-0.21		
Lipinski	Yes		
Bioavailability score	0.55		
Glabsorpsion	High		
Coco-2 Permeability	-4.69		
BBB permeant	Yes		
T ₁ / ₂	0.787		
CL	5.131		
MolLogP	3.67		
MolLogS	-3.51 (in Log(moles/l)) 79.10 (in mg/l)		
MolPSA	55.95 A ²		
Log K _p (cm/s)	-5.35		
Top chemical roles	Radical scavenger and antioxidant		
Ton biological roles	activity Plant metabolite, myosin-light-chain		
Top biological roles	kinase inhibitor, anti-inflammatory		
	agent, anti-neoplastic agent, and		
	hepatoprotective agent		

Table 2:	Pharmacological	properties	and	drug-likeness	results
	of chrysin				

BBB = blood-brain barrier, DL = drug-likeness, WS = water solubility, GI = gastrointestinal absorption, Hacc = hydrogen bond acceptors, Hdon = hydrogen bond donors, MW = molecular weight, Rbon = rotatable bonds, TPSA = topological polar surface area, CL = clearance. As shown in the Figure 2, chrysin predicted to possess a good druglikeness activity with the score of -0.21, as well as good brain barrier permeability (BBB score = 3.71). In addition, LogP, one of the important components of Lipinski's Rule of 5, was determined as 3.67 that means chrysin can be as an oral drug. *In silico* druglikeness possibilities of chrysin are given in the Figure 2.





In silico toxicological parameters of chrysin were evaluated using ProTox-II software, and the results are presented in Table 3. Oral toxicity prediction results were determined as LD_{50} (lethal dose₅₀) values in 3919 mg/kg body weight and the predicted toxicity class of chrysin was 5 according to the globally harmonized system of classification of labelling of chemicals. Furthermore,

the results revealed that chrysin showed no observable toxicity, including carcinogenicity, cytotoxicity, hepatotoxicity, immunotoxicity, and mutagenicity. Table 3 also showed that chrysin is a component involved in Tox21-nuclear receptor signaling and Tox21-stress response pathways targeting various ligands.

Table 3:	Toxicity	prediction	report	including	targets (of chrysin
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Classification	Target	Prediction	Probability
Organ toxicity	Hepatotoxicity	Inactive	0.68
Toxicity end points	Carcinogenicity	Inactive	0.62
Toxicity end points	Immunotoxicity	Inactive	0.99
Toxicity end points	Mutagenicity	Inactive	0.57
Toxicity end points	Cytotoxicity	Inactive	0.87
Tox21-Nuclear receptor signaling pathways	Aryl hydrocarbon receptor (AhR)	Active	1.0
Tox21-Nuclear receptor signaling pathways	Androgen Receptor (AR)	Inactive	0.99
Tox21-Nuclear receptor signaling pathways	Androgen Receptor Ligand Binding Domain (AR-LBD)	Inactive	1.0
Tox21-Nuclear receptor signaling pathways	Aromatase	Active	0.61
Tox21-Nuclear receptor signaling pathways	Estrogen Receptor Alpha (ER)	Active	1.0
Tox21-Nuclear receptor signaling pathways	Estrogen Receptor Ligand Binding Domain (ER-LBD)	Active	1.0
Tox21-Nuclear receptor signaling pathways	Peroxisome Proliferator Activated Receptor Gamma (PPAR-Gamma)	Active	1.0
Tox21-Stress response pathways	Nuclear factor (erythroid-derived 2)-like 2/antioxidant responsive element (nrf2/ARE)	Inactive	0.99
Tox21-Stress response pathways	Heat shock factor response element (HSE)	Inactive	0.99
Tox21-Stress response pathways	Mitochondrial Membrane Potential (MMP)	Active	1.0
Tox21-Stress response pathways	Phosphoprotein (Tumor Suppressor) p53	Active	1.0
Tox21-Stress response pathways	ATPase family AAA domain-containing protein 5 (ATAD5)	Active	0.96

3.3 Results of top gene enrichment analysis

In this research, the gene targets of chrysin against AD were collected using GeneCards, DisGeNET, PharmGKB, and SMILES into SwissTargetPrediction. Based on the results, 133 targets were collected to determine the chrysin-associated targets, and 2542 targets were collected Alzheimer disease-related targets. Accordingly, a total of thirty-eight genes were identified as the intersection targets. The detailed information about the targets of chrysin against AD was shown in Table 4.

