



Review Article : Open Access

A review on the importance of two medicinal plants of North East India: *Paris polyphylla* Smith and *Kaempheria parviflora* Wall. ex Baker

A.K. Phurailatpam*[◆], Anju Choudhury**[◆], Tasso Yatung*** and Kalkame Ch. Momin*

* Department of Floriculture, Medicinal and Aromatic Plants, College of Horticulture and Forestry, Central Agricultural University, Pasighat-791102, Arunachal Pradesh, India

** Department of Social Sciences, College of Horticulture and Forestry, Central Agricultural University, Pasighat-791102, Arunachal Pradesh, India

*** Department of Vegetable Sciences, College of Horticulture and Forestry, Central Agricultural University, Pasighat-791102, Arunachal Pradesh, India

Article Info

Article history

Received 8 August 2022

Revised 25 September 2022

Accepted 26 September 2022

Published Online 30 December-2022

Keywords

Medicinal plants

Paris polyphylla Smith*Kaempheria parviflora* Wall.

Ex Baker

Vulnerable

North East India

Abstract

Paris polyphylla Smith and *Kaempheria parviflora* Wall ex Baker are two important medicinal plants, found mainly in the north eastern parts of India. These rhizomatic plants are in great demands and extracted injudiciously from the wild. The rhizome is the economic part which is used for its various medicinal properties. The present article gives an account of updated information on its phytochemical, pharmacological properties, its ethnomedicinal uses and their conservation aspects in the region. The review reveals that a wide numbers of phytochemical constituents have been isolated from these plants. The rhizomes of the several species of the genus *Paris* have been used as haemostatic and anti-inflammatory agent to treat traumatic injuries, snake bites, abscess, parotitis and mastitis. The rhizome of *K. parviflora* has several uses in treating various ailments including gastrointestinal disorders, fungal infections, allergies and alleviate male impotence, antiallergic, anti-inflammatory, antifungal, antiplasmodial, antimycobacterial, anti HIV-1, antispasmodic effects, etc. For the last few decades or so, extensive research work has been done to prove its biological activities and pharmacology of its extracts. Excessive injudicious collection and harvesting from the wild has pushed these species towards extinction. Domestication, cultivation and strict laws are the need of the hour to save these species from extinction.

1. Introduction

The traditional knowledge of utilization of plants and herbs as medicine is passed on from generations till today. The use of plants and herbs for medicinal purposes has been documented from many tribal communities. Plant and plant based traditional system of healthcare has been practiced in many parts of the world and the knowledge is slowly getting lost in the modern society (Aziz *et al.*, 2016; Aziz *et al.*, 2017; Mahmood *et al.*, 2011). In India, the sacred Vedas give many references of medicinal plants used in our country for the last many centuries. Due to its fewer side effects as compared to allopathic drugs, there has been a tremendous increase in the demand of plant based health products in developing as well as developed countries, resulting in popularity of herbal products and medicine around the world. It is estimated that about 80% of the world population residing in the rural areas of the developing and under developed countries greatly rely mainly on medicinal plants and its extracts in the absence of affordable and accessible source of primary healthcare system (WHO, 2002). It is well known that

plants play a significant role as a raw material source for various AYUSH-medicine formulations (<https://main.ayush.gov.in>). A more practical, productive and easily accessible source of medicine are the common medicinal herbs (Husain, 2021). Use of traditional herbs regularly improves immunity, fights many ailments, rejuvenates the body and age associated disorders. They have the potential to combat chronic conditions such as hypertension, high cholesterol, diabetes, AIDS, influenza, a persistent cold or cough, infections, chronic fatigue, and inflammatory conditions (Ansari and Ahmad, 2019). It is believed that many people still treat themselves with traditional medicines made from plants, herbs and spices. The WHO, which launched the traditional medicine strategy (2014-2023) with the goal of maintaining global population health by prioritizing the use of traditional and complementary medicines, has also acknowledged the importance of herbal medicine. Scientists are now conducting research on a variety of spices and herbs to provide a rational explanation for their use in the treatment and prevention of common illnesses (Mehrotra, 2021). Traditional medicine system of Ayurveda, Siddha and Unani has been existed in our country for centuries and plants are mentioned as a medicine in ancient texts like the Rig veda and the Atharva veda (2000 BC) (Warrier, 2021).

Many modern allopathic drugs and compounds are extracted from plants and also the modern medicine and pharmaceutical companies rely on plants for many important and novel drugs. The development

Corresponding author: Arunkumar Phurailatpam

Associate Professor College of Horticulture and Forestry Central Agricultural University Pasighat-791102, Arunachal Pradesh

E-mail: arunkumarph77@gmail.com

Tel.: +91-9612975356

Copyright © 2022 Ukaaz Publications. All rights reserved.

Email: ukaaz@yahoo.com; Website: www.ukaazpublications.com

of biomedicine, which has achieved wonders in medicine, has been greatly supported by the enormous advancements made by investigative biochemistry, electronics and other scientific fields. But, it also resulted into extra ordinary shoot up in costs of these advance medicines and putting it away from the grasp of the most of the world's population (Sharma, 2021). Many plants and plant products are evaluated and analysed based on their traditional uses for the development or discovery of new and novel drugs and compounds. Continuously unsustainable and injudicious extractions of these plants from their natural habitat have pushed some of the important species towards extinction. With the increase in world population and urbanization a rapid loss in medicinal plant species and knowledge seems to be inevitable in the near future. Citing an example, *Taxus wallichiana* Zucc., an important medicinal plant has pushed towards extinction due to unsustainable extraction from the wild after the discovery of its efficacy in cancer treatment. It contains a very important chemical, paclitaxel (taxol), one of the most effective drugs used for a variety of cancers (Cragg and donal, 1993; Goldspiel, 1997). In recognition of its anticancerous properties during 1980s the species has suffered large scale unsustainable extraction, which led to its present status (Paul *et al.*, 2013). IUCN has documented this plant as one of the most threatened species (Thomas and Farjon, 2011). Presently, many plant species comes under threatened and endangered category due over harvesting and injudicious collection from the wild. In this review, two endangered medicinal plant species; namely, *P. polyphylla* and *K. parviflora* found in North eastern part of India is taken up. *P. polyphylla* Smith and *K. parviflora* are two important medicinal plants found mainly in the north eastern parts of India (Gajurel *et.al.*, 2015; Devi *et. al.*, 2016). These plants are economically very important for the region and have high demand but they are very less documented.

2. *Paris polyphylla* Smith

P. polyphylla is a herbaceous plant belonging to Melanthiaceae family. The plant is found in many parts of Asia and Europe (two species). In Asia, it is mainly found in Indo-China region and South East Asia region and 24 species with 12 endemic species are reported to be found in China alone. Two species; namely, *P. polyphylla* and *P. tibetica* with about 6 intraspecific taxa are found in India (Liang and Soukup, 2000).

Several species of Paris as per World checklist of selected plant families (WCSP) are given below (Govaerts, 2011; Liang *et al.*, 2012).

- *P. polyphylla* var. *alba*
- *P. polyphylla* var. *chinensis*
- *P. polyphylla* var. *latifolia*
- *P. polyphylla* var. *nana*
- *P. polyphylla* var. *panxiensis*
- *P. polyphylla* var. *polyphylla*
- *P. polyphylla* var. *stenophylla*
- *P. polyphylla* var. *yunnanensis*

Five additional varieties were also identified by The flora of China, three of which are placed in different species by the WCSP:

- *P. polyphylla* var. *appendiculata*=*P. tibetica*
- *P. polyphylla* var. *brachystemon*=*P. polyphylla* var. *stenophylla*
- *P. polyphylla* var. *kwantungensis*=*P. polyphylla* var. *polyphylla*
- *P. polyphylla* var. *minor*=*P. delavayi*
- *P. polyphylla* var. *pseudothibetica*=*P. delavayi*

P. polyphylla is an erect, perennial, herbaceous plant with unbranched shoot. The shoot contains a single stem-like structure and 2 to 3 whorls of leaves. The underground part is a rhizome with nodes. The flowering period starts in April and May last up to three months. It has odd flowers with long yellow radiating anthers (Shah *et al.*, 2012). The stems are smooth and 50-100 cm in height with a diameter of 1-2.5 cm in diameter. The leaves are simple and arranged in whorls; petiolate, lanceolate and reticulate with 3 primary veins. The flowers are solitary and hermaphrodite borne at the axial terminal. The flowers are yellowish green in colour and the outer sepaloid is larger than the inner with 3-5 tepals. The androecium is free containing 6-11 stamens. The gynoecium contains only one pistil and 3-5 carpels; syncarpous and superior, ovary. The seeds are reddish orange in colour and a mature fruit contains about 50-60 seeds (Puwein and Thomas, 2020). In North East region the flowers start blooming in the month of April and lasts upto 3 months.



Figure 1: *P. polyphylla* plant and rhizome.

P. polyphylla is a shade loving plant which grows luxuriously under the forest canopy of more than 80% shade at an altitude of 1200-3000 msl. Its natural habitats are mainly thick moist forest understory (Liang *et al.*, 2000). Germination is very slow in this species and usually needs 7-8 months to sprout from the seed. It is best suited in humus rich and well drained soil and waterlogging detrimental for this plant. The seeds failed to germinate or grow well or failed to flower or set seed when tried to grow in other environment out of the natural habitat. The growth of this plant is very slow and takes many years (5-7 years) to have mature and economical rhizomes. The slow growth nature of the plant makes it vulnerable to extinction from its natural habitat.

2.1 Ethnomedicinal uses

This plant has many traditional medicinal uses in many countries. This plant is very popular in China and Nepal for its medicinal properties. Some of the traditional uses are as analgesic, diphtheria, boils, removes heat, antispasmodic, depurative stomach ache, appendicitis, Japanese B encephalitis, snake bites, antitussive, ulcers, insect bites, tonsillitis, *etc.* ((Farooque *et al.*, 2004; Maity *et al.*, 2004; Tiwari *et al.*, 2010; Jamir *et al.*, 2012; Lalsangluaii *et al.*, 2013; Pfoze *et al.*, 2013; Mir *et al.*, 2014; Sharma and Samant 2014). In North East India, the leaves and rhizome is used for treating diabetes, stomachache, stomach worms and as tonic.

2.2 Pharmacology

Numerous pharmacological studies showed that compounds and extracts from this plant have a wide range of pharmacological effects, including antitumor, cytotoxic, antimicrobial, antifungal, hemostatic and anti-inflammatory properties. According to the evaluations of toxicity, *Rhizome paridis* may cause mild liver damage (Ding *et al.*, 2021).

2.2.1 Antitumour activity

Polyphyllin D, which is a steroidal saponin from *P. polyphylla* is a potential drug for breast cancer treatment (Lee *et al.*, 2005). Treatment of MCF-7 and MDA-MB-231 cells with polyphyllin D resulted in the inhibition of viability and induction of apoptosis. It was reported that main steroidal saponin of *P. polyphylla* i.e. {Diosgenin-3-a-L-arabinofuranosyl (1-4)-[a-L-rhamnopyranosyl (1-2)]-β-D-Glycopyranoside} is found to have cytotoxicity and caused typical apoptosis (Yan *et al.*, 2009). Anticancer activity against lung adenocarcinoma cell lines were observed when treated with *Rhizoma paridis* saponins, both *in vivo* and *in vitro*. It was also found that *P. polyphylla* showed inhibitory effect on all the cell lines with IC₅₀ values ranging from 10/μg/ml to 30/μg/ml which shows its effectiveness against digestive cancer (Sun *et al.*, 2007). Wang *et al.* (2007) isolated antitumour constituents from *P. polyphylla* var. *yunnanensis* by column chromatography with silica gel which are given below:

- i. Diosgenin-3-O- a -L- arabinofuranosyl (1-4) β-D-glycopyranoside
- ii. Pennogenin-3-O-a -L- arabinofuranosyl (1-4) β-D-glycopyranoside
- iii. Isorhamnetin-3-O- β -D- glycopyranoside
- iv. Ethyl - a -D-fructofuranoside
- v. Pennogenin-3-O-a-L-rhamnopyranosyl(1-4)[a-L-rhamnopyranosyl(1-2)]β-D-glycopyranoside
- vi. Pennogenin-3-O-a-L-rhamnopyranosyl (1-4)[a -L- rhamnopyranosyl (1-4)] a -L-[a -L-
- vii. Rhamnopyranosyl (1-2)] b-D-glycopyranoside

2.2.2 Immunostimulating properties

It was reported that the compounds (1) 3 -O- a -1 - rhamnopyranosyl (1-4) - a -1-rhamnopyranosyl (1-4)-[a-1-rhamnopyranosyl (1-2)- a -d-glucopyranoside, (2) Diosgenin-3-O-a-1- rhamnopyranosyl (1-2)- a -1-arabinofuranosyl- b- d-glucopyranoside and (3) Diosgenin from *P. polyphylla* showed phagocytic activity, respiratory burst and no production in RAW 264.7 (Zhang *et al.*, 2007).

2.2.3 Antibacterial action

The rhizomes of *P. polyphylla* have antibacterial action against various bacteria like *Bacillus dysentery*, *B. typhi*, *B. paratyphi*, *E. coli*, *Staphylococcus aureas*, *Haemolytic streptococci* and *Meningococci* (Anon, 2002). Mayirnao and Bhat (2017) found that the rhizome extract of *P. polyphylla* species had significant antimicrobial activity. At a concentration of 5 mg/ml of the sample, 95-97% of *E. coli*, *S. aureus* and *A. niger* growth were inhibited, while 74% of *T. reesei* were found to be inhibited. *Bacillus dysentery*, *B. paratyphi*, *B. typhi*, *Staphylococcus aureas*, *Escherichia coli*, *Haemolytic streptococci*

and *Meningococci* were found to be inhibited by the rhizomes of *P. polyphylla* (Sharma *et al.*, 2015).

2.2.4 Spermicidal action

Plant extract of *P. polyphylla* was found to be effective as spermicidal. The application of the plant's extract (100 mg/ animal) on vagina prevented pregnancy upto 60% on the tested subjects (Devkota, 2005). Effective spermicidal activity was demonstrated by the plant extracts of *P. polyphylla* against rat and human sperm. When tested on rabbits, it prevented pregnancy by as much as 60% (Pande *et al.*, 2007).

2.2.5 Antifungal

Antifungal activity of *P. polyphylla* saponin has been reported effective against *Cladosporium cladosporioides* and *Candida* species (Deng *et al.*, 2008). *P. polyphylla* var. *yunnanensis* was effective against *Candida albicans*, *Cladosporium cladosporioides* and *Saccharomyces cerevisiae* Meyen ex E.C. Hansen (Sharma *et al.*, 2015). Others a-paristypnin extracted from *P. polyphylla* is found to have a depressant action on carotid pressure, myocardium and respiratory movements. It also affects vasodilation in the spleen and limbs, vasoconstriction in kidney and stimulation in the intestines (Devkota *et al.*, 2005).

2.3 Chemical composition

Devkota *et al.* (2005) had isolated 6 (six) compounds from *P. polyphylla* collected from Parbat district of Nepal. The compounds are:

- i. Przewalskinone B
- ii. Polyphyllin C
- iii. Polyphyllin D
- iv. Saponin-1
- v. Stigmasterol
- vi. Stigmasterol-3-O- β -D-glucoside.

A new saponin, Saponin-polyphyllin A-H has been isolated from the rhizome of *P. polyphylla* (Rastogi and Mehrotra, 1993). A novel steroidal saponin along with the 12 known compounds from *P. polyphylla* var. *chinensis* was also separated from *P. polyphylla* (Yun *et al.*, 2007).