Table 4: The list of top genes interacts with chrysin against AD

Gene Symbol	Unipot Gene Id	Protein Description
ABCB1	P08183	ATP-dependent translocase
ABCC1	P33527	Multidrug resistance-associated protein-1
ABCG2	Q9UNQ0	ATP-binding cassette transporter
ACHE	P22303	Acetylcholinesterase
AKT1	P31749	AKT serine/threonine kinase 1
APP	P05067	Amyloid-beta precursor protein
AR	P10275	Androgen receptor
BAX	Q07812	Apoptosis regulator BAX
BCHE	P06276	Butyrylcholinesterase
BCL2	P10415	Apoptosis regulator Bcl-2
BDNF	P23560	Brain-derived neurotrophic factor
CASP3	P42574	Caspase 3
CLU	P10909	Clusterin
ECE1	P42892	Endothelin converting enzyme 1
EGFR	P00533	Epidermal growth factor receptor
ESR1	P03372	Estrogen receptor 1
ESR2	Q92731	Estrogen receptor 2
GSK3B	P49841	Glycogen synthase kinase 3 beta
HMOX1	P09601	Heme oxygenase 1
IGF1	P05019	Insulin-like growth factor
IL17A	Q16552	Interleukin 17A
IL1B	P01584	Interleukin 1 beta
IL6	P05231	Interleukin 6
MAOA	Q5ULA9	Monoamine oxidase A
MAPK1	P28482	Mitogen-activated protein kinase
MET	P08581	MET proto-oncogene, receptor tyrosine kinase
MPO	P05164	Myeloperoxidase
NFKB1	P19838	Nuclear factor kappa B subunit 1
NOS3	P29474	Nitric oxide synthase, endothelial
PLAU	Q03405	Plasminogen activator, urinary
PLD2	O14939	Phospholipase D2
PPARG	P37231	Peroxisome proliferator activated receptor gamma
PSEN1	P49768	Presenilin 2
PSEN2	P49810	Presenilin 2
PTGS2	P35354	Prostaglandin G/H synthase 2
TNF	P01375	Tumor necrose factor
T P 5 3	P04637	Tumor protein p53
VEGFA	P15692	Vascular endothelial growth factor A

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3.4 Results of protein – protein interaction network

The relationship of a total of 38 proteins between each other were constructed from STRING database with PPI enrichment p-value < 1.0e-16 (FDR < 0.05). This enrichment value means that these

proteins have more interactions among themselves than what would be expected for a random set of proteins of the same size and degree distribution drawn from the genome. PPI network was presented in the Figure.

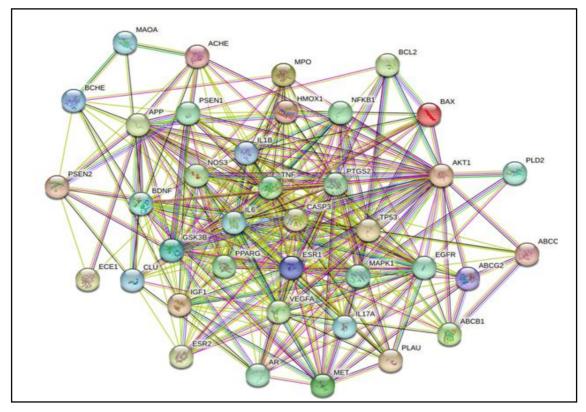


Figure 3: Protein-protein interaction networks of chrysin against AD-related targets.

As can be seen in the Figure, CASP, ESR1, MAPK1, NOS3, IL1B, VEGFA, and PTGS2 are the proteins that located in the center of the network. ILB, IL6, TNF, MAPK1, CASP3, PSEN1, PSEN2, PTGS2,

NFKB1, AKT1, GSK3B, and APP were selected as top core targets that may play a significant role in AD treatment.

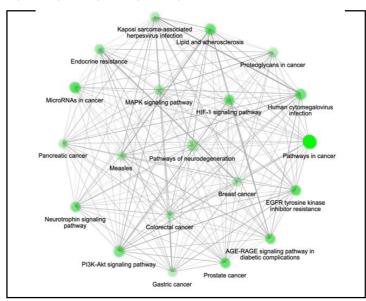


Figure 4: Network of top related pathways construction with KEGG enrichment.