- i. Diosgenin
- ii. Pennogenin
- iii. Diosgenin-3-O-a-L-rhamnopyranosyl (1 → 2)-b-D-glucopyranoside
- iv. Pennogenin-3-O-a-L-rhamnopyranosyl(1 → 2)-b-D-glucopyranoside
- v. Diosgenin-3-O-a-L-rhamnopyranosyl (1 → 4)-[a-Larabinofuranosyl (1 → 4)]-b-D-glucopyranoside
- vi. Pennogenin-3-O-a-L-rhamnopyranosyl (1 → 2)-[a-Larabinofuranosyl (1 → 4)]-b-D-glucopyranoside
- vii. Diosgenin-3-O-a -L-rhamnopyranosyl (1 → 2) [b-Dglucopyranoside (1 → 3)]-b-D-glucopyranoside

- viii. Diosgenin-3-O-a-L-rhamnopyranosyl (1 → 4)-a-Lrhamno-
pyranosyl (1 → 4)[a-L-rhamnopyranosyl (1 → 2)]-b-D-
glucopyranoside
- ix. Pennogenin-3-O-a-Lrhamnopyranosyl (1 → 4)-a-Lrhamno-
pyranosyl (1 → 4) [a-L-rhamnopyranosyl (1 → 2)]-b-D-
glucopyranoside
- x. 3-O-a-Larabinofuranosyl(1 → 4)[a-Lrhamnopyranosyl (1 → 2)]-
b-D-glucopyranoside
- xi. b-D-chacotriosyl-26-O-b- D-glucopyranoside
- xii. 2b,3b,14a,20b,22a,25b hexahydroxycholest- 7-en-6-one, and
- xiii. 2b,3b,14a,20b,24b,25b hexahydroxycholest-7-en-6-one

2.4 Propagation

P. polyphylla grows best in moist humus rich soil under partial or complete shade as in jungle understory. Propagation by seed is a big problem due to its long seed dormancy and slow germination. The best way to propagate this plant is by rhizome or rhizome cuttings. Rhizomes from the wild are collected and used for propagation as well as for medicinal purposes. This leads to a great decline of the herb population. Excessive illegal harvesting and collection for many years has pushed this species towards extinction (Zhang *et al.*, 2011). Matured rhizomes can only be harvested economically after 5-7 years and which is one of the main reasons for the shortage of this plant. Fully economical plant can only be harvested after growing for 5-7 years, which aggravates the shortage of its resources (Wen *et al.*, 2012). In order to conserve this species from extinction, a successful propagation method is the need of the hour (Jianjun *et al.*, 2013). There is an urgent need to discover alternate ways to produce propagules and cultivation methods. A systematic domestication and cultivation method on large scale is really needed for conserving this species and a sustainable supply of this plant material in the market. Rhizome cuttings and tissue cultured planting materials are the prospective methods of propagation of this threatened species.

2.5 Conservation

P. polyphylla is one of the medicinal plants listed as vulnerable by the IUCN (Madhu *et al.*, 2010). This plant has a long seed dormancy period and the germination of the seed takes lots of time. There is difficulty in germination of the seeds under laboratory conditions with different treatments. The growth of the rhizome is very slow and takes almost a year to increase its node number. In addition to its slow growth rate and long dormancy period, the fate of this precious plant is sealed with the increase in its demand, overharvesting, unsustainable collection of rhizomes, harvesting of plants before seed maturity and the increase in urbanization and construction of roads, rail tracks and deforestation. The whole plant, either matured or immature, is often uprooted at the time of harvesting, thereby killing the whole plant. This unsustainable harvesting practice, combined with illegal/cross-border trades of rhizome, deforestation and loss of habitat are common as a result the species is vanishing day-by-day. Market driven collection has pushed to a greedy and unsustainable collection and habitat degradation. Cultivation of the species coupled with *in situ* conservation could be a solution to address the escalated herbal demand (Cunningham *et al.*, 2018).

The College of Horticulture and Forestry, Central Agricultural University, Pasighat, Arunachal Pradesh, India has started the work on micropropagation of this plant species by tissue culture method for the conservation of this vulnerable species. Tissue cultured plants are produced from rhizomes in the laboratory of College of Horticulture and Forestry, Pasighat, Arunachal Pradesh. The plants will be planted in the wild that it can flourish and increases its population in nature away from human disturbance.

3. *Kaempferia parviflora* Wall. ex Baker

K. parviflora commonly known as 'Black galangale', is an important herbaceous plant which is rhizomatous in nature. It belongs to Zingiberaceae family and it has high medicinal value (Techaprasan *et al.*, 2010). The rhizomes are the economical part and it is mainly cultivated for its various medicinal properties. They have been widely used in traditional medicines to cure fevers, hypertension, metabolic disorders, inflammation, urinary tract infections, coughs, erectile dysfunction, abdominal and gastrointestinal ailments, asthma, wounds, rheumatism, epilepsy, and skin diseases (Abdelsamed *et al.*, 2019). More than 60 species of *Kaempferia* species are distributed from India to Southeast Asia, whereas, Thailand has the richest *Kaempferia* biodiversity with more than 20 different species (Sirirugsa, 1992; Larsen and Larsen *et al.*, 2006). The rhizomes are popularly added as a flavoring agent in local cuisine and also used as traditional medicine for a wide range of ailments in Thailand (Sutthanut *et al.*, 2007; Konkumnerd *et al.*, 2010). The genus *Kaempferia* is subdivided into two sub-genera: *K. subgen. Kaempferia* and subgen. *Protanthium* (Horan.) Baker based on the positioning of the inflorescence. *Kaempferia* subgen. *Kaempferia* produces terminal inflorescences while the subgen. *Protanthium* produces the inflorescences directly from the rhizome (Saensouk *et al.*, 2021).

It is a perennial herb with reddish black rhizome. It has thin ovate green leaves with a dimension of 13.5-23.2 cm × 9.3-13.5 cm. The petioles are 9-10 cm in length. The inflorescence is pedunculate and 5.1-5.4 cm in length. The flowers are sessile and white with purple tinged colour. The bracts are lanceolate and slightly greenish in colour measuring 1.4-1.7 × 3-3.5 cm in size. Calyx is white (2.4 × 0.7 cm) and corolla lobes are linear white (1.5 cm). Staminodes are lateral, linear, white in colour measuring 1.1 × 0.3 cm. Labellum is white tinged with purple at the base and lip is lilac in colour, obovate, cuneate, bifid and 1.2 × 0.8 cm in size. Anthers are sessile, crested and 0.4 × 0.1 cm long. Stigma is 4.2 cm in size and the ovary is velutinous, trilocular and elliptic in shape (Devi *et al.*, 2016).



Figure 2: *K. parviflora* plant and rhizome.

In Thailand, the rhizomes of this plant are widely used as a flavoring agent and as a traditional medicine for a wide range of ailments (Sutthanut *et al.*, 2007; Konkumnerd *et al.*, 2010). *K. parviflora* is also used as an aphrodisiac, to treat colic disorder, peptic and duodenal ulcers, to lower blood glucose levels, improve blood circulation and to increase vitality (Yenjai *et al.*, 2004, Akase *et al.*, 2011). Numerous pharmacological studies have been prompted by the recognition of high potential of this plant for the development of various health products (Chivapat *et al.*, 2010).

3.1 Pharmacology

K. parviflora has many medicinal uses and found to be effective against knee osteoarthritis, inhibition of a breast cancer resistance protein (BCRP), wound healing, antidrug resistant strains of *Mycobacterium tuberculosis*, neuroprotective, antinociceptive, human immunodeficiency virus type-1 (HIV-1) inhibitory activity, *in vitro* antiallergenic, and larvicidal activity against *Aedes aegypti* are some of the pharmacological properties of this plant. The following is a list of some of the reported biological activities of *Kaempferia* plant and its extracts (Ekor, 2013; Techaprasan *et al.*, 2010; Wutythamawech, 1997; Yenjai *et al.*, 2004; Akase *et al.*, 2011; Nakao *et al.*, 2011; Sivarajan and Balachandran, 1994; Hidaka *et al.*, 2017).

3.1.1 Anticancer activity

The ethanol extract of *K. parviflora* suppresses the PI3K/AKT and MAPK signalling pathways, which in turn promotes cell apoptosis in HeLa cells from cervical cancer patients. HeLa cell invasion and migration are also prevented by *K. parviflora*. This could have something to do with the inhibition of MMP-2 expression. It has been demonstrated that the ethanol extract of *K. parviflora* has an effect on human cholangiocarcinoma cell lines (HuCCA-1 and RMCCA-1) with CC_{50} values of 46.1 g/ml and 62.0 g/ml, respectively. During cancer treatment, the ATP-binding cassette (ABC) protein P-glycoprotein (P-gp) extrudes multiple chemotherapeutic agents, resulting in multidrug resistance. It was discovered that the ethanol extract of *K. parviflora* and compound-1 increased the accumulation of rhodamine 123 and daunorubicin, which are the substrates of P-gp (Patanasethanont *et al.*, 2007). Multidrug resistance may also be caused by the multidrug resistance associated proteins (MRPs), such as MRP-1, -2, and -3. Unlike the usual P-gp inhibitor, verapamil, MRPs inhibitors can significantly increase the accumulation of calcein, a MRPs substrate. By inhibiting MRP functions, it was discovered that the ethanol extract of *K. parviflora* and the methoxyflavones increased the dose-dependent accumulation of calcein and doxorubicin in A549 cells. Doxorubicin accumulation in A549 cells is stimulated most strongly by compound 6. On the other hand, it would appear that 5-hydroxy-methoxyflavones are not associated with these stimulatory effects (Saranyapin *et al.*, 2017).

3.1.2 Antiobesity activity

K. parviflora extracts were tested against obesity-induced dermatopathy system. Ethanolic extract, a polymethoxyflavonoid-rich fraction (PMF), and a polymethoxyflavonoid-poor fraction from *K. parviflora* were used in the experiment. It was found that the ethanolic extract reduced the body weight of the mouse and the thickness of the subcutaneous fat layer more effectively (Hidaka *et al.*, 2017).

3.1.3 Antiinflammatory activity

The ethanol extract of *K. parviflora* had a significant effect on PGE2 inhibition. It was also reported that the plant extract and 3',4',5,7-tetramethoxyflavone (86) both inhibited iNOS-mRNA expression in a dose-dependent manner. In addition, rat paw oedema was reduced by the soluble subfractions of H₂O, C₂H₅OH, C₄H₈O₂, CHCl₃ and n-hexane, which demonstrated excellent *in vivo* antiinflammatory activity (Sae-wong *et al.*, 2009). Ethanol extract of 80 per cent was found to have antioxidative activity of polyphenolics against the oxidizing effects of UV radiation (Lee *et al.*, 2018). The methoxylated flavones in the 60% ethanol and ethyl acetate soluble fraction of the 100% methanol extracts of *K. parviflora* likely contributed to the reduction in knee osteoarthritis (Kobayashi *et al.*, 2018). Methoxylated flavonoids were isolated from *K. parviflora*; namely, 5-hydroxy-3,7,3',4'-tetramethoxyflavone (96), 5-hydroxy-7,4'-dimethoxyflavone (93), and 5-hydroxy-3,7,4'-trimethoxyflavone (95) which has IC₅₀ values of 16.1 M, 24.5 M. They described methoxyflavones 96, 93, and 95 from a hexane extract of *K. parviflora* rhizomes were active against NO release in RAW264.7 cells (Tewtrakul *et al.*, 2009).

3.1.4 Anticholinesterase activity

Thirteen known methoxyflavones (1-13) were isolated from *K. parviflora* and their structures were completely elucidated based on NMR analysis. 5,7,4'-trimethoxyflavone (6) and 5,7-dimethoxyflavone (7) were the highest potential cholinesterase inhibitors towards AChE and BChE. The compounds bearing 5,7-dimethoxy groups and a free substituent at C-3 had a significant inhibitory effect at a concentration of 0.1 mg/ml, but those bearing a 5-hydroxyl group reduced the inhibitory potency. Significant finding in the treatment of Alzheimer's disease was that a methanol extract as well as compounds (86-87) isolated from *K. parviflora* rhizomes inhibited acetylcholinesterase and butyryl cholinesterase more effectively than (86) (Sawasdee *et al.*, 2009).

3.1.5 Antimutagenicity activity

Soluble CH₂Cl₂ and ethyl acetate fractions of *K. parviflora* exhibited antimutagenicity and glucosidase inhibitory activity. With IC₅₀ values of 0.40, 0.40, 0.42, and 0.47 nmol/plate, respectively, isolated methoxylated compounds from these extracts (86, 97, 84, and 92) demonstrated potent activity. With IC₅₀ values of 20.4 M, 54.3 M, and 64.3 M, respectively, compounds 88, 87, and 91 also demonstrated significant activity (Azuma *et al.*, 2011).

3.1.6 Effect on cytochromes

It was found that continuous consumption of compound (88) isolated from *K. parviflora* lowers liver CYP3A expression, which raises levels of CYP3A-metabolized substances like midazolam. *K. parviflora* extract significantly induced CYP1A1, CYP1A2 enzyme activities following short-term treatment (Ochiai *et al.*, 2018). CYP2B enzyme activities were markedly increased by all *K. parviflora* extract treatment, whereas *K. parviflora* extract significantly enhanced CYP2E1 activity only after long-term treatment (Mekjaruskul *et al.*, 2012).

3.1.7 Adaptogenic activity

K. parviflora extracts made from hexane, chloroform, methanol and ethanol had more adaptogenic properties than the reference's crude

ginseng root powder. In humans, the whole-body potential expenditure was increased by a single oral dose of *K. parviflora* rhizome (60 per cent ethanol extract) (Pripdeevech *et al.*, 2012). Improvement in physical fitness and health was observed when treated *K. parviflora* (Wattanathorn *et al.*, 2012).

3.1.8 Xanthine oxidase inhibitory activity

With IC_{50} values of 0.9 and >4 mM, respectively, of the isolated methoxylated flavonoids from *K. parviflora* (87 and 86) inhibit xanthine oxidase activity (Nakao *et al.*, 2011).

3.1.9 Allergenic activity

In addition to H_2O , CH_2Cl_2 , ethyl acetate and isolated polymethoxyflavones from *K. parviflora* (86,97) antiallergenic activity was observed highest with compounds (92) and (94) extracted from *K. parviflora* (Kobayashi *et al.*, 2015).

3.1.10 Neurological activity

By increasing levels of serotonin, norepinephrine, and dopamine in the rat hippocampus, a methanolic extract of *K. parviflora* (95 per cent methanol) exhibited neuroprotective activity (Plaingam *et al.*, 2017). Tonsomboon *et al.* (2021) found out that in HT-22 cells, co-treatment of glutamate with *K. parviflora* extract significantly inhibited glutamate-mediated cytotoxicity and decreased intracellular ROS production. Additionally, the glutamate-induced apoptosis and apoptotic-inducing factor (AIF) translocation were blocked by *K. parviflora* extract co treatment. Western blot analysis also demonstrated that *K. parviflora* extract significantly diminished extracellular signal-regulated kinase (ERK) phosphorylation induced by glutamate, and brain-derived neurotrophic factor (BDNF) was recovered to the control. Moreover, this *K. parviflora* extract treatment prolonged the lifespan of *C. elegans*. Altogether, this study suggested that *K. parviflora* extract possesses both neuroprotective and longevity-inducing properties, thus serving as a promising candidate for development of innovative health products.