3.5 Results of KEGG enrichment pathway

According to the KEGG enrichment pathway analyses, a total of 158 distinct pathways were identified as the probably modulated pathways by chrysin. A network corresponding to 38 protein targets are schematized in Figure 4, summarizing the correlations between the major pathways listed in the enrichment network. As presented in the Figure 4, several target proteins are simultaneously involved in one pathway, while one target protein is also present in many pathways.

Accordingly, pathways in cancer, microRNAs in cancer, lipid and atherosclerosis, hypoxia-inducible factor 1 (HIF-1) signaling pathway, PI3K-Akt signaling pathway, mitogen-activated protein kinase (MAPK) signaling pathway, epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor resistance, advanced glycation endproductsreceptor for advanced glycation endproducts (AGE-RAGE) signaling in diabetic complications, neurotrophin signaling pathway, sphingolipid signaling pathway, interleukin-17 (IL-17) signaling pathway, neurodegeneration, and Alzheimer's disease were determined as the top pathways associated with chrysin-regulated proteins with the lowest false discovery rate (FDR<0.05) (Figure 4).

4. Discussion

Network-based pharmacology is a promising approach for identifying effective phytochemicals against various diseases, including Alzheimer disease. This approach involves exploring the biological function, protein-phytochemical/drugs network, and up-down regulation of pathological host target proteins to understand the mechanism of phytochemicals against the disease. Network pharmacology combines phytochemical information with bioinformatics tools to illustrate herbal formulae holistically in the context of phytochemical basis and therapeutic mechanisms. Studies have used network pharmacology to investigate significant phytochemicals, pathways, and targets against AD and to explore the molecular evidence of herbal formulae. Accordingly, this approach can be used to identify potential targets and efficacy prediction to uncover the therapeutic effect of herbal drugs (Boezio et al., 2017; Dragomir et al., 2018). Although, biological effects and pharma-cological profiling of chrysin have been studied, network-based genomic targets of chrysin, molecular signaling pathways and pharmacokinetic properties of chrysin have not been revealed until now (Akram and Nawaz, 2017; Gezici and Sekeroglu, 2019; Sekeroglu and Gezici, 2019; Singh et al., 2021; Choudhir et al., 2022; Wu et al., 2022). In this context, this research was aimed to investigate the molecular targets and potential interactions of chrysin against AD by gene-set enrichment and bioinformatics approach.

In the current research, we first identified the pharmacokinetic properties and toxicity of chrysin via electronic databases. Chrysin compliance with Lipinski rule of 5, and with a higher LD_{50} value. The literature has previously reported oral administration of chrysin (5000 mg/kg) showed 40% mortalityin acute oral toxicity, while daily oral administration of chrysin (1000 mg/kg) showed significantly decreased body weight and significantly increased liver weight in male rats in the sub-chronic toxicity study (Yao *et al.*, 2023), as consistent with the estimated results of this work. In other research aimed to investigate toxicity of chrysin by acute and sub-chronic oral toxicity in rats, LD_{50} value estimated for the chrysin was 4350 mg/kg, in fact, chrysin caused 40% of dead's in both male and female rats at the dose of 5000 mg/kg body weight (Falbo and

Aiello, 2023), which is close to the results of this bioinformatic research.

Recent studies have focused on the role of chrysin in targeting specific genes involved in Alzheimer's disease, such as the amyloid-beta precursor protein (APP). Chrysin has been found to protect against the accumulation of beta-amyloid plaques in the brain, which are a hallmark of Alzheimer's disease. Additionally, chrysin has been shown to have anti-inflammatory and antioxidant properties that may help mitigate the effects of neurodegeneration (Calderon-Garcidueñas and Duyckaerts, 2018; Ju and Tam, 2022). Therefore, chrysin-associated targets and Alzheimer disease-related targets were determined in the present research. This research showed that chrysin regulates the activities of genes related to AD in a way that causes an increase in the expression levels of some of the genes, e.g., antioxidant molecules, while it causes a decrease in others, e.g., IL-1B, IL-10 and TNF- α . Further research is needed to fully understand the potential benefits of chrysin in treating Alzheimer's disease, but early studies show promise for this natural compound as a potential therapeutic option. Liu et al. (2021) investigated the "hidden" multi-target strategy of novel chrysinderivatives in combination with its molecular targets for the treatment of Alzheimer's disease. Amongst the synthesis derivatives, compound 3 was found as a potential hidden multifunctional candidate in the therapy of AD, thanks to its good ADMET (absorption, distribution, metabolism, excretion and toxicity) score (Liu et al., 2021). Nonetheless, a recent meta-analysis of genome-wide association studies on Alzheimer's disease and related dementias identified new loci and enabled the generation of a new genetic risk score associated with the risk of Alzheimer's disease (Bellenguez et al., 2022).