3.1.11 Effects on sexual performance

7-methoxyflavones (86, 87, 89, 91, 93-95) from *K. parviflora* rhizomes were found to increase male sexual activity by inhibiting PDE5, with 86 being the most effective. According to Temkitthawon *et al.* (2011), methoxyls at positions C5 and C752 were responsible for the activity. In healthy men, standardized *K. parviflora* rhizome extracts of 5% DMF also improve erectile function (Stein *et al.*, 2018). In addition to boosting testosterone production, *K. parviflora* extract and 5,7-dimethoxyflavones reduce age-related diseases and hypogonadism (Horigome *et al.*, 2016). When streptozotocin (STZ)-induced diabetic rats were given *K. parviflora* extract (aqueous with 1% Tween-80) 49, their testosterone levels, sperm count and sexual performance are all increased (Lert-Amornpat *et al.*, 2017).

Miscellaneous

In mice, the rhizome extract of *K. parviflora* (95 per cent ethanol) reduced obesity by inhibiting adipogenesis, lipogenesis and muscle atrophy (Lee *et al.*, 2018). *K. parviflora* derivatives of the 5-hydroxy-7-methoxyflavone, on the other hand, cause skeletal muscle hypertrophy (Ono *et al.*, 2019). Plant extract of *K. parviflora* (95 per cent ethanol) has activity against propioni bacteria, sebostatics, and inflammation and as a potential acne treatment (Jin and Lee, 2018). In a study for rheumatoid arthritis, ethanol extract significantly

reduced the levels of gene expression for matrix-degrading enzymes, inflammatory mediators, and pro-inflammatory cytokines, it did not alter the cell cycle or induce apoptosis. Without interfering with the NF- κ B pathway, they also reported that the alcoholic extract inhibits cell migration, reduces the mRNA expression of cadherin-11, and selectively reduces the phosphorylation of signal transducers, activators of transcription 1 (STAT1), and STAT3 signaling molecules (Kongdang *et al.*, 2019).

3.2 Chemical composition

14 compounds, including two polyoxygenated cyclohexanes (1 and 2), eleven flavonoids (3-13), and sitosterol (14), were identified through phytochemical analysis of *K. parviflora* extract. The primary methoxyflavones in *K. parviflora* have been identified structurally, according to phytochemical analysis (Kim *et al.*, 2007 and Sutthanut *et al.*, 2007). Five compounds including 5,7-dimethoxyflavone (1), 3,5,7-trimethoxyflavone (2), di-O-methylpinocembrin (3), bisdemethoxycurcumin (4), aloe-emodin (5) were isolated from the n hexane extract of *K. parviflora* rhizomes (Tri *et al.*, 2021). Structure-activity relationship (SAR) studies of *K. parviflora* methoxyflavones reveal that methoxylation at position 5 and vicinal methoxylation at positions 3' and 4' play crucial roles in promoting this activity in order to investigate the antiallergic activity in inhibiting degranulation (Azuma *et al.*, 2008; Okabe *et al.*, 2014). It has been demonstrated that compounds 4 and 6 have the greatest inhibitory effects on the activity of acetylcholinesterase (AChE) and butyrylcholinesterase (BChE). According to the SAR study, hydroxylation at position 5 reduces the inhibitory effects on AChE and BChE, while dimethoxylation at positions 5 and 7 is required for these effects to occur. Another study demonstrates that PDE-5 inhibition relies heavily on the methoxyl group at position 5 in 7-methoxyflavones. Methoxylation at position 4', on the other hand, has no effect on PDE inhibition (Temkitthawon *et al.*, 2011). Additionally, B16 melanoma 4A5 cells are less cytotoxic to methoxyflavones when methylated at position 5. As a result, compound-11 is more cytotoxic than compound-359. KP methoxyflavone structural modifications have been investigated. Compound-6's various oxime derivatives are synthesized and found to be cytotoxic to HepG2 and T47D cell lines (Sutthanut *et al.*, 2007). For the purpose of examining the characteristics of pharmacokinetics, three methoxyflavones, 3,5,7,3',4'-pentamethoxyflavone (compound 1), 5,7,4'-trimethoxyflavone (compound 4) and compound 6, were chosen. Within 1 to 2 h of oral administration to rats, they quickly reach their peak concentration. However, they exhibit a very low bioavailability of one to four per cent. Pitakpawasutthi *et al.* (2018) reported that *K. parviflora* oil contains a copaene, dauca 5, 8 diene, camphene, b pinene, borneol, and linalool as major components.

3.3 Conservation

K. parviflora is one of the important medicinal plants which has high demand but has not been domesticated or grown extensively in the region. The plant is highly sought after due to its high medicinal value. It is difficult for the plant to survive and reproduce in its natural environment when it is collected from the wild in an uncontrolled and unwise manner. Currently, this crop is being grown on a large and small scale in the region. The application of tissue culture to the large-scale multiplication of this crop has been taken up by our Institute at the College of Horticulture and Forestry. In order for the tissue-cultured plants to grow and multiply in the wild

without being harmed by human activities, they were relocated to their natural or similar habitats. The species can be preserved and its population can grow in this way. The availability of *K. parviflora* materials in the market and the prevention of its extinction will be aided by strict forest policies, wild collection policies, and cultivation subsidies.

4. Conclusion

P. polyphylla and *K. parviflora* are two important medicinal plants that are found in the North-eastern part of India. Both of these plants are under threat from overexploitation and a lack of cultivation efforts. Numerous ailments can be treated with the help of these two plants. These plants are very expensive in the market because there are fewer of them in the wild and there are fewer of them available from cultivation. At present, the marketing of these species are not organized yet and there is a huge collection of these species from the wild and traded illegally. Unsustainable and injudicious harvesting from the wild along with illegal trading of these species push them towards extinction. *P. polyphylla* typically thrives in temperate climates at high altitudes as a forest understory. The propagation of this species takes a long time in its natural habitat and with long seed dormancy period makes it difficult to multiply and propagate in nature as well. The species is illegally taken from the wild and causes a decline in population as a result of its high demand on the global market. The IUCN classifies this species as a threatened species. Since *K. parviflora* can easily be grown from its rhizomes, many people have started cultivating this species. In addition, there is a significant market demand for this plant, many of which can be met through cultivation. Before it's too late, scientists and policymakers need to develop conservation policies and conduct research on these two species.

Conflict of interest

The authors declare that there are no conflicts of interest relevant

References

- Abdelsamed, I.E.; Tarik, A.M.; Ahmed, F.E.; Ahmed M.A.; Ali, S.A.; Abdelaaty, A.S.; Tatsuro, Y.; Abdel, R.H.F.; Masaaki N.; Hesham R.E.; Akemi U.; Paul, W.P. and Mohamed, E.F.H. (2019). Recent advances in *Kaempferia* phytochemistry and biological activity: A comprehensive review. *Nutrients*, 11(10):2396.
- Akase, T.; Shimada, T.; Terabayashi, S.; Ikeya, Y.; Sanada, H. and Aburada, M. (2011). Antiobesity effects of *Kaempferia parviflora* in spontaneously obese type II diabetic mice. *Journal of Natural Medicines*, 65(1):73-80.
- Anonymous (2002). Newsfinder, pp:1-4.
- Ansari, M.H.R. and Ahmad S. (2019). Herbs that heal: Natural remedies for health promotion and longevity. *Ann. Phytomed.*, 8(1):7-18.
- Aziz, M.A.; Adnan, M.; Khan, A.H.; Rehman, A.U.; Jan, R. and Khan, J. (2016). Ethnomedicinal survey of important plants practiced by indigenous community at Latha subdivision, South Waziristan agency, Pakistan. *Journal of Ethnobiology and Ethnomedicine*, 12(1):53.
- Aziz, M.A.; Khan, A.H.; Adnan, M. and Izatullah, I. (2017). Traditional uses of medicinal plants reported by the indigenous communities and local herbal practitioners of Bajaur Agency, Federally Administrated Tribal Areas Pakistan. *Journal of Ethnopharmacology*, 198:268-281.
- Azuma, T.; Kayano, S.I.; Matsumura, Y.; Konishi, Y.; Tanaka, Y. and Kikuzaki, H. (2011). Antimutagenic and α -glucosidase inhibitory effects of constituents from *Kaempferia parviflora*. *Food Chemistry*, 125(2):471-475.
- Azuma, T.; Tanaka, Y. and Kikuzaki, H. (2008). Phenolic glycosides from *kaempferia parviflora*. *Phytochemistry*, 69(15):2743-2748.
- Chivapat, S.; Chavalittumrong, P.; Attawish, A. and Rungsipat, A. (2010). Chronic toxicity study of *Kaempferia parviflora* Wall ex. extract. *Thai Journal of Veterinary Medicine*, 40(4):377-383.
- Cragg, J.G. and Donald, S.G. (1993). Testing identifiability and specification in instrumental variable models. *Economic Theory*, 9(2):222-240.
- Cunningham, A.B.; Brinckmann, J.A.; Bi, Y.F.; Pei, S.J.; Schippmann, U. and Luo, P. (2018). Paris in the spring: A review of the trade, conservation and opportunities in the shift from wild harvest to cultivation of *Paris polyphylla* (Trilliaceae). *Journal of Ethnopharmacology*, 222:208-216.
- Deng, D.; Lauren, D.R.; Cooney, J.M.; Jensen, D.J.; Wurms, K.V.; Upritchard, J.E.; Cannon, R.D.; Wang, M.Z. and Li, M.Z. (2008). Antifungal saponins from *Paris polyphylla* Smith. *Planta Medica*, 24(11):1397-1402.
- Devi, N.B.; Das, A.K. and Singh, P.K. (2016). *Kaempferia parviflora* (Zingiberaceae): A new record in the Flora of Manipur. *International Journal of Innovative Science, Engineering and Technology*, 3(7):2348-7968.
- Devkota, K.P. (2005). Bioprospecting studies on *Sarcococca hookeriana* Bail, *Sonchus wightianus* DC, *Paris polyphylla* Smith and related Medicinal Herbs of Nepal: Ph D Thesis, HEJ Research Institute of Chemistry, International Centre for Chemical Science, University of Karachi, Karachi-75270, Pakistan.
- Ding, Y.G.; Zhao, Y.L.; Zhang, J.; Zuo, Z.T.; Zhang, Q.Z. and Wang, Y.Z. (2021). The traditional uses, phytochemistry, and pharmacological properties of *Paris* L. (Liliaceae): A review. *Journal of Ethnopharmacology*, 278:114293.
- Ekor, M. (2013). The growing use of herbal medicines: Issues relating to adverse reactions and challenges in monitoring safety. *Frontiers in Pharmacology*, 4:177.
- Farooquee, N.A.; Majila, B.S. and Kala, C.P. (2004). Indigenous knowledge systems and sustainable management of natural resources in a high altitude society in Kumaun Himalaya, India. *Journal of Human Ecology*, 16:33-42.
- Goldspiel, B.R. (1997). Clinical overview of the taxanes. *Pharmacotherapy*, 17(5):110-125.
- Govaerts, R.H.A. (2011). World checklist of selected plant families published update facilitated by the Trustees of the Royal Botanic Gardens, Kew.
- Hidaka, M.; Horikawa, K.; Akase, T.; Makihara, H.; Ogami, T.; Tomozawa, H.; Tsubata, M.; Ibuki, A. and Matsumoto, Y. (2017). Efficacy of *Kaempferia parviflora* in a mouse model of obesity-induced dermatopathy. *Journal of Natural Medicines*, 71(1):59-67.
- Husain, M. K. (2021). Herbs that heal: Relevance of traditional natural remedies in promotion of health. *Ann. Phytomed.*, 10(2):4-21.
- Jamir, N.S.; Lanusunep and Pongener, N. (2012). Medico-herbal medicine practiced by the Naga tribes in the state of Nagaland (India). *Indian Journal of Fundamental and Applied Life Sciences*, 2:328-333.