Afterwards, PPI analyses of targeting proteins were conducted and CASP, ESR1, MAPK1, NOS3, IL1B, VEGFA, and PTGS2 are identified as the core proteins that located in the center of the network.In addition, target signaling pathways modulated by chrysin were determined using KEGG enrichment in this research. It is well-known that multiple signaling pathways interact with each other in the metabolic processes in living organisms. Based on the pathway analyses, most of the genes regulated by chrysin are found to closely associate with cancer, microRNAs in cancer, HIF-1 signaling pathway, PI3K-Akt signaling pathway, MAPK signaling pathway, EGFR tyrosine kinase inhibitor resistance, AGE-RAGE signaling in diabetic complications, neurotrophin signaling pathway, sphingolipid signaling pathway, IL-17 signaling pathway, neurodegeneration, and Alzheimer's disease. ILB, IL6, TNF, MAPK1, CASP3, PSEN1, PSEN2, PTGS2, NFKB1, AKT1, GSK3B, and APP genes are involved in Alzheimer's disease. In agreement with the findings from this network-based research, previous reports indicated that chrysin is used to synaptic dysfunctions such as synapses loss, deficits in synaptic plasticity, senile dementia, and progressive disability, which are closely associated with the development of AD (Akram and Nawaz, 2017; Calderon-Garcidueñas and Duyckaerts, 2018; Espay et al., 2019; Ju and Tam, 2022; Wu et al., 2022).

5. Conclusion

Alzheimer's disease (AD) is a complex neurodegenerative disease that is attributed to a combination of multiple factors, including synaptic dysfunctions, deficits in synaptic plasticity, senile dementia, and progressive disability. It is characterized by changes in the brain that result in the loss of neurons and their connections, including the development of amyloid plaques and neurofibrillary, or tau, tangles. The main causes of AD and other neurodegenerative diseases are aggregated protein accumulation and oxidative damage. Phytochemicals such as terpenoids and flavonoids, have been found to possess biological activities. In particularly, have been found to possess antioxidant activity and the potential to increase the level of acetylcholine, while flavonoids have been found to inhibit acetylcholinesterase, butyrylcholinesterase, Tau protein aggregation, β-secretase, oxidative stress, inflammation, and Alzheimer's disease. Chrysin is a natural flavonoid that possess remarkable biological and pharmacological activities in the human metabolism. The results showed that the network-based techniques could provide a new approach to discover the mechanisms of chrysin forthe prevention and treatment of AD. Even though it is rare to use chrysin alone as a medicine nowadays, it is expected that chrysin will be most likely to be used in the future drug discovery. Further studies, especially clinical trials are necessary to confirm the pharmacokinetic properties and molecular targets of chrysin against AD.

Conflict of interest

The authors declare no conflict of interest relevant to this article.

References

- Akram, M. And Nawaz, A. (2017). Effects of medicinalplants on Alzheimer's disease and memory deficits. Neural Regeneration Research, 12(4):660.
- Aoki-Kinoshita, K. F. and Kanchisa, M. (2007). Gene annotation and pathway mapping in KEGG. In Comparative genomics pp:71-91. Humana Press.
- Athanasios, A.; Charalampos, V. and Vasileios, T. (2017). Protein-protein interaction (PPI) network: Recent advances in drug discovery. Current Drug Metabolism, 18(1):5-10.
- Banerjee, P.; Eckert, A. O.; Schrey, A. K. and Preissner, R. (2018). ProTox-II: A webserver for the prediction of toxicity of chemicals. Nucleic Acids Research, 46(W1):W257-W263.
- Bellenguez, C.; Küçükali, F.; Jansen, I. E.; Kleineidam, L.; Moreno-Grau, S.; Amin, and Goldhardt, O. (2022). New in sights into the geneticetiology of Alzheimer's disease and relateddementias. Nature Genetics, 54(4):412-436.
- Boezio, B.; Audouze, K.; Ducrot, P. And Taboureau, O. (2017). Network basedapproaches in pharmacology. Molecularinformatics, 36(10): 1700048.
- Calderon-Garcidueñas, A. L. and Duyckaerts, C. (2018). Alzheimer disease. Handbook of Clinical Neurology, 145:325-337.
- Choudhir, G; Sharma, S. and Hariprasad, P. (2022). A combina-torial approach to screen structurally diverse acetylcholinesterase inhibitory plant secondary metabolites targeting Alzheimer's disease. Journal of Biomolecular Structure and Dynamics, 40(22):11705-11718.
- Daina, A.; Michielin, O. and Zoete, V. (2017). Swiss ADME: A free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. Scientific Reports, 7(1):42717.
- Daina, A.; Michielin, O. and Zoete, V. (2019). Swiss Target Prediction: Updated data and new features for efficient prediction of protein targets of small molecules. Nucleic Acids Research, 47(W1): W357-W364.
- Dragomir, A.; Vrahatis, A. G. and Bezerianos, A. (2018). A networkbasedperspective in Alzheimer's disease: Currentstate and an integrative framework. IEEE Journal of Biomedical and Healthin Formatics, 23(1):14-25.