- Jianjun, Q.; Zheng, N.A.; Zhang, B.; Sun, P.; Hu, S.; Xu, W.; Ma, Q.; Zhao, T.; Zhou, L.; Qin, M. and Xianen, L. (2013). Mining genes involved in the stratification of *Paris polyphylla* seeds using high-throughput embryo transcriptome sequencing. *BMC Genomics*, **14**:358.
- Jin, S. and Lee, M.Y. (2018). *Kaempferia parviflora* extract as a potential anti-acne agent with anti-inflammatory, sebostatic and anti-propionibacterium acnes activity. *International Journal of Molecular Sciences*, **19**(11): 3457.
- Kim, J.; Park, K.; Lee, C. and Chong, Y. (2007). Synthesis of a complete series of O-Methyl analogues of naringenin and apigenin. *Bulletin of the Korean Chemical Society*, **28**(12):2527-2530.
- Kobayashi, H.; Suzuki, R.; Sato, K.; Ogami, T.; Tomozawa, H.; Tsubata, M.; Ichinose, K.; Aburada, M.; Ochiai, W. and Sugiyama, K. (2018). Effect of *Kaempferia parviflora* extract on knee osteoarthritis. *Journal of Natural Medicines*, **72**:136-144.
- Kobayashi, S.; Kato, T.; Azuma, T.; Kikuzaki, H. and Abe, K. (2015). Anti-allergenic activity of polymethoxyflavones from *Kaempferia parviflora*. *Journal of Functional Foods*, **13**:100-107.
- Kongdang, P.; Jaitham, R.; Thonghoi, S.; Kuensaen, C.; Pradit W. and Ongchai, S. (2019). Ethanolic extract of *Kaempferia parviflora* interrupts the mechanisms-associated rheumatoid arthritis in SW982 culture model via p38/STAT1 and STAT3 pathways. *Phytomedicine*, **59**: 152755.
- Konkumnerd, J.; Karnchanatat, A. and Sangvanich, P. (2010). A thermostable lectin from the rhizomes of *Kaempferia parviflora*. *Journal of the Science of Food and Agriculture*, **90**(11):192-1925.
- Lalsangluaii, F.; Chinlapianga, M. and Shukla, A.C. (2013). Efficacy and potency of *Paris polyphylla* Smith, an ethno-medicinal plant of Mizoram. *Science and Technology Journal*, **1**:36-40.
- Larsen, K. and Larsen, S.S. (2006). Gingers of Thailand. *Queen Sirikit Botanic Garden, Chiang Mai*, pp:1-130.
- Lee, M.H.; Han, A.R.; Jang, M.; Choi, H.K.; Lee, S.Y.; Kim, K.T. and Lim, T.G. (2018). Antiskin inflammatory activity of black ginger (*Kaempferia parviflora*) through antioxidative activity. *Oxidative Medicine and Cellular Longevity*, 5967150. doi: 10.1155/2018/5967150.
- Lee, M.S.; Yuet-Wa, J.C.; Kong, S.K.; Yu, B.; Eng-Choon, V.O.; Nai-Ching, H.W. and Chung-Wai, T.M. et al. (2005). Effects of polyphyllin D, a steroidal saponin in *Paris polyphylla*, in growth inhibition of human breast cancer cells and in xenograft. *Cancer Biology and Therapy*, **4**: 1248-1254.
- Lee, S.; Kim, C.; Kwon, D.; Kim, M.B. and Hwang, J.K. (2018). Standardized *Kaempferia parviflora* Wall. ex Baker (Zingiberaceae) extract inhibits fat accumulation and muscle atrophy in ob/ob mice. *Evidence Based Complementary and Alternative Medicine*, 8161042. <https://doi.org/10.1155/2018/8161042>.
- Lert-Amornpat, T.; Maketon, C. and Fungfuang, W. (2017). Effect of *Kaempferia parviflora* on sexual performance in streptozotocin-induced diabetic male rats. *Andrologia*, **49**(10):1-6.
- Liang, S. and Soukup, V.G. (2012). "*Paris polyphylla*", retrieved 2012-05-01, in Wu, Zhengyi; Raven, Peter H. & Hong, Deyuan, eds. (1994 onwards), *Flora of China* (online), eFloras.org, retrieved 2012-05-01.
- Liang, S.J. and Soukup, V.G. (2000). *Paris Linnaeus*. In: Chen Sing-chi, Liang Song-jun, Xu Jiemei, Tamura MN. (eds) *Flora of China*. Vol. 24. Flagellariaceae through Marantaceae. Science Press (Beijing), China and Missouri Botanic Garden Press (St. Louis), USA. pp:88-95.
- Liang, S.Y. and Soukup, V.G. (2000). *Paris Linnaeus*. In: *Flora of China Volume 24 Flagellariaceae through Marantaceae*. Wu ZY, Raven PH ed. (Beijing, China: Science Press and St Luis, USA: Botanical Garden Press), pp:88-95.
- Madhu, K.C.; Sussana, P. and Pramod, K.J. (2010). *Ecological Study of Paris polyphylla* Sm. *Ecological Society*, **17**:87-93.
- Mahmood, A.; Mahmood, A.; Shaheen, H.; Qureshi, R.A.; Sangi, Y. and Gilani, S.A. (2011). Ethnomedicinal survey of plants from district Bimber Azad Jammu and Kashmir. *Pakistan Journal of Medicinal and Aromatic Plants*, **5**(11):2348-2360.
- Maity, D.; Pradhan, N. and Chauhan, A.S. (2004). Flock uses of some medicinal plants from North Sikkim. *Indian Journal of Traditional Knowledge*, **3**:66-71.
- Mayirnao, H. and Bhat, A.A. (2017). Evaluation of antioxidant and antimicrobial activity of *Paris polyphylla* Sm. *Asian journal of pharmaceutical and clinical research*, **10**(11):315-319.
- Mehrotra, N. (2021). Herbs that heal: Nature's pharmacy endowed remedies for better health. *Ann. Phytomed.*, **10**(1):6-22.
- Mekjaruskul, C., Jay, M. and Sripanidkulchai, B. (2012). Modulatory effects of *Kaempferia parviflora* extract on mouse hepatic cytochrome P450 enzymes. *Journal of Ethnopharmacology*, **141**(3):831-839.
- Mir, A.H.; Upadhaya, K. and Choudhury, H. (2014). Diversity of endemic and threatened ethnomedicinal plant species in Meghalaya, North-east India. *International Research Journal of Environmental Sciences*, **3**:64-78.
- Nakao, K.; Murata, K.; Deguchi, T.; Itoh, K.; Fujita, T.; Higashino, M.; Yoshioka, Y.; Matsumura, S.I.; Tanaka, R. and Shinada, T. (2011). Xanthine oxidase inhibitory activities and crystal structures of methoxyflavones from *Kaempferia parviflora* rhizome. *Biological and Pharmaceutical Bulletin*, **34**(7):1143-1146.
- Ochiai, W.; Kobayashi, H.; Kitaoka, S.; Kashiwada, M.; Koyama, Y.; Nakaishi, S.; Nagai, T.; Aburada, M. and Sugiyama, K. (2018). Effect of the active ingredient of *Kaempferia parviflora*, 5,7-dimethoxyflavone, on the pharmacokinetics of midazolam. *Journal of Natural Medicines*, **72**(3):607-614.
- Okabe, Y.; Shimada, T.; Horikawa, T. et al. (2014). Suppression of adipocyte hypertrophy by polymethoxyflavonoids isolated from *Kaempferia parviflora*. *Phytomedicine*, **21**(6):800-806.
- Ono, S.; Yoshida, N.; Maekawa, D.; Kitakaze, T.; Kobayashi, Y.; Kitano, T.; Fujita, T.; Okuwa-Hayashi, H.; Harada, N. and Nakano, Y. (2019). 5-Hydroxy-7-methoxyflavone derivatives from *Kaempferia parviflora* induce skeletal muscle hypertrophy. *Food Science and Nutrition*, **7**:312-321.
- Pande, P.C.; Tiwari, L. and Pande, H.C. (2007). Ethnoveterinary plants of Uttaranchal- A review. *Indian Journal of Traditional Knowledge*, **6**:444-458.
- Patanasethanont, D.; Nagai J, Yumoto R, Murakami, T; Sutthanut, K; Bung-Orn Sripanidkulchai, B.O; Jenjai, C. and Takano, M. (2007). *Journal of Pharmaceutical Sciences*, **96**(1):223-233.
- Paul, A.; Bharali, S.; Khan, M.L. and Tripathi, O.P. (2013). Anthropogenic disturbances led to risk of extinction of *Taxus wallichiana* Zuccarini, an endangered medicinal tree in Arunachal Himalaya. *Natural Areas Journal*, **33**:447-454.
- Paul, A.; Gajurel, P.R. and Das A.K. (2015). Threats and conservation of *Paris polyphylla* an endangered, highly exploited medicinal plant in the Indian Himalayan region. *Biodiversitas*, **16**(2):295-302.

- Pfoze, N.L.; Kumar, Y. and Myrboh, B. (2013). Screening of bioactive phytochemicals obtained from lesser known ethnomedicinal plants of Senapati district of Manipur, India. *Pleione*, 7:489-500
- Pitakpawasutthi, Y.; Palanuvej, C. and Ruangrunsi, N. (2018). Quality evaluation of *Kaempferia parviflora* rhizome with reference to 5,7-dimethoxyflavone. *Journal of Advanced Pharmaceutical Technology and Research*, 9:26-31.
- Plaingam, W.; Sangsuthum, S.; Angkhasirisap, W. and Tencomnao, T. (2017). *Kaempferia parviflora* rhizome extract and *Myristica fragrans* volatile oil increase the levels of monoamine neurotransmitters and impact the proteomic profiles in the rat hippocampus: Mechanistic insights into their neuroprotective effects. *Journal of Traditional and Complementary Medicine*, 7(4):538-552.
- Pokhrel, G.; Upadhyaya, A. and Thapa, M.S. (2019). Threats and Conservation of *Paris polyphylla*: Vulnerable Medicinal Plant in Panchase Protected Forest, Nepal. *Forestry: Journal of Institute of Forestry*, 16:14-30.
- Pripdeevech, P.; Pitija, K.; Rujjanawate, C.; Pojanagaroon, S.; Kittakoop, P. and Wongpornchai, S. (2012). Adaptogenic-active components from *Kaempferia parviflora* rhizomes. *Food Chemistry*, 132(3):1150-1155.
- Puwein, A. and Thomas, S.C. (2020). An overview of *Paris polyphylla*, a highly vulnerable medicinal herb of Eastern Himalayan region for sustainable exploitation. *The Natural Products Journal*, 10:314.
- Rastogi, P. and Mehrotra, B.N. (1993). *Compendium of Indian Medicinal Plants: CDRI-Lucknow and National Institute of Science and Communication New Delhi*, 3:479-480.
- Sae-wong, C.; Tansakul, P. and Tewtrakul, S. (2009). Antiinflammatory mechanism of *Kaempferia parviflora* in murine macrophage cells (RAW 264.7) and in experimental animals. *Journal of Ethnopharmacology*, 124:576-580.
- Saranyapin, P.; Siriwoot, S.; Mingkwan, N.T.; Pitchaya, M.; Nitwara, W.; Duncan, R.S. and Wutigri, N. (2017). *Kaempferia parviflora* extract exhibits anticancer activity against HeLa Cervical cancer cells. *Frontiers in Pharmacology*, 8:630.
- Sawasdee, P.; Sabphon, C.; Sitthiwongwanit, D. and Kokpol, U. (2009). Anticholinesterase activity of 7-methoxyflavones isolated from *Kaempferia parviflora*. *Phytotherapy Research*, 23(12):1792-1794.
- Shah, A.S.; Mazumder, P.B. and Choudhury, M.D. (2012). Medicinal properties of *Paris polyphylla* smith: A review. *Journal of Herbal Medicine and Toxicology*, 6(1):27-33.
- Sharma, A.; Kalita, P. and Tag, H. (2015). Distribution and phytomedicinal aspects of *Paris polyphylla* Smith from the Eastern Himalayan Region: A review. *Humanitas Medicine*, 5(3):15.
- Sharma, P. and Samant, S.S. (2014). Diversity, distribution and indigenous uses of medicinal plants in Parbati Valley of Kullu district in Himachal Pradesh, Northwestern Himalaya. *Asian Journal of Advanced Basic Sciences*, 2:77-98.
- Sharma, V. (2021). Ayurveda and remedial plants in medication. *Ann. Phytomed.*, 10(1):1-5.
- Siriruga, P. (1992). Taxonomy of the genus *Kaempferia* (Zingiberaceae) in Thailand. *Thai Forest Bulletin*, 19:1-15.
- Sivarajan, V. and Balachandran, I. (1994). *Ayurvedic Drugs and their Plant Sources* Oxford and IBH Publishing Co. Oxford & IBH Pub. Co; New Delhi, India.
- Stein, R.A.; Schmid, K.; Bolivar, J.; Swick, A.G.; Joyal, S.V. and Hirsh, S.P. (2018). *Kaempferia parviflora* ethanol extract improves self-assessed sexual health in men: A pilot study. *Journal of Integrative Medicine*, 16(4):249-254.
- Sun, J.; Liu, B.R.; Hu, W.J.; Yu, L.X. and Qian, X.P. (2007). In vitro anticancer activity of aqueous extracts and ethanol extracts of fifteen traditional Chinese medicines on human digestive tumor cell lines. *Phytotherapy Research*, 21(11):1102-1104.
- Sutthanut, K.; Sripanidkulchai, B.; Yenjai, C. and Jay, M. (2007). Simultaneous identification and quantitation of 11 flavonoid constituents in *Kaempferia parviflora* by gas chromatography. *Journal of Chromatography A*, 1143(1-2):227-233.
- Techaprasan, J.; Klinbunga, S.; Ngamriabsakul, C. and Jenjittikul, T. (2010). Genetic variation of *Kaempferia* (Zingiberaceae) in Thailand based on chloroplast DNA (psbA-trnH and petA-psbJ) sequences. *Genetics and Molecular Research*, 9(4):1957-1973.
- Temkitthawon, P.; Hinds, T.R.; Beavo, J.A.; Viyoch, J.; Suwanborirux, K.; Pongamornkul, W.; Sawasdee, P. and Ingkaninan, K. (2011). *Kaempferia parviflora*, a plant used in traditional medicine to enhance sexual performance contains large amounts of low affinity PDE5 inhibitors. *Journal of Ethnopharmacology*, 137(3):1437-1441.
- Tewtrakul S.; Subhadhirasakul S.; Karalai C.; Ponglimanont C. and Cheenpracha S. (2009). Anti-inflammatory effects of compounds from *Kaempferia parviflora* and *Boesenbergia pandurata*. *Food Chemistry*, 115:534-538.
- Thomas, P. and Farjon, A. (2011). *Taxus wallichiana*. The IUCN red list of threatened species, <https://doi.org/10.2305/IUCN.UK.2011-2.RLTS.T46171879A9730085.en>.
- Tiwari, J.K.; Ballabha, R. and Tiwari, P. (2010). Ethnopaediatrics in Garhwal Himalaya, Uttarakhand, India. (*Psychomedicine and Medicine*). *New York Science Journal*, 3:123-126.
- Tonsomboon, A.; Prasanth, M.L.; Plaingam, W. and Tencomnao, T. (2021). *Kaempferia parviflora* rhizome extract Inhibits Glutamate-Induced toxicity in HT-22 Mouse Hippocampal neuronal cells and extends longevity in *Caenorhabditis elegans*. *Biology (Basel)*, 10(4):264.
- Tri, M.D.; Phat, N.T.; Tho, P.Q.; Khiem, T.H.; Dang, S. and Dan, N.T.N. (2021). Chemical constituents of the rhizomes of *Kaempferia parviflora*. *Can Tho University Journal of Science*, 57(1). DOI:10.22144/ctu.jvn.2021.007.
- Wang, Y.; Zhang, Y.J.; Gao, W.Y. and Yan, L.L. (2007). Anti-tumor constituents from *Paris polyphylla* var. yunnanensis. *China journal of Chinese materia medica*, 32(4):1425-1428.
- Warrier, R.R. (2021). Authentication of herbal products to attract global markets. *Ann. Phytomed.*, 10(2):1-3.
- Wattanathorn, J.; Muchimapura, S.; Tong-Un, T.; Saenghong, N.; Thukhum-Mee, W. and Sripanidkulchai, B. (2012). Positive modulation effect of 8-week consumption of *Kaempferia parviflora* on health related physical fitness and oxidative status in healthy elderly volunteers. *Evidence Based Complementary and Alternative Medicine*, 132: 1150-1155.
- Wen, F.; Yin, H.; Chen, C.; Liu, X. Xue, D.; Chen, T.; He, J. and Zhang, H. (2012). Chemical characteristics of saponins from *Paris fargesii* var. brevipetala and cytotoxic activity of its main ingredient, Paris saponin H. *Fitoterapia*, 83(4):627-635.
- Wutythamawech, W.E (1997). *Encyclopedia of Thai Herbs*. OS Printing; Bangkok, Thailand. pp:365.
- Yan, L.L.; Zhang, Y.J.; Gao, W.Y.; Man, S.L. and Wang, Y. (2009). In vitro and in vivo anticancer activity of steroid saponins of *Paris polyphylla* var. yunnanensis. *Experimental Oncology*, 31(1):27-32.

Yenjai, C.; Prasanphen, K.; Daodee, S.; Wongpanich, V. and Kittakoop, P. (2004). Bioactive flavonoids from *Kaempferia parviflora*. *Fitoterapia*, **75**(1):89-92.

Yun, H.; Lijian, C.; Wenhong, Z.; Yuhong, D.; Yongli, W.; Qiang, W. and Ding, Z. (2007). Separation and identification of steroidal compounds with cytotoxic activity against human gastric cancer cell lines *in vitro* from the rhizomes of *Paris polyphylla* var. *chinensis*. *Chemistry of Natural Compounds*, **43**(6):672-677.

Zhang, M.; Li, Y.W.; Li, Z.Y.; Huang, X.L.; Zhu; Liu, Q.S. (2011). Progress on studies of endangered ethno-medicine of *Rhizoma Paris*. *Journal of Central South University*, **20**:65-69.

Zhang, X.; Cui, Y.; Huang, J.; Zhang, Y.; Nie, Z.; Wang, L.; Yan, B.; Tang, Y. and Liu, Y. (2007). Immuno stimulating properties of diosgenyl saponins isolated from *Paris polyphylla*. *Bioorganic and Medicinal Chemistry Letters*, **17**(9):2408-2413.

Citation

A.K. Phurailatpam, Anju Choudhury, Tasso Yatung and Kalkame Ch. Momin (2022). A review on the importance of two medicinal plants of North East India: *Paris polyphylla* Smith and *Kaempferia parviflora* Wall. ex Baker. *Ann. Phytomed.*, **11**(2):214-223. <http://dx.doi.org/10.54085/ap.2022.11.2.23>.