- Espay, A. J.; Vizcarra, J. A.; Marsili, L.; Lang, A. E.; Simon, D. K.; Merola, A. and Leverenz, J. B. (2019). Revisiting protein aggregation as pathogenic in sporadic Parkinson and Alzheimer diseases. Neurology, 92(7):329-337.
- Falbo, F. and Aiello, F. (2023). Chrysin: A polyedricflavone as a tool to explore new phytotherapeutic applications and drug design. Archiv der Pharmazie, 356(2):2200347.
- Fishilevich, S.; Zimmerman, S.; Kohn, A.; Iny Stein, T.; Olender, T.; Kolker, E. and Lancet, D. (2016). Genic insights from integrated human proteomics in Gene Cards. Database, pp:16.
- Gezici, S. and Sekeroglu N. (2022). Medicinal plants and phytochemicals of potential importance in the developmental process and treatment of alzheimer's disease. Hacettepe University Journal of the Faculty of Pharmacy, 42(2):121-133.
- Gezici, S. and Sekeroglu, N. (2019). Neuroprotectivepotential and phytochemicalcomposition of acornfruits. Industrial Crops and Products, 128:13-17.
- Harel, A.; Inger, A.; Stelzer, G; Strichman-Almashanu, L.; Dalah, I.; Safran, M. and Lancet, D. (2009). GIFtS: Annotation landscape analysis with GeneCards. BMC Bioinformatics, 10(1):1-11.
- Hastings, J.; Owen, G; Dekker, A.; Ennis, M.; Kale, N.; Muthukrishnan, V. and Steinbeck, C. (2016). ChEBI in 2016: Improved services and an expanding collection of metabolites. Nucleic Acids Research, 44(D1):D1214-D1219.
- Ju, Y. and Tam, K. Y. (2022). Pathological mechanisms and therapeutic strategies for Alzheimer's disease. Neural Regeneration Research, 17(3):543.
- Kanehisa, M.; Furumichi, M.; Tanabe, M.; Sato, Y. and Morishima, K. (2017). KEGG: new perspectives on genomes, pathways, diseases and drugs. Nucleic Acids Research, 45(D1):D353-D361.
- Kasala, E. R.; Bodduluru, L. N.; Madana, R. M.; Gogoi, R. and Barua, C. C. (2015). Chemopreventive and therapeutic potential of chrysin in cancer: Mechanistic perspectives. Toxicology Letters, 233(2):214-225.
- Khoo, B. Y., Chua, S. L. and Balaram, P. (2010). Apoptotic effects of chrysin in human cancer cell lines. International Journal of Molecular Sciences, 11(5):2188-2199.
- Lagunin, A.; Ivanov, S.; Rudik, A.; Filimonov, D. and Poroikov V. DIGEP-Pred (2013). Web service for *in silico* prediction of drug-induced gene expression profiles based on structural formula. Bioinformatics, 29(16):2062e3.
- Lee, H. S.; Lee, I. H.; Kang, K.; Park, S. L; Kwon, T. W. and Lee, D. Y. (2020). An investigation of the molecular mechanisms underlying the analgesic effect of Jakyak-Gamcho Decoction: A Network Pharmacology Study. Evidence-Based Complementary and Alternative Medicine, pp:20.
- Li, Y.; Yang, R.; Huang, X.; Chen, C.; Dou, D.; Wang, Q. and Sun, T. (2022). Chrysin targets myeloid-derived suppressor cells and enhances tumour response to anti-PD-1 immunotherapy. Clinical and Translational Medicine, 12(9).
- Liu, C.; Kou, X.; Wang, X.; Wu, J.; Yang, A. and Shen, R. (2021). Novelchrys in derivatives as hidden multifunctional agents for anti-alzheimer's disease: Design, synthesis and *in vitro* evaluation. European Journal of Pharmaceutical Sciences, 166:105976.
- Mani, R. and Natesan, V. (2018). Chrysin: Sources, beneficial pharmacological activities, and molecular mechanism of action. Phytochemistry, 145:187-196.
- Pinero, J.; Ramírez-Anguita, J. M.; Saüch-Pitarch, J.; Ronzano, F.; Centeno, E.; Sanz, F. and Furlong, L. I. (2020). The DisGeNET knowledge platform for disease genomics: 2019 update. Nucleic Acids Research, 48(D1):D845-D855.

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- Satyanarayana, K.; Sravanthi, K.; Shaker, I. A.; Ponnulakshmi, R. and Selvaraj, J. (2015). Role of chrysin on expression of insulin signaling molecules. Journal of Ayurveda and Integrative Medicine, 6(4):248.
- Sekeroglu, N. and Gezici, S. (2019). Astragalus neuro carpus Bioss. as a potentialsource of natural enzyme inhibitor associated with Alzheimer's and Parkinson diseases along with its rich polyphenolic content and antioxidant activities. Ann. Phytomed., 8(1):82-87.
- Shooshtari, M. K.; Sarkaki, A.; Mansouri, S. M. T.; Badavi, M.; Khorsandi, L.; Ghasemi Dehcheshmeh, M. and Farbood, Y. (2020). Protective effects of chrysin against memory impairment, cerebral hyperemia and oxidative stress after cerebral hypoperfusion and reperfusion in rats. Metabolic Brain Disease, 35:401-412.
- Singh, A. K.; Rai, S. N.; Maurya, A.; Mishra, G.; Awasthi, R.; Shakya, A. and Singh, M. P. (2021). Therapeuticpotential of phytoconstituents in management of Alzheimer's disease. Evidence-Based Complementary and Alternative Medicine, 21:1-19.
- Stompor-Gor'cy, M.; Bajek-Bil, A. and Machaczka, M. (2021). Chrysin: Perspectives on contemporary status and future possibilities as pro-health agent. Nutrients, 13(6):2038.

- Sun, Q.; Xie, N.; Tang, B.; Li, R. and Shen, Y. (2017). Alzheimer's disease: from genetic variants to the distinct pathological mechanisms. Frontiers in Molecular Neuroscience, 10:319.
- Thorn, C. F.; Klein, T. E. and Altman, R. B. (2013). PharmGKB: The pharmacogenomics knowledge base. Pharmacogenomics: Methods and Protocols, pp:311-320.
- Wu, J.; Vallenius, T.; Ovaska, K.; Westermarck, J.; Mäkelä, T. P. and Hautaniemi, S. (2009). Integrated network analysis platform for protein-protein interactions. Nature methods, 6(1):75-77.
- Wu, X.; Zheng, X.; Tang, H.; Zhao, L.; He, C.; Zou, Y. and Ye, G (2022). A network pharmacology approach to identify the mechanisms and molecular targets of curcumin against Alzheimer disease. Medicine, 101(34).
- Yao, W.; Cheng, J.; Kandhare, A. D.; Mukherjee-Kandhare, A. A.; Bodhankar, S. L. and Lu, G (2021). Toxicological evaluation of a flavonoid, chrysin: morphological, behavioral, biochemical and histopathological assessments in rats. Drug and Chemical Toxicology, 44(6):601-612.
- Zeinali, M.; Rezaee, S. A. and Hosseinzadeh, H. (2017). An overview on immunoregulatory and anti-inflammatory properties of chrysin and flavonoids substances. Biomedicine and Pharmacotherapy, 92: 998-1009.

Citation Sevgi Gezici and Nazim Sekeroglu (2023). Bioinformatics approach to identify molecular targets of chrysin against Alzheimer's disease. Ann. Phytomed., 12(1):259-268. http://dx.doi.org/10.54085/ap.2023.12.1.38